

## 1 **2. Methods**

### 2 **2.3 CIMS**

3 Experiments utilizing the bisulfate/sulfate buffer (IO3-5, IO8, SW3-5, SW8, and CL1) sometimes  
4 exhibited cyclical CIMS signal changes for Br<sub>2</sub> (*m/z* 285, 287, 291), IBr (*m/z* 333, 335) with no attributable  
5 cause. These signal changes occurred seemingly at random and to varying extents. In Fig. S1a, Experiment  
6 IO4 (pH = 1.7, includes H<sub>2</sub>O<sub>2</sub>) demonstrates the most extreme example of this behaviour that almost appears  
7 to affect the analysis. First at t = -3, the Br<sub>2</sub> rises briefly before falling. Then at t=2, the Br<sub>2</sub> signal begins  
8 to resemble a sine wave. All data beyond t=2 is not considered for this specific experiment. In Fig S1b, the  
9 effect during Experiment SW5 (pH = 1.7, includes H<sub>2</sub>O<sub>2</sub>) is more muted, beginning at approximately t = -  
10 6 for IBr and Br<sub>2</sub>. As represented by these figures, this behaviour being farther away from our periods of  
11 integration is typical of the remaining experiments. Because these signal changes occurred outside of the  
12 experimental periods analyzed (i.e., before irradiation, and after O<sub>3</sub> had been active for one hour), they are  
13 therefore not believed to affect our results and their interpretation.

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## 15 **3. Results**

### 16 **3.1 Dark reaction production of I<sub>2</sub>**

17 In cases without OH precursors at pH < 2, significant photochemical I<sub>2</sub> production still occurs  
18 (integrated production of 14 ± 10 nmol for IO8, and 6.0 ± 2.0 nmol for SW8), while Br<sub>2</sub> and Cl<sub>2</sub>  
19 concentrations remain below limits of detection (consistent with Abbatt et al., (2010), in which no Br<sub>2</sub> was  
20 observed without an OH-precursor) (Table 2, main text). This production likely stems from the mechanisms  
21 outlined by Kim et al. (2016) (R13-14, R10-R12), discussed in the Sect. 1. As discussed in Sect. 3.1, H<sub>2</sub>O<sub>2</sub>  
22 or NO<sub>2</sub><sup>-</sup> can react directly with I<sup>-</sup>, thereby reducing the available [I<sup>-</sup>] for photochemical OH oxidation when  
23 pH < 2. When H<sub>2</sub>O<sub>2</sub> was the oxidant, integrated I<sub>2</sub> production amounts were found to be ≤ 0.82 nmol (IO4,  
24 IO5, and SW5), likely due to this initial dark depletion. When instead NO<sub>2</sub><sup>-</sup> is used (as in IO3 and SW3),

25 initial amounts of I<sub>2</sub> on flowtube connection to CIMS were less than when H<sub>2</sub>O<sub>2</sub> was used (Table S1, Fig.  
26 S3). To estimate how much I<sup>-</sup> may have been lost from our frozen sample by these dark mechanisms, we  
27 convert the integrated I<sub>2</sub> production amounts from Table S1 to I<sup>-</sup> (by multiplying by 2) and subtract from  
28 the maximum possible moles of I<sup>-</sup> in our samples (0.0800 L \* 1.6 x 10<sup>-6</sup> M = 1.28 x 10<sup>-7</sup> moles I<sup>-</sup>). For the  
29 samples that use hydrogen peroxide, as little as 36–91% of I<sup>-</sup> is available for reaction, while 94-97% remain  
30 when using NO<sub>2</sub><sup>-</sup>. However, it is certain that not all of the I<sub>2</sub> produced by this mechanism went into the  
31 CIMS by the nature of having to break the flow tube seal in order to connect it to the CIMS, and so these  
32 are only estimates that could be affected by the length of time the tube is open to the environment and not  
33 connected to the CIMS, or sealed shut.

