Effect of mid-term drought on *Quercus pubescens* BVOCs emissions seasonality and their dependence to light and/or temperature

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Abstract. Biogenic volatile organic compounds (BVOCs) emitted by plants represent a large source of carbon compounds released into the atmosphere where they account for precursors of tropospheric ozone and secondary organic aerosols. Being directly involved in air pollution and indirectly in climate change, understanding what factors drive BVOC emissions is a prerequisite for modelling their emissions and predict air pollution. The main algorithms currently used to model BVOCs emissions are mainly light and/or temperature dependent. Additional factors such as seasonality and drought also influence isoprene emissions, especially in the Mediterranean region which is characterized by a rather long drought period in summer. These factors are increasingly included in models but only for the principal studied BVOC, namely isoprene but there are still some discrepancies in estimations of emissions. In this study, the main BVOCs emitted by *Quercus pubescens*: isoprene, methanol, acetone, acetaldehyde, formaldehyde, MACR, MVK and ISOPOOH (these 3 last compounds detected under the same ion), were monitored with a PTR-ToF-MS over an entire seasonal cycle, under both *in situ* natural and amplified drought which is expected with climate change. Amplified drought impacted all studied BVOCs by reducing emissions in spring and summer while increasing emissions in autumn. All six BVOCs monitored showed daytime light and temperature dependencies while three BVOCs (methanol, acetone and formaldehyde) also showed emissions during the night despite the absence of light under constant temperature. Moreover, methanol and acetaldehyde burst in the early morning and formaldehyde deposition/uptake were also punctually observed which were not assessed by the classical temperature and light models.

1 Introduction
Plants contribute to global emissions of volatile organic compounds (VOCs) with an estimated emission rate of $10^{15}$ gC yr$^{-1}$ (Guenther et al. 1995; Harrison et al. 2013). The large variety of compounds released by plants represents, at the global scale, 2-3% of the total carbon released in the atmosphere (Kesselmeier & Staudt 1999).

Under strong photochemical conditions, BVOCs, together with NOx, can significantly contribute to tropospheric ozone concentration (Xie et al. 2008; Papiez et al. 2009). In addition to its greenhouse effect, O$_3$ has strong effects on plant metabolism (Reig-Armiñana et al. 2004; Beauchamp et al. 2005) as well as on human health (Lippmann 1989). BVOCs are also rapidly oxidized by OH radical and NO$_3$ (Hallquist et al. 2009; Liu et al. 2012), which account for an important fraction of the total mass of secondary organic aerosols (SOA, Jimenez et al. 2009).

Methanol and acetone are, after isoprene, the principal BVOC released to the atmosphere. Isoprene emissions represent between 400-600 TgC yr$^{-1}$ at the global scale (Arnett et al. 2008) whereas methanol emissions vary between 75 and 280 TgC yr$^{-1}$ (Singh et al. 2000; Heikes et al. 2002, respectively) and acetone emissions represent only 33 TgC yr$^{-1}$ (Jacob et al. 2002). Other compounds such as acetaldehyde, methacrolein (MACR), methyl vinyl ketone (MVK), isoprene hydroxy hydroperoxides (ISOPOOH) and formaldehyde, whose biogenic origin has been poorly investigated, are better known to be anthropogenic and/or secondary VOCs issued from atmospheric oxidations (Hallquist et al. 2009). However, acetaldehyde is also a by-product of plant metabolism and its emissions represent 23 Tg yr$^{-1}$ at the global scale (Millet et al. 2010). Formaldehyde, MACR, MVK and ISOPOOH are released by plants through oxidations of methanol and isoprene, respectively, within leaves but they can have other leaf precursors (Oikawa & Lerdau 2013). Thus, it is thereby important to model all this panel of BVOCs emissions with the aim of predicting their effect on secondary atmospheric chemistry.

Current models allow to predict BVOCs emissions according to the type of vegetation, biomass density, leaf age, specific emission factor for many vegetal species, as well as the impact of some environmental factors. Models, such as the MEGAN (Guenther et al. 2006; Guenther et al. 2012) or CHIMERE (Menut et al. 2014) model, include at least two main algorithms that allow to model light and temperature emissions dependence (called $L+T$ algorithm afterwards) and a temperature dependent algorithm (called $T$ algorithm afterwards), both described in Guenther et al. (1995). The $L+T$ algorithm is typically used for BVOCs emissions whose synthesis rapidly relies on photosynthesis, and hence include de novo emissions. The $T$ algorithm is used for BVOCs emissions that do not directly rely on BVOCs synthesis when, for example, they originate from permanent large storage pools (Ormeno et al. 2011). The dependence to light and/or temperature is well documented for isoprenoids (Owen et al. 2002; Rinne et al. 2002; Dindorf et al. 2006) but there is still a lack of knowledge about highly volatile BVOCs (e.g. methanol, acetone, acetaldehyde). However, many of these compounds are very reactive in the atmosphere (Hallquist et al. 2009) and, could be emitted in large quantities to the atmosphere at global scale. The characterization of their emissions and sensitivity to light and/or temperature is, thus, necessary in order to obtain reliable predictions of atmospheric processes in order not to miss this important part of the atmospheric reactivity.