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### 35 **3.2 Hydroxyl radical-induced halogen production at pH ≈ 4.7**

36 Considering the values of I<sub>2</sub> production from Table 2 (main text), IO<sub>2</sub>, appears to have produced  
37 ~10 times less I<sub>2</sub> based on the chosen period of integration. It was noted that I<sub>2</sub> appeared to already be  
38 present within the flow tube on connecting the flow tube to the CIMS (Fig. S2). This production could  
39 possibly be induced by room light during the transfer of the flow tube from the cooling box to the  
40 experimental light box. However, this is believed to be minimal relative to the production from the solar  
41 simulator bulbs in the photolysis box; the relative absorption spectrum for hydrogen peroxide ceases  
42 significant absorption above 400 nm (Phibbs and Giguère, 1951), while the emission spectrum of typical  
43 fluorescent lights begins significant emission above 400 nm. However, the experiment otherwise  
44 eventually produces the same qualitative features as the other three experiments after light activation (Fig.  
45 S2). If instead the limits of integration are chosen starting when the I<sub>2</sub> signal begins rising (i.e., during a  
46 period that qualitatively resembles the other experiments), the integrated I<sub>2</sub> production amounts (1.1 ± 0.6  
47 nmol) more closely resemble the aforementioned experiments. In these conditions and in the time span of  
48 the experiment, relatively little Br<sub>2</sub> was produced (maximum of 34 ± 2 pmol, experiment IO2), and Cl<sub>2</sub>  
49 concentrations remained below limits of detection.

### 50 3.4 Effects of O<sub>3</sub> on halogen production

51 As discussed in the main text, HOX compounds were observed when O<sub>3</sub> was added to the flow  
52 tube. With regard to the extent to which it affects our observed signal, we believe volatile organic  
53 compounds, such as aldehydes and ketones, that may form gas phase HX could originate from our cylinder  
54 of zero air. However, we believe this source would be effectively scrubbed by our activated charcoal trap  
55 (Fig. 1), mitigating any gas phase production of HX. There also exists organic matter in the condensed  
56 phase, averaging 70 mg/L in each Instant Ocean sample (Sect. 2 of the main text). This carbon-matter is  
57 presumably uncharged and would freeze throughout the formed ice (i.e., no freeze concentration effect),  
58 therefore making only a small fraction of the total carbon available at the frozen surface for reaction.

59 If any of this solution-based carbon were involved in making HX, it would be expected that the SW  
60 and IO experiments produce different amounts of IOHX<sup>-</sup>, given that the SW experiments were found to  
61 average ~5 mg/L of dissolved organic matter. However, there is no difference in the signal changes between  
62 corresponding SW and IO experiments (Figs. 3-4, S3). Therefore, we believe the primary source of IOHX<sup>-</sup>  
63 in the CIMS is, indeed, HOX formed in the flow tube.

## 64 4. Discussion

65 Through flow tube experiments involving condensed phase hydroxyl radical production, I<sup>-</sup>, Br<sup>-</sup>, and  
66 Cl<sup>-</sup> were oxidized to produce I<sub>2</sub>, Br<sub>2</sub>, and Cl<sub>2</sub>, respectively, from frozen saline surfaces. At low pH (~2),  
67 and with H<sub>2</sub>O<sub>2</sub> as our OH precursor, we noted a large outflux of I<sub>2</sub> on connecting the flow tube to the CIMS.  
68 Br<sub>2</sub> production was readily observed in the presence of light, and enhanced when the samples were exposed  
69 to O<sub>3</sub>, as in Fig 2b. However, experiment SW3 (Fig. S4), which was performed with NO<sub>2</sub><sup>-</sup> as the hydroxyl  
70 radical precursor, exhibited photochemical I<sub>2</sub> production on the introduction of radiation. Only after the  
71 introduction of O<sub>3</sub> was Br<sub>2</sub> observed (under proper isotope ratios).

72 In most cases, it was also found that extending limits of integration beyond 1 h after addition of O<sub>3</sub>  
73 did not produce I<sub>2</sub> in amounts that exhausted the supply of I<sup>-</sup>. In an example experiment (IO2, Fig. S5), the  
74 limits of integration were extended to t = 15 hours after the initiation of lights. While the signal appeared

75 to stabilize below the I<sub>2</sub> LOD of 9 pmol mol<sup>-1</sup>, the calculated I<sub>2</sub> production amount of 70 nmol for this  
76 extended integration period only accounts for 46% of the 152 total nmol of I<sup>-</sup> available. When repeated for  
77 the other experiments at pH = 4.7, it is found that at least 16% of the original I<sup>-</sup> remains unreacted after  
78 similarly extended limits of integration. This suggests that all of the I<sup>-</sup> in our frozen samples may not be  
79 completely excluded to the disordered interface, and may exist within the ice bulk or inaccessible brine  
80 channels throughout the ice, and that differences in integration production amounts can originate from  
81 differences in I<sup>-</sup> distribution during freezing (Bartels-Rausch et al., 2014; Malley et al., 2018).