Other factors than light and temperature can drive BVOCs emissions such as water stress. Most studies dealing with BVOCs response to water stress have, however, focused on terpene-like compounds and have been carried out after weeks of watering restriction or removal under controlled conditions (for a review, see studies cited in Peñuelas and Staudt 2010). Considerable uncertainty remains in our understanding of emission mechanisms since some works showed increases (Funk et al. 2004; Monson et al. 2007) or decreases of isoprene emissions (Brüggemann & Schnitzler 2002; Fortunati et al. 2008) and there is a lack of knowledge on the impact of water stress on highly BVOCs emissions. Moreover, the understanding of isoprene sensitivity and highly volatile BVOCs...
to recurrent water stress (few years) under in situ conditions is clearly missing. Likewise, the capacity of current
L+T and T algorithms to predict emission shifts under different drought scenarios in the context of climate change
needs to be addressed for isoprene and highly volatile compounds. This is of especial interest for the Mediterranean
area where the most severe climatic scenario of the IPCC predicts an intensification of summer drought consisting
on a rain reduction that can locally reach 30%, an extension of the drought period as well as a temperature rise of
3.4°C, (Giorgi & Lionello 2008; IPCC 2013; Polade et al. 2014) for 2100.
In the present investigation, we aimed (i) to study the emission factors of each studied BVOC released by Q.
pubescens, including isoprene and highly volatile compounds that originate from plant metabolism under water
stress (ii) to test the performance of the L+T and T algorithms to predict isoprene and highly volatile BVOC
emissions over the seasonal cycle and under two recurrent water stress treatments. Q. pubescens was chosen as
vegetal model because this species is highly resistant to drought and well widespread in the Northern
Mediterranean area occupying 2 million ha (Quézel & Médail 2003). It also represents the major source of isoprene
emissions in the Mediterranean area and the second one at the European scale (Keenan et al. 2009).

2 Material and methods

2.1 Experimental site

Our study was performed at the O3HP site (Oak Observatory at OHP, Observatoire de Haute Provence), located
60 km North of Marseille, France (5°42'44" E, 43°55'54" N), at an elevation of 650m above the sea level. The
O3HP (955m²), free from direct human disturbance for 70 years, is a homogeneous forest mainly composed of Q.
pubescens (≈ 90 % of the biomass and ≈ 75 % of the trees) with a mean diameter of 1.3 m. The remaining 10 %
of the biomass is mostly represented by Acer monspessulanum trees, a very low isoprene-emitter species (Genard-
Zielinski et al. 2015). The O3HP site was created in 2009 in order to study the impact of climate change on a Q.
pubescens forest. Using a rainfall exclusion device (an automated monitored roof deployed during chosen rain
events) set up over part of the O3HP canopy, it was possible to reduce natural rain by 30% and to extend the
drought period in an attempt to mimic the current climatic model projections for 2100 (Giorgi & Lionello 2008;
IPCC 2013; Polade et al. 2014). Two plots were considered in the site: a plot receiving natural precipitation where
trees grew under natural drought (300m² surface, used as control plot) and a second plot submitted to amplified
drought (232m² surface). Rain exclusion on this latter plot started on May 2012 and was continuously applied
every year, principally, during the growth period. Ombrothermic diagrams indicate that the drought period was
extended for 2 months in 2012, 4 months in 2013 and 3 months in 2014 for amplified drought relative to natural
drought (Fig 1). Data on cumulative precipitation show that 35% of rain was excluded in 2012 (from 29 April from
to 27 October), 33.5% in 2013 (from 7 July from to 29 December), 35.5% in 2014 (from 8 April to 8 December).
This experimental set up involved a recurrent drought in the amplified drought plot. Sampling was performed at
the branch-scale at the top of the canopy during three campaigns from October 2013 to July 2014, covering an
entire seasonal cycle: in autumn (14 to 28 October 2013, 2nd year of amplified drought), in spring (12 to 19 May
2014, 3rd year of amplified drought) and in summer (13 to 25 July 2014, 3rd year of amplified drought). Spring,
summer and autumn campaigns corresponded to the end of leaf growth, leaf maturation and the beginning of the
leaf senescence, respectively. The same five trees per plot were selected and investigated throughout the study.
2.2 Branch scale-sampling methods