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83 **References**

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113 **Tables**

114 Table S1: Integrated I<sub>2</sub> production amounts prior to irradiation or addition of O<sub>3</sub> from low pH experiments  
115 involving samples with an OH precursor. The period of integration was chosen to be immediately after  
116 connection of flow tube to the CIMS until sample was irradiated. Average LODs for I<sub>2</sub> across experiments  
117 was  $9 \pm 2$  pmol mol<sup>-1</sup>. “IO#” represents samples composed of Instant Ocean, and “SW#” represents  
118 “saltwater” samples, composed of reagent salts.

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Experiment	Oxidant	pH	I <sub>2</sub> produced (nmol)	Integration time (hours)	Estimated Percent of I <sup>-</sup> remaining for reaction
IO3	NO <sub>2</sub> <sup>-</sup>	2.0	4.0(±0.1)	0.55	93.7
SW4	NO <sub>2</sub> <sup>-</sup>	2.2	2.5(±0.1)	0.43	96.1
SW3	NO <sub>2</sub> <sup>-</sup>	1.8	2.0(±0.1)	0.83	96.8
IO4	H <sub>2</sub> O <sub>2</sub>	1.7	41(±14)	7.28	36.2
IO5	H <sub>2</sub> O <sub>2</sub>	1.7	5.7(±1.9)	2.92	91.1
SW5	H <sub>2</sub> O <sub>2</sub>	1.8	41(±14)	4.95	35.5

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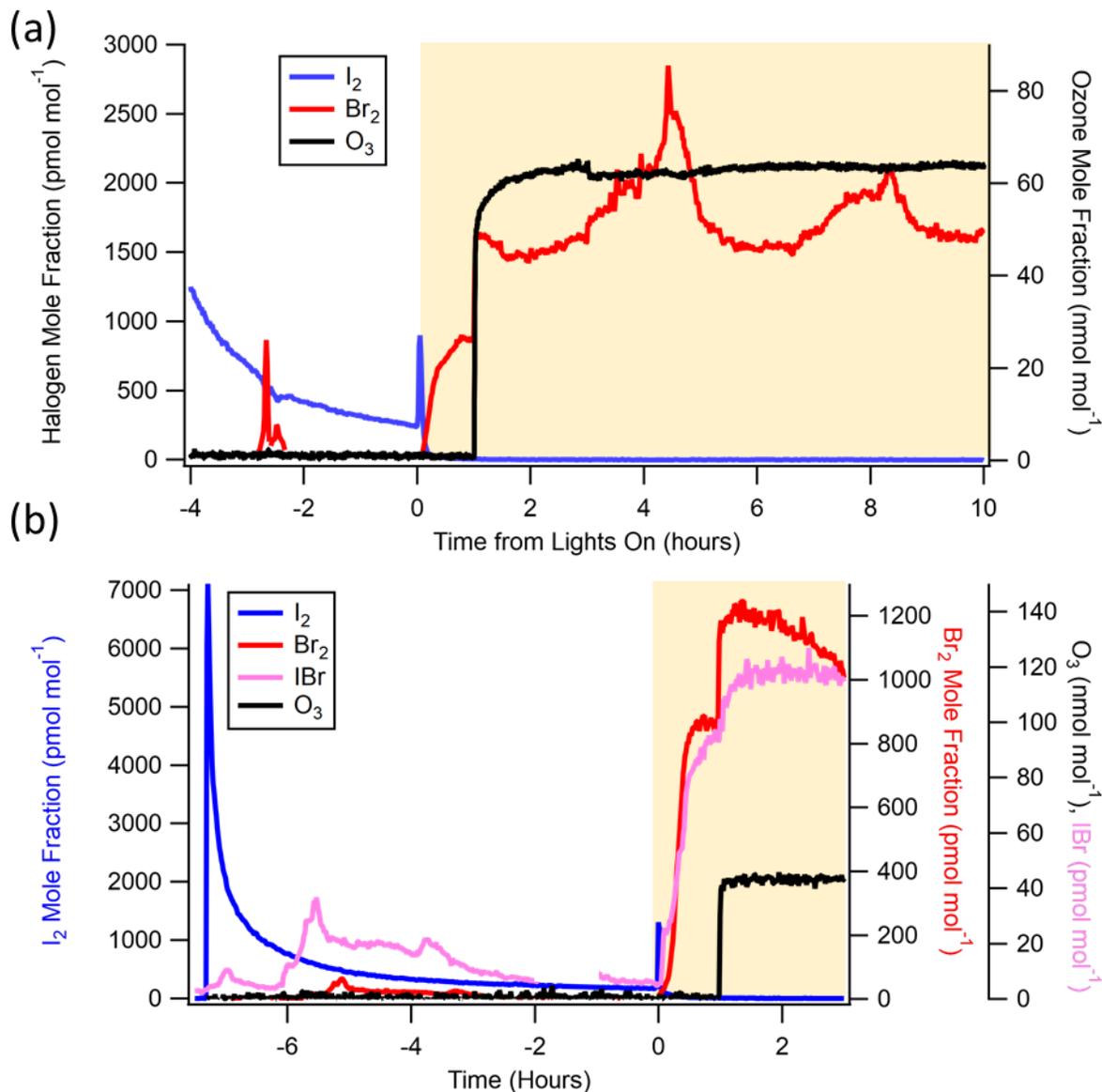
Table S2: Integrated I<sub>2</sub> produced from pH = 4.7 experiments involving samples with an OH precursor. The period of integration begins at sample irradiation and extends past the limits of analysis used in the main text. Average LODs for I<sub>2</sub> across experiments was  $9 \pm 2$  pmol mol<sup>-1</sup>. “IO#” represents samples composed of Instant Ocean, and “SW#” represents “saltwater” samples, composed of reagent salts.