Two identical dynamic branch enclosures were used for sampling gas exchange (in terms of CO₂, H₂O and BVOCs) as fully described in Genard-Zielinski et al. (2015) with some modifications. Branches were enclosed in a = 30L PTFE (polytetrafluoroethylene) frame closed by a 50µm thick PTFE film. One tree from natural and one tree from amplified drought plot were analysed concomitantly during 1 or 2 days. Inlet air was introduced at 9L.min⁻¹, controlled by mass flow controllers (MFC, Bronkhorst), using a pump, inside, by PTFE (KNF N840.1.2FT.18®, Germany) allowing for air renewal inside the chamber every ~ 3min. Ozone was removed from inlet air by placing PTFE filters impregnated with sodium thiosulfate (Na₂S₂O₃) as described by Pollmann et al. (2005), so that oxidation of BVOCs due to ozone within the enclosed atmosphere is negligible. The excess of air humidity was removed using drierite. A PTFE fan ensured a rapid mixing of the chamber air and a slight positive pressure within the enclosure enabled the PTFE film to be held away from leaves to minimise biomass damage.

Microclimate (temperature, relative humidity and photosynthetically active radiation or PAR) was continuously (every minute) monitored by a data logger (LI-COR 1400®; Lincoln, NE, USA) with a relative humidity and temperature probe placed inside the chamber (RHT probe, HMP60, Vaisala, Finland) and a quantum sensor (PAR, LI-COR, PAR-SA 190®, Lincoln, NE, USA) placed outside the chamber. The climatic conditions in terms of PAR and temperatures are summarized in Fig. S1 (in supplementary files) for each field campaigns. All air flow rates were controlled by mass flow controllers (MFC, Bronkhorst) and all tubing lines were made of PTFE. Chambers were installed the day before measurements and flushed overnight. Enclosed branches contained 8 to 12 leaves corresponding to a range of 1.4 to 3.6 g of dry matter and 110 to 320 cm² of leaf surface, respectively.

2.3 Ecophysiological parameters

Exchange of CO₂ and H₂O from the enclosed branches was continuously (every min) measured using infrared gas analysers (IRGA 840A®, LI-COR) concomitantly with BVOCs emission measurements (cf. 2.4). Gas exchange values were averaged by taking into account all the data measured between 12h and 15h (local time). Net photosynthesis ($P_n$, µmolCO₂ m⁻² s⁻¹) and stomatal conductance to water ($G_w$, mmolH₂O m⁻² s⁻¹) were calculated using equations described by Von Caemmerer and Farquhar (1981) as used in Genard-Zielinski et al. (2015) (for more details, see Appendix A, equations A1 to A4). Leaves from enclosed branches were directly collected after gas exchange sampling to accurately measure leaf surface with a leaf area meter. $P_n$ and $G_w$ were hence expressed in a leaf surface basis. After that, leaves were freeze-dried to assess their dry mass.

2.4 BVOCs analysis

A PTR-ToF-MS 8000 instrument (Ionicon Analytik GmbH, Innsbruck, Austria) was used for online measurements of BVOCs emitted by the enclosed branches. A multi-position common outlet flow path selector valve system (Vici) and a vacuum pump were used to sequentially select air samples from: amplified drought, inlet air, natural drought, ambient air and catalyst. The catalyst consists in a 25 cm long stainless steel tubing, filled with platinum wool and heated at 350°C to efficiently remove VOCs from sample and measure potential instrumental background levels. Each sample was analysed every hour, with 15min of analysis. Mass spectra in the range 0-500amu were recorded at 1min integration time. The reaction chamber pressure was fixed at 2.1mbar, the drift tube voltage at 550V and the drift tube temperature at 313 K corresponding to an electric field strength applied to the drift tube
(E) to a buffer gas density (N) ratio of 125Td (1Td = 10^{-17} \text{ V cm}^2). A calibration gas standard, consisting of a mixture of 14 aromatic organic compounds (TO-14A Aromatic Mix, Restek Corporation, Bellefonte, USA, 100 ± 10ppb in Nitrogen), was used to experimentally determine the ion relative transmission efficiency. BVOCs targeted in this study and their corresponding ions include formaldehyde (m/z 31.018), methanol (m/z 33.033), acetaldehyde (m/z 45.03), acetone (m/z 59.05), isoprene (m/z 41.038, 69.069) and MACR+MVK+ISOPOOH (m/z 71.049, these three compounds were detected with the same ion with PTR-MS). The signal corresponding to protonated VOCs was converted into mixing ratios by using the proton transfer rate constants k given by Cappellin et al. (2012). Formaldehyde concentrations were calculated according to the method described by Vlasenko et al. (2010) to account for its humidity dependent sensitivity. BVOCs emissions rates (ER) were calculated by considering the BVOCs concentrations in the inlet and outlet air as follows (equation 1):

\[ ER = \frac{Q_0 (C_{out} - C_{in})}{B} \] (1)

where \( ER \) was expressed in \( \mu \text{gC g}_{\text{DM}}^{-1} \text{ h}^{-1} \), \( Q_0 \) was the flow rate of the air introduced into the chamber (L h^{-1}), \( C_{out} \) and \( C_{in} \) were the concentrations in the inflowing and outflowing air (\( \mu \text{gC L}^{-1} \)), respectively, and \( B \) was the total dry biomass matter (g_{DM}). Daily cycles were made by averaging measured emissions of all trees every hour.

2.5 Emission algorithms

The light and/or temperature dependence of Q. pubescens BVOCs (isoprene and highly volatile compounds) under natural and amplified drought was tested using both the L+T and T algorithms. Emission rates calculated according to these algorithms (afterwards, called \( ER_{L+T} \) and \( ER_T \), respectively) were calculated using the equations described in Guenther et al. (1995) (for more details, see Appendix B, equations B1 to B5). The empirical coefficient \( \beta \) (used in the T algorithm) was determined for each compound according to the season and the treatment through the slope of correlation between the natural logarithm of emissions rates (measured emissions, \( \mu \text{gC g}_{\text{DM}}^{-1} \text{ h}^{-1} \)) and experimental temperature (K). Emissions factors (EF), that are emissions rates at standard conditions of light and temperature, 1000\( \mu \text{mol m}^{-2} \text{ s}^{-1} \) and 30°C, were used to calculate modelled emissions and were determined for each compound under each season and treatment. EF values correspond to the slope of the correlation between experimental emission rates and \( C_t^\ast C_l \) when using the L+T algorithm or \( C_T \) when using the T algorithm (without forcing data to pass through the origin, see Appendix B for a full description of \( C_t^\ast C_l \) and \( C_T \)). All parameters used for the calculation of modelled emissions are presented in supplementary files (Table S1 and S2).

2.6 Data treatment

Data treatment was performed with the software STATGRAPHICS® centurion XV (Statpoint, Inc). After having checked the normality of the data set, two-way repeated measures ANOVA were carried out to evaluate the variability of \( Pn, Gw \) and BVOC emission rates according to the drought treatment and season. Correlation coefficient (R²) and slope (called “sl” afterwards) from Pearson's correlations between measured and modelled emissions were used to evaluate the algorithm (L+T or T) that better predicted Q. pubescens emissions under the different drought conditions and seasonal cycle. The slope of those correlations indicate if there was an under- or over- estimation of modelled emissions when sl < 1 and sl > 1, respectively. For that, slope comparison tests were
performed to check for slope significant differences from 1. These correlations were obtained without forcing data to pass through the origin.

3. Results and discussion

3.1 Ecophysiological parameters

The physiology of *Q. pubescens* was slightly impacted by amplified drought over the whole study (Fig. 2), with a decrease of Gw under amplified drought compared to natural drought – ranging from 44 % in spring (P < 0.1) to 55 % in summer (P < 0.01, Table 1). In autumn, there was no significant difference between both treatments. Pn was only slightly reduced in summer by 36 % (P < 0.1) with no difference for the others season. Thus, the stomatal closure observed had a slight impact on carbon assimilation. Indeed, *Q. pubescens* has a high stem hydraulic efficiency (Nardini & Pitt 1999) which compensates stomatal closure since it allows to use water more efficiently, thus, maintaining Pn. Moreover, it must be noted that an increase of Pn was observed in autumn and could likely be attributed to autumnal rains. These results showed that the amplified drought artificially applied to *Q. pubescens* at O₃HP led to a moderate drought for this species, based on a moderate reduction of the physiological performances (Niinemets 2010).

3.2 Effect of drought on BVOCs emissions

Emissions of all BVOCs followed during this experimentation were reduced under amplified drought compared to natural drought, especially in spring and summer (Table 1) except for acetaldehyde emissions. Indeed, acetaldehyde was not significantly different between both treatments probably due to a large variability of the data set. In autumn, for all BVOCs, there was no difference between both plots. The decrease of oxygenated BVOCs in spring and summer under amplified drought (e.g. methanol, MACR+MVK+ISOPOOH, formaldehyde, acetone) could be explained by stomatal closure in spring and summer under amplified drought since emissions of these compounds are strongly bound to Gw (Niinemets et al. 2004). Isoprene emissions were also reduced in spring and summer during the 3rd year of this experiment whereas an increase had been observed in the first year (Génard-Zielinski et al. in prep) as well as what had been shown by Brüggemann and Schnitzler (2002) but this work was conducted with potted plants. The isoprene decrease observed in our experiment cannot be explained by the stomatal closure because this compound could also be emitted through the cuticle (Sharkey & Yeh 2001). It could rather be due to the decrease of Pn which reduced the carbon availability to produce isoprene. Moreover, carbon assimilated through Pn can be also invested into the synthesis of other defense compounds leading to a decrease of isoprene production and emission.