Experiment	Oxidant	pH	I <sub>2</sub> produced (nmol)	Integration time (hours)	Estimated Percent of I <sup>-</sup> remaining for reaction
IO1	H <sub>2</sub> O <sub>2</sub>	4.7	31(±10)	30	59
IO2	H <sub>2</sub> O <sub>2</sub>	4.7	35(±20)	15	54
SW1	H <sub>2</sub> O <sub>2</sub>	4.7	63(±23)	23	17
SW2	H <sub>2</sub> O <sub>2</sub>	4.5	63(±20)	17	16

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132 **Figures**



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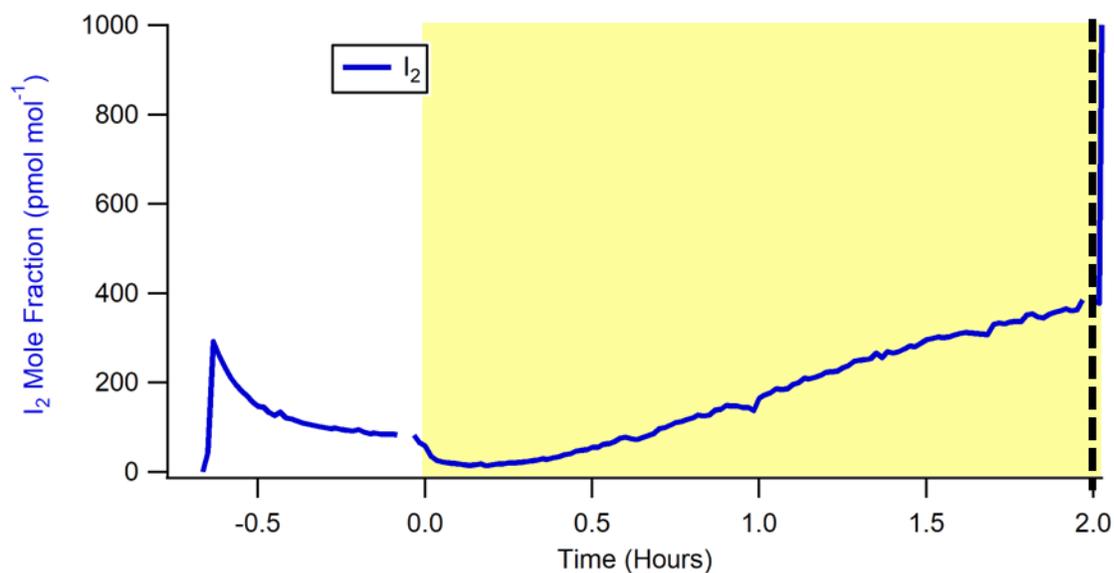
134 Figure S1: a) Experiment IO4 (pH < 2, includes H<sub>2</sub>O<sub>2</sub>) time series demonstrating cyclical increases in signal  
135 Br<sub>2</sub> signals, especially at t = -3 and beginning again at t = 2. Period of analysis in main text includes t = 0  
136 until t = 2. b) Experiment SW5 (pH < 2, includes H<sub>2</sub>O<sub>2</sub>) time series demonstrating cyclical signals for IBr  
137 and Br<sub>2</sub>, beginning predominately at t = -6 until shortly before t = 0.