3.3 Effect of drought on light and/or temperature dependence through a seasonal cycle

All six BVOCs monitored showed daytime light and temperature dependencies (isoprene, degradation products of isoprene and acetaldehyde), while three BVOCs (methanol, acetone and formaldehyde) also showed emissions during the night despite the absence of light under constant temperature.
Regarding the light and temperature dependencies, the daily cycle of isoprene emissions (Fig. 3) showed that this compound clearly responds to light and temperature as already known (Guenther et al. 1993) and that this response is not impacted by amplified drought. Isoprene can protect thylakoids from oxidative damage (Velikova et al. 2011) occurring mainly during the day which can explain this kind of dependence. Yet, our results show the intensity of isoprene emission factor under natural and amplified drought is very different independently of the season. The modelled emissions were very representative of measured emissions except in spring under natural drought when we obtained a slight underestimation of emissions ($sl = 0.84$, $P < 0.05$) maybe, because light and temperature, in spring, were not the only parameters driving isoprene emissions. At this season, plants likely needed to produce more isoprene to protect the establishment of the photosynthetic machinery in the new leaves. MACR+MVK+ISOPOOH emissions, as isoprene, seemed to respond better to light and temperature than to only temperature (Fig. S2 in supplementary files) since correlations between measured emissions and $ER_{L,T}$ were always better than correlations with $ER_T$. Since MACR+MVK+ISOPOOH are oxidation products of isoprene (Oikawa & Lerdau 2013), it is not surprising that these compounds followed the same pattern than isoprene in terms of dependence to light and temperature. The estimations of $ER_{L,T}$ were quite good except in spring under natural drought where a slight underestimation was observed ($sl = 0.87$, $P < 0.05$).

The dependence of acetaldehyde emissions to light and/or temperature is very contrasted; studies have shown that they are bound to both light and temperature (Jardine 2008; Fares et al. 2011) or to temperature only (Hayward et al. 2004). Our results suggested that acetaldehyde emissions were mainly bound to light and temperature (Fig. 4). Indeed, correlations between measured and $ER_{L,T}$ were always better than with $ER_T$. However, some discrepancies were observed. Under natural drought, underestimations were observed in spring and summer ($sl = 0.72$, and $sl = 0.57$, $P < 0.05$, respectively) whereas in autumn, there was a good estimation ($sl = 0.86$, $P > 0.05$). Under amplified drought, underestimation was only observed in summer ($sl = 0.80$, $P < 0.05$). Daily cycles of acetaldehyde emissions presented also an emissions burst in the morning (at 7h, local time) in spring (under both treatments) and in summer (only under natural drought). Acetaldehyde can be produced due to an overflow of pyruvic acid during light-dark transitions. Cytosolic pyruvic acid levels rise rapidly and it can be converted into acetaldehyde by pyruvate decarboxylase (Fall 2003). This mechanism could explain the morning burst for this compound and the fact that no emissions during the night was observed.