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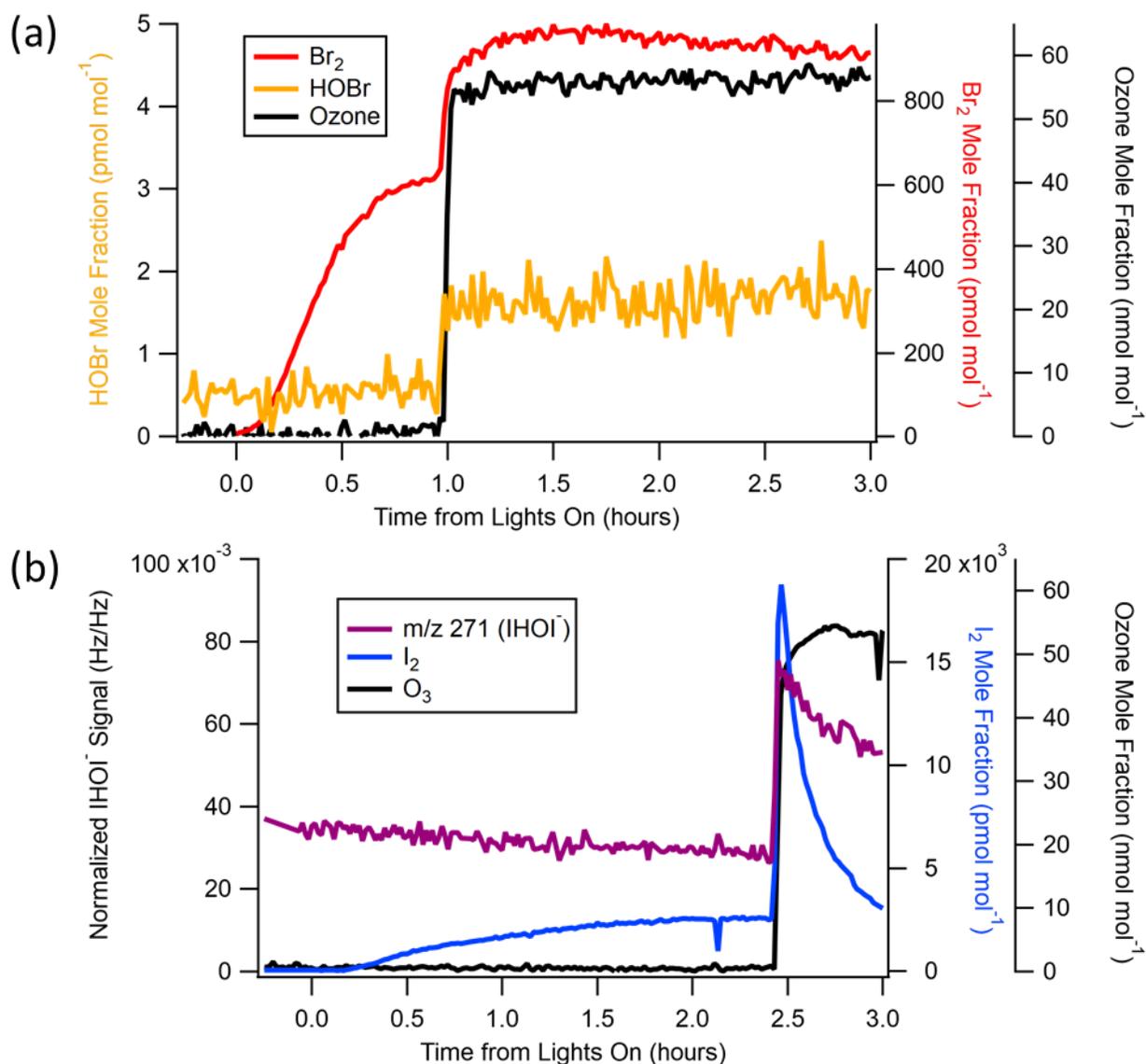