We observed emissions of methanol, acetone and formaldehyde during the night under no light and constant temperature (around 20°C, see supplementary files S1). Correlations between $ER_{L,T}$ or $ER_T$ and measured methanol emissions were very similar especially in spring and summer (Fig. 5). However, some observed phenomena suggested that methanol emission was sustained by temperature in the absence of light. Indeed, the burst in the early morning (at 7h, local time), similar to acetaldehyde, was observed when stomata opened in spring and summer, independently of the drought treatment although it was clearer under natural than amplified drought. This burst can be explained by a strong release of this compound that has been accumulated in the intercellular air space and leaf liquid pools (due to the relative high polarity of methanol) at night when stomata are closed (Hüve et al. 2007). Moreover, for both drought treatments, methanol emissions during the night were observed at any seasons (especially autumn) which could be explained by nocturnal temperatures (roughly constant) that sufficed to maintain the biochemical processes involved in methanol formation. Methanol emissions, which result from the demethylation of pectin during the leaf elongation, has already been described to be temperature dependent alone
(Hayward et al. 2004; Folkers et al. 2008). However, our results suggest that methanol emissions respond strongly to light and temperature during the day. This kind of diurnal emissions cycle has already been described by Smiatek and Steinbrecher (2006). Our results about daily cycles of acetone emissions (Fig. S3 in supplementary files) showed that this compound responded better to light and temperature than only temperature since correlations were better with $ER_{L+T}$. Under natural drought, the modelled emissions were well representative of measured emissions in summer. By contrast, in spring and in autumn, slight underestimations were observed ($sl = 0.88, P < 0.05$ and $sl = 0.69, P < 0.05$, respectively). Under amplified drought, good estimations were observed in summer and autumn but in spring, there was an overestimation of modelled emissions ($sl = 1.27, P < 0.05$). Previous studies have shown that acetone rather depends on temperature alone (Fares et al. 2011) or to light and temperature (Jacob et al. 2002), indicating that its dependence on light and/or temperature remains unclear. During the day, acetone emissions were dependent on light and temperature and emissions still occurred during the night, especially in autumn. Alike methanol, nocturnal temperatures could allow to maintain acetone formation (Smiatek & Steinbrecher 2006). Acetone is a by-product of plant metabolism (Jacob et al. 2002) and its production can be enzymatic and non-enzymatic (Fall 2003) which can explain these observed differences through the day. We can suppose that acetone emissions observed during the day could come from the enzymatic activity and, on the contrary, during the night, they could come from the non-enzymatic production.

Formaldehyde emissions followed the same pattern than methanol and acetone emissions (Fig. S4 in supplementary files), especially in autumn. By considering only the daytime (correlation with $L+T$ modelled emissions), there were good estimations in summer and autumn and a slight underestimation was observed in spring ($sl = 0.89, P < 0.05$) for natural drought. Under amplified drought, correlations indicated that $L+T$ modelled emissions were well representative of measured emissions, but some negative emissions were observed in summer which suggested a deposition or an uptake of this compound by leaves as already highlighted by Seco et al. (2008).

This phenomenon could have a role in stress tolerance, since formaldehyde can be catabolised (mainly through oxidations) within leaves leading to CO$_2$ formation (Oikawa & Lerdau 2013). Emissions during the night suggest that formaldehyde came from another source than oxidation within leaves since oxidations occur mainly during the day due to an excess of light in chloroplasts, principal place of reactive oxygen species production (Asada 2006). Thus, formaldehyde emissions observed during the night could result from, for example, the glyoxylate decarboxylation or the dissociation of 5,10-methylene-THF (Oikawa & Lerdau 2013).

Predicting emissions rates of these 3 compounds (methanol, acetone and formaldehyde), during the night, seem to require other parameters such as a temperature threshold, below which methanol, acetone and formaldehyde synthesis and so emissions do not occur (Ghirardo et al. 2010).

### 4 Conclusion

After 3 years of amplified drought, all BVOC emissions were reduced in spring and summer compared to natural drought whereas, in autumn, an increase was observed for some compounds. These results are in opposition with the results obtained after only one year of amplified drought (2012), especially for isoprene, where an increase was observed for this compound (Génard-Zielinski et al. in prep). Amplified drought did not seem to shift the dependence to light and/or temperature which remained unchanged between treatments.
Moreover, two different dependence behaviours were found: (i) all six BVOCs monitored showed daytime light and temperature dependencies while (ii) only three BVOCs (methanol, acetone and formaldehyde) also showed that their emissions were maintained during the night with no light at rather constant nocturnal temperatures. Moreover, some phenomena, such as methanol and acetaldehyde emissions bursts in early morning or the formaldehyde deposition/uptake (formaldehyde), were not assessed by either L+T or T algorithm.

Appendix A: calculation of ecophysiological parameters

Net photosynthesis ($Pn$, $\mu$molCO$_2$ m$^{-2}$ s$^{-1}$) was calculated using equations described by Von Caemmerer and Farquhar (1981) as follows:

$$Pn = \frac{F*(Cr-Cs)}{S} - CS * E$$  \hspace{1cm} (A1)

Where $F$ is the inlet air flow (mol s$^{-1}$), $Cs$ and $Cr$ are the sample and reference CO$_2$ molar fraction respectively (ppm), $S$ is the leaf surface (m$^2$), $CS*E$ is the fraction of CO$_2$ diluted in water evapotranspiration and $E$ (molH$_2$O m$^{-2}$ s$^{-1}$) then transformed in mmolH$_2$O m$^{-2}$ s$^{-1}$, afterward) is the transpiration rate calculated as follow:

$$E = \frac{F*(Ws-Wr)}{S*(1-Ws)}$$  \hspace{1cm} (A2)

where $Ws$ and $Wr$ are the sample and the reference H$_2$O molar fraction respectively (molH$_2$O mol$^{-1}$).