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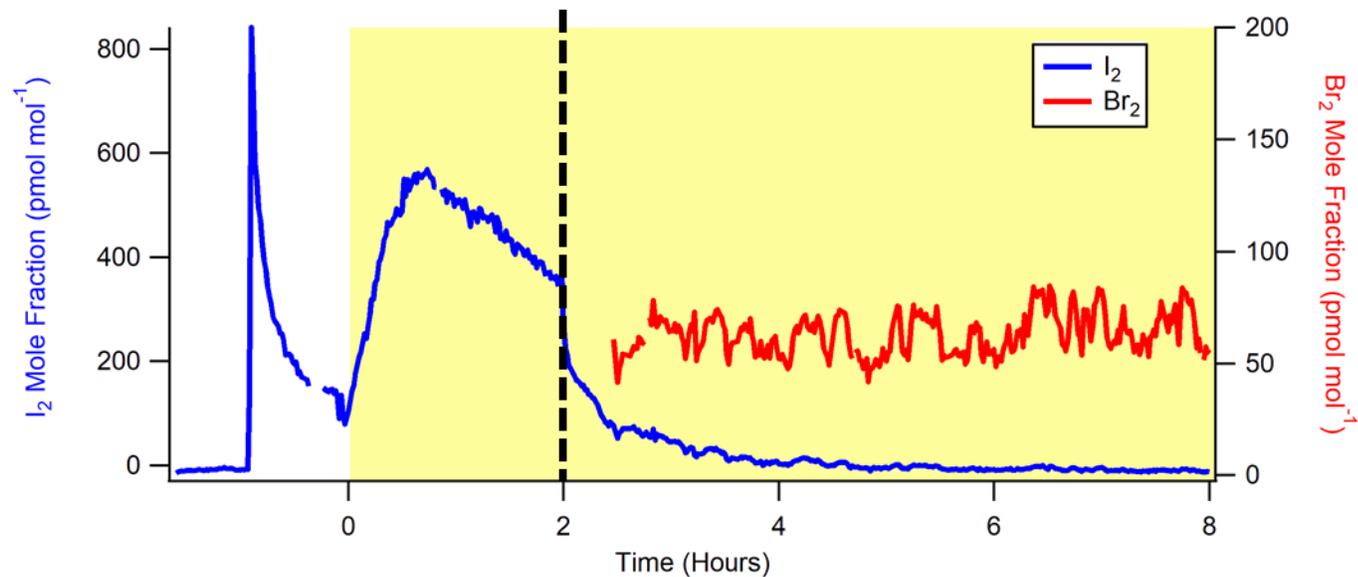
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144 Figure S2: Experimental timeseries for experiment IO2. The key difference between this experiment and  
145 others at pH ~4.68 is that there was some initial I<sub>2</sub> present when the flow tube was connected to the CIMS.  
146 On activating the lights, these concentrations lowered, before ultimately rising due to OH-induced I<sub>2</sub>  
147 production. Beginning the integration when the signal begins rising leads to similar production values as  
148 those experiments without this initial I<sub>2</sub> present. Vertical dashed line represents when O<sub>3</sub> was introduced  
149 to the system.

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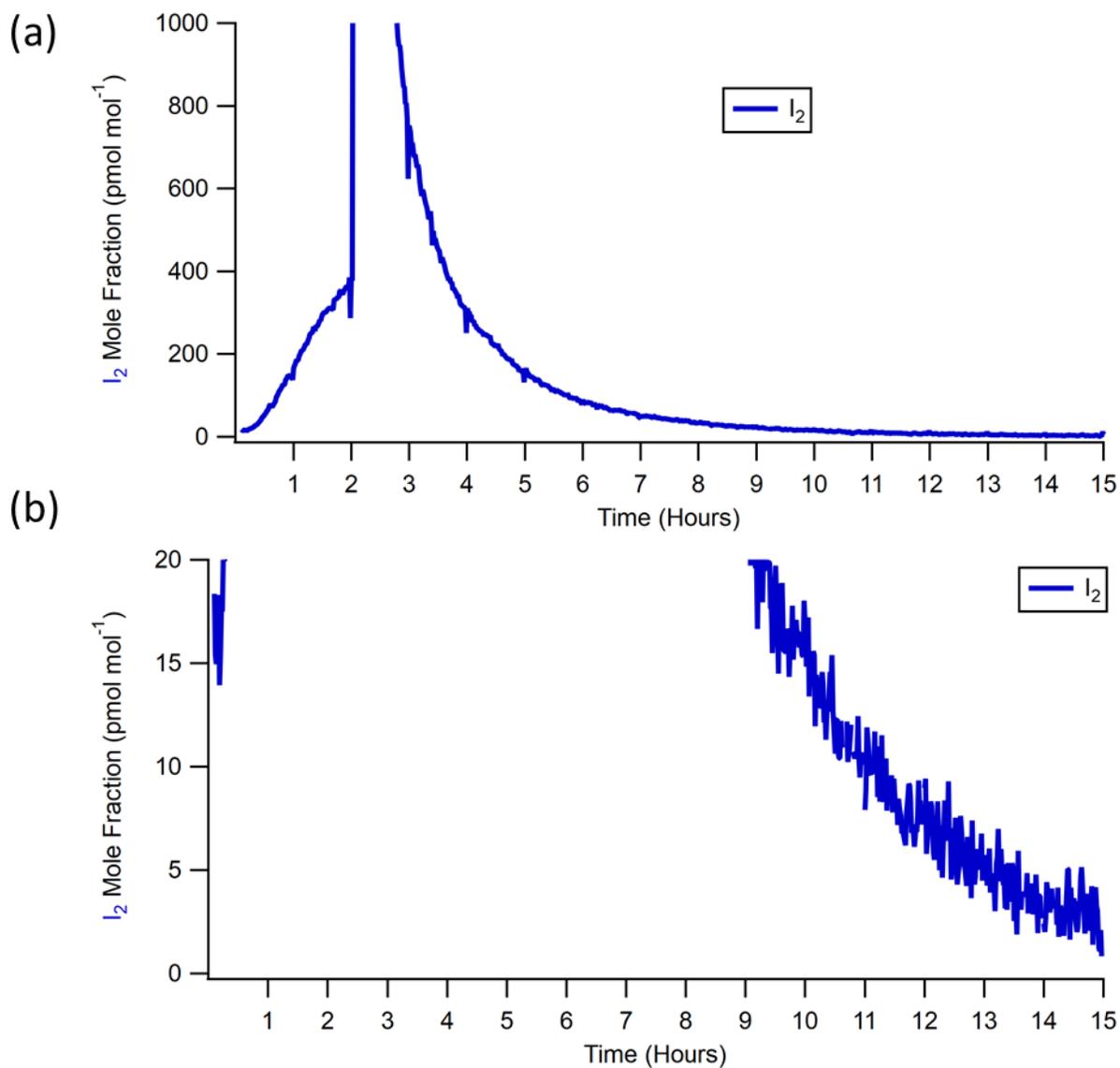
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 152 Figure S3: a) Experiment IO5, using Instant Ocean at pH = 1.7, in which  $\text{H}_2\text{O}_2$  acted as our hydroxyl radical  
 153 precursor (analogous to SW5, Fig. 4). Comparison of  $\text{Br}_2$  mole fractions to HOBr. Note that the HOBr  
 154 signal, while calibrated, should be used only for qualitative purposes as its identity could not be confirmed  
 155 using isotopic ratios with  $m/z$  223 due to its relatively large background signal.  $\text{Br}_2$  data filtered based on  
 156 correctness of isotope ratios between  $m/z$  285 and 287 (IBrBr<sup>-</sup>). b) Experiment SW2 (analogous to IO2, Fig.  
 157 3) showing effect of  $\text{O}_3$  on  $\text{I}_2$  and HOI.  
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161 Figure S4: Experiment SW3, using synthetic seawater at pH = 1.8, in which NO<sub>2</sub><sup>-</sup> acted as our hydroxyl  
 162 radical precursor. Ozone was introduced at hour two (indicated by dashed vertical line), coincident with  
 163 the I<sub>2</sub> concentration decrease. Br<sub>2</sub> data filtered based on correctness of isotope ratios between *m/z* 285 and  
 164 287 (IBrBr).

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169 Figure S5: Iodine time series from experiment IO2, using Instant Ocean at pH = 4.7, in which H<sub>2</sub>O<sub>2</sub> acted  
 170 as our hydroxyl radical precursor. The x-axis begins on light introduction to the flow tube, while ozone  
 171 was introduced at hour two as indicated by the sudden increase in signal. (a) The time series signal rapidly  
 172 increases at t=2 coincident with the addition of 60 nmol mol<sup>-1</sup> of O<sub>3</sub>, and then returns to baseline by hour  
 173 13. (b) Zoomed in version of the same plot