Stomatal conductance to water ($Gw$, molH$_2$O m$^{-2}$ s$^{-1}$) was calculated using the following equation:

$$Gw = \frac{E*(1-Ws)}{Wl-Ws}$$  \hspace{1cm} (A3)

where $Wl$ is the molar concentration of water vapour within the leaf (molH$_2$O mol$^{-1}$) calculated as follows:

$$Wl = \frac{Vpsat}{P}$$  \hspace{1cm} (A4)

where $Vpsat$ is the saturated vapour pressure (kPa) and $P$ was the atmospheric pressure (kPa).

Appendix B: Modelled emissions calculation

The modelled emissions rates according to light and temperature ($ER_{L+T}$) or the temperature algorithm ($ER_T$) were calculated according to algorithms described in Guenther et al. (1995) as follows:

$$ER_{L+T} = EF_{L+T} * Cl * Ct$$  \hspace{1cm} (B1)

where $EF_{L+T}$ is the emission factor at 1000 $\mu$mol m$^{-2}$ s$^{-1}$ of photosynthetically active radiation (PAR) and 30°C of temperature (obtained with the slope of the correlation between experimental emissions and $Cl * Ct$ without forcing data to pass through the origin), $Cl$ and $Ct$ correspond to light and temperature dependence factors respectively and were calculated with the following formulae:

$$Cl = \frac{aCl1L}{\sqrt{1+a^2L}}$$  \hspace{1cm} (B2)

$$Ct = \frac{exp \frac{CT1(T-Ts)}{RT_sF}}{1+exp \frac{CT2(T-TM)}{RT_sF}}$$  \hspace{1cm} (B3)
where $\alpha = 0.0027$, $C_L = 1.066$, $C_{T1} = 95000 \text{ J mol}^{-1}$, $C_{T2} = 230000 \text{ J mol}^{-1}$, $T_M = 314 \text{ K}$ are empirically derived constants, $L$ is the photosynthetically active radiation (PAR) flux ($\mu\text{mol m}^{-2} \text{ s}^{-1}$), $T$ is the leaf experimental temperature (K) and $T_0$ is the leaf temperature at standard condition (303K). Modelled emissions according to temperature alone that is $ER_T$, was calculated as follows:

$$ER_T = EF_T * C_T$$

(B4)

where $EF_T$ is the emission factor at 30°C of temperature (obtained with the slope of the correlation between experimental emissions and $C_T$ without forcing data to pass through the origin) and $C_T$ is a temperature dependence factor calculated as follows:

$$C_T = \exp[\beta(T - T_S)]$$

(B5)

where $\beta$ is an empirical coefficient (with a standard variation value of 0.09 K$^{-1}$ used in literature when not measured) determined, in this study, for each compound according to the season and the treatment through the slope of the correlation between the natural logarithm of measured emissions rates ($ER, \mu\text{gC g}_{DM}^{-1} \text{ h}^{-1}$) and experimental temperature (expressed in K), $T$ is the leaf experimental temperature (K) and $T_S$ is the standard temperature (303K).

**Author contribution**

AS, EO and CF designed the research and the experimental design. AS, BTR, EO and CF conducted the research. AS, CB, BTR, and CL collected and analyzed the data. AS, EO, CB, HW, BTR, AA and CF wrote the manuscript

**Competing interests**

The authors declare that they have no conflict of interest.

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**References**


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Table 1: Net photosynthesis ($P_n$, µmolCO$_2$ m$^{-2}$ s$^{-1}$), stomatal conductance to water ($G_w$, mmolH$_2$O m$^{-2}$ s$^{-1}$) and emission rates (µgC g$_{DM}$$^{-1}$ h$^{-1}$) according to treatment and season. Values represent an average of all data measured between 12h and 15h (local time). Letters denote the difference between drought treatments with a > b and values showed represent the mean ± SE, n=5. ND: natural drought and AD: amplified drought with ns = non-significant, (*) = 0.05 < $P$ < 0.1, *= 0.01 < $P$ < 0.05, ** = 0.001 < $P$ < 0.01.

<table>
<thead>
<tr>
<th>Season</th>
<th>Spring</th>
<th></th>
<th></th>
<th></th>
<th>Summer</th>
<th></th>
<th></th>
<th>Autumn</th>
<th></th>
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<tbody>
<tr>
<td>Treatments</td>
<td>ND</td>
<td>AD</td>
<td>P</td>
<td>ND</td>
<td>AD</td>
<td>P</td>
<td>ND</td>
<td>AD</td>
<td>P</td>
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<tr>
<td>Pn</td>
<td>11 ± 1 a</td>
<td>9 ± 2 a</td>
<td>ns</td>
<td>14 ± 2 a</td>
<td>9±1.2 b</td>
<td>(※)</td>
<td>7 ± 1 a</td>
<td>9 ± 1 a</td>
<td>ns</td>
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<tr>
<td>Gw</td>
<td>110 ± 19 a</td>
<td>57 ± 13 b</td>
<td>(※)</td>
<td>285 ± 38 a</td>
<td>126 ± 17 b</td>
<td>**</td>
<td>122 ± 23 a</td>
<td>74 ± 21 a</td>
<td>ns</td>
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<tr>
<td>Isoprene</td>
<td>20 ± 4 a</td>
<td>10 ± 2 b</td>
<td>*</td>
<td>124 ± 10 a</td>
<td>81 ± 11 b</td>
<td>*</td>
<td>3 ± 1 a</td>
<td>5 ± 2 a</td>
<td>ns</td>
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<tr>
<td>MACR+MVK+ISOPOOH</td>
<td>0.1 ± 0.03a</td>
<td>0.1 ± 0.01 a</td>
<td>ns</td>
<td>0.4 ± 0.1 a</td>
<td>0.2 ± 0.02 b</td>
<td>*</td>
<td>0.04 ± 0.01 a</td>
<td>0.1 ± 0.01 a</td>
<td>ns</td>
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<tr>
<td>Methanol</td>
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<td>0.5 ±0.04 b</td>
<td>*</td>
<td>1 ± 0.2 a</td>
<td>0.6 ± 0.03 b</td>
<td>*</td>
<td>0.2 ± 0.03 a</td>
<td>0.2 ± 0.1 a</td>
<td>ns</td>
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<tr>
<td>Acetaldehyde</td>
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<td>1 ± 0.3 a</td>
<td>ns</td>
<td>2 ± 0.5 a</td>
<td>1 ± 0.1 a</td>
<td>ns</td>
<td>1 ± 0.3 a</td>
<td>1 ± 0.3 a</td>
<td>ns</td>
</tr>
<tr>
<td>Acetone</td>
<td>0.5 ± 0.1 a</td>
<td>0.2 ± 0.02 a</td>
<td>ns</td>
<td>1 ± 0.2 a</td>
<td>0.5 ± 0.04 b</td>
<td>**</td>
<td>0.4 ± 0.1 a</td>
<td>0.4 ± 0.1 a</td>
<td>ns</td>
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<td>Formaldehyde</td>
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<td>ns</td>
<td>0.4 ± 0.1 a</td>
<td>0.1 ± 0.02 b</td>
<td>**</td>
<td>0.2 ± 0.1 a</td>
<td>0.3 ± 0.1 a</td>
<td>ns</td>
</tr>
</tbody>
</table>
Figure legends

**Figure 1**: Ombrothermic diagram for natural and amplified drought in 2012, 2013 and 2014. Bars represent mean monthly precipitation (mm) and curves represent mean monthly temperature (°C). On each amplified drought graph, the percentage represents the proportion of excluded rain compared to the natural drought plot.

**Figure 2**: Diurnal pattern of stomatal conductance ($G_w$) and net photosynthesis ($P_n$) according to drought treatment and season. Values showed represent means ± SE, n=5.

**Figure 3**: Diurnal pattern of isoprene emissions rates, where points represent measured emission and the yellow line corresponds to modelled emissions rates according to the $L+T$ algorithm ($ER_{L+T}$). R² and slope (sl) of correlations between measured and modelled emissions are presented in the yellow frame. Correlations were obtained without forcing data to pass through the origin. Values are mean ± SE, n=5.

**Figure 4**: Diurnal pattern of acetaldehyde emissions rates, where points represent measured emission, the yellow line corresponds to modelled emissions rates according to the $L+T$ algorithm ($ER_{L+T}$) and the dotted line corresponds to modelled emissions rates according to the $T$ algorithm ($ER_T$). R² and slope (sl) of correlations between measured and modelled emissions are presented in the yellow frame for $L+T$ and in the white frame for $T$. Correlations were obtained without forcing data to pass through the origin. Values are mean ± SE, n=5.

**Figure 5**: Diurnal pattern of measured methanol emissions rates. Points represent measured emission, the yellow line corresponds to modelled emissions rates according to the $L+T$ algorithm ($ER_{L+T}$) and the dotted line corresponds to modelled emissions rates according to the $T$ algorithm ($ER_T$). R² and slope (sl) of correlations between measured and modelled emissions are presented in the yellow frame for $L+T$ and in the white frame for $T$. Correlations were obtained without forcing data to pass through the origin. Values are mean ± SE, n=5.
Figure 1:
Figure 2:
Figure 3:
Figure 4:
Figure 5: