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**Emissions and ambient distributions of Biogenic Volatile Organic Compounds (BVOC) in a Ponderosa pine ecosystem: interpretation of PTR-MS mass spectra**

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Abstract

Two proton-transfer-reaction mass spectrometry systems were deployed at the Bio-
hydro-atmosphere interactions of Energy, Aerosols, Carbon, H₂O, Organics and
Nitrogen-Southern Rocky Mountain 2008 field campaign (BEACHON-SRM08; July to
September 2008) at the Manitou Forest observatory in a Ponderosa pine woodland
near Woodland Park, Colorado USA to simultaneously measure BVOC emissions and
ambient distributions of their oxidation products. Here, we present mass spectral anal-
ysis in a wide range of masses (m/z=40⁺ to 210⁺) to assess our understanding of
BVOC emissions and their photochemical process inside of the forest canopy. The
biogenic terpenoids, 2-methyl-3-butene-2-ol (MBO, 50.2%) and several monoterpenes
(MT, 33.5%) were identified as the dominant BVOC emissions from a transmission cor-
rected mass spectrum, averaged over the daytime (11 am to 3 p.m., local time) of three
days. To assess contributions of oxidation products of local BVOC, we calculate a oxida-
tion product spectrum with the OH- and ozone-initiated oxidation product distribution
mass spectra of two major BVOC at the ecosystem (MBO and β-pinene) that were
observed from laboratory oxidation experiments. A majority (∼73%) of the total signal
could be explained by known compounds. The remainder are attributed to oxidation
products of BVOC, emitted from nearby ecosystems and transported to the site, and
oxidation products of unidentified BVOC emitted from the Ponderosa pine ecosystem.

1 Introduction

Emissions and oxidation processes of biogenic volatile organic compounds (BVOC) in
the troposphere have been intensively studied for several decades (Wiedinmyer et al.,
2004; Guenther et al., 1995; Fuentes et al., 2000; Andreae and Crutzen, 1997) due
to their impact on tropospheric photochemistry, secondary organic aerosol formation
potential, and the global carbon cycle. These studies indicate that global biogenic VOC
emissions (∼1150 Tg C/year; Guenther et al., 1995) are an order of magnitude higher
than anthropogenic emissions (~160 Tg C/year; Benkovitz et al., 2004). The oxidation processes of BVOC affect local and global air quality. For example, Ryerson et al. (2001) found that reactive BVOC (e.g. isoprene) are an important factor controlling ozone formation downwind of power plant plumes in the southeastern US Kanakidou et al. (2005) reviewed the importance of BVOC for processes influencing the formation of secondary organic aerosol and cloud condensation nuclei, which have significant implications for global climate (IPCC, 2007). In addition, Guenther (2002) and Kesselmeier et al. (2002) concluded that reactive BVOC have significant implications for the global carbon cycle.

Our understanding of BVOC emissions and photochemistry of BVOC has been significantly advanced in the past decade (e.g Guenther et al., 2006; Atkinson and Arey, 2003) yet a number of recent laboratory and field studies have indicated that significant issues related to unexplored BVOC or their oxygenated products still remain unsolved. For example, Zhao et al. (2004) reported a significant yield (~30%) of C4 and C5 carbonyl compounds from the oxidation of isoprene (C₅H₈), yet these species have not been measured in ambient air due to technical difficulties (Goldstein and Galbally, 2007). A field study in a Ponderosa pine forest canopy (Lee et al., 2005) reported that total monoterpene (MT) concentrations, measured by proton transfer reaction-mass spectrometry (PTR-MS) were consistently higher (~30%) than sums of the speciated MT concentrations, measured by the gas chromatography method. The investigators suggest that this difference indicates that there are significant amounts of reactive MT that have not been detected using conventional Gas Chromatography (GC) analytical methods. Indeed, significant missing OH reactivity (Di Carlo et al., 2004) and unexpected chemical ozone loss (Kurpius and Goldstein, 2003), reported inside of forest canopies suggest that undetected BVOC could have significant implications for photochemistry.

Here we present a detailed mass spectral analysis (m/z=40⁺ to 210⁺) of BVOC emissions using PTR-MS and GC-MS. We compare branch enclosure BVOC emission measurements with ambient VOC distributions in a Ponderosa pine forest. The
universal sensitivity of PTR-MS for most VOC exhibiting a proton affinity greater than water provided a capability to examine unidentified BVOC along with complementary measurements made by GC-MS techniques. Recent work on quantifying the impact of mass discrimination characteristics of the PTR-MS system (from $m/z = 40^+$ to $m/z = 210^+$; e.g. Kim et al., 2009; Taipale et al., 2008) provide new constraints for quantification of unidentified BVOC and their oxidation products in the higher molecular weight mass range (e.g. >100 amu). Using these constraints, we compare the observed ambient spectra of forest air with the laboratory observed OH and ozone initiated oxidation product mass spectra of two major BVOC emissions in the ecosystem, 2-methyl-3-butene-2-ol (MBO) and $\beta$-pinene. Our analysis examines the main uncertainties associated with sources of missing organic carbon in forest environments (e.g. primary production of BVOC vs. photochemical production) (Di Carlo et al., 2004; Holzinger et al., 2005).

2 Methods

2.1 Site

The field study was conducted at the Manitou Forest observatory (MFO, https://wiki.ucar.edu/display/mfo/Manitou+Forest+Observatory) from 21 July to 19 September 2008. The MFO is located in the Manitou Experimental Forest (elevation 2286 m, latitude 39°6′0″, and longitude 105°5′30″), maintained by the US Forest Service. The site is representative of the Central Rocky Mountains Ponderosa pine zone. Year long trace gas monitoring at the site (Smith et al., 2009) indicates that clean air masses from the southwest predominate and anthropogenic pollution plumes from nearby cities such as Colorado Springs and Denver were rarely transported to the site.
2.2 Branch enclosure system

A 5L Teflon bag with two \(1/4''\) Teflon tube fittings (Welch Fluorocarbon, Dover, NH) enclosed a Ponderosa pine branch with mature needles that was located approximately four meters above the ground. Ambient air was pumped through a charcoal scrubber to remove VOC and ozone and then introduced into the enclosure at a rate of \(\sim 2.5\) slm (standard liter per minute). VOC concentrations in the scrubbed air entering the enclosure were measured once every hour for background subtractions. Mass spectra obtained by PTR-MS during the night when BVOC emissions were significantly lower than those during the day indicated that there was no significant background interference with the BVOC emission measurements. Air temperature inside the enclosure was measured using a K-type thermocouple and logged on a computer. We found that BVOC emissions were always higher during the first day following installation of the enclosure. To avoid BVOC artifacts due to mechanical disturbance during installation, data collected during the first day after the installation were therefore excluded. Measurements of BVOC emissions were conducted for four full days after emissions had stabilized. After each measurement period, the enclosure was opened, exposing the branch to ambient conditions for 2 days before enclosing it again for subsequent measurements. At the conclusion of the experiment, the branch was harvested, and needles were dried at 70°C for \(\sim 48\) h and weighed.

2.3 PTR-MS and GC-MS

Two high sensitivity PTR-MS systems (IONICON Analytik, Innsbruck Austria) were deployed during the field campaign. General information for the analytical technique can be found in Blake et al. (2009) and references therein. Kim et al. (2009) and Karl et al. (2005) presented information on the analytical characteristics and calibration procedures for the specific instruments used in this study. Since we explore a wide range of molecular masses \((m/z=40^+\) to \(210^+)\), the mass discrimination of the system was carefully characterized before and after the field campaign with a certified standard gas.
(Matheson Tri Gas, USA) containing 14 aromatic compounds from benzene (MW 78) to trichlorobenzene (MW 180). The relative sensitivity over the mass range generally declines in an exponential function. In addition, we calibrated relative sensitivities at \( m/z = 205^+ \), the molecular weight of the protonated sesquiterpene (SQT) parent ion with a triisopropylbenzene standard generated by a capillary diffusion system (CDS) located at the University of Colorado at Boulder (Helmig et al., 2004). The experiment demonstrated that the exponential fitting line, based on the certified aromatic standards, predicts the transmission efficiency reasonably well (Kim et al., 2009). The average deviation from the transmission curve is estimated to be \( \sim 15\% \), an error very similar to the estimation of Taipale et al. (2008).

Branch enclosure and ambient air samples were also analyzed using GC-MS methods. In situ measurements were made at the field site using a portable GC-MS with an internal microtrap concentrator and a 30 m × 0.32 mm ID 1 mm film DB-1 column, temperature programmed from 40°C to 200°C (Hapsite Smart, INFICON, East Syracuse, NY, USA). Air from the enclosure was directly routed to the portable GC-MS to collect 1 L samples on the microtrap and used for quantitative speciation of MBO, and the dominant MT, oxygenated-MT and qualitative speciation of SQT. Higher volume branch enclosure and ambient air samples (6 L) were collected onto stainless steel two-stage sorbent tubes filled Tenax and Carbotrap, and then thermally desorbed and analyzed with the portable GC-MS. An ozone trap (LpDNPH ozone scrubber (KI), Supelco USA) was used on the ambient air-sampling inlet. To examine tree-to-tree variations of BVOC emission, leaf cuvette samples with Tenax/Carbotrap sorbent tubes were also analyzed using a GC-flame ionization detector (FID). Additionally, diurnal variations of ambient BVOC were measured by collecting air in stainless steel canisters (2 L), which were subsequently analyzed by an Agilent GC-MS in laboratories in Boulder, CO and Portland, OR.
2.4 Environmental chamber

After the field campaign, we conducted laboratory oxidation experiments for two of the major BVOC species (MBO, β-pinene) observed at the Manitou forest site using a stainless steel environmental chamber (Shetter et al., 1987), 2 m in length with a volume of 47 L. For each oxidation experiment, the concentrations of reactants and products were monitored by both a Bomem DA3.01 Fourier transform infrared spectrometer and the PTR-MS system. The experimental details of OH and ozone-initiated experiments can be found in Orlando et al. (2000).

3 Results

3.1 Fragmentation patterns

For accurate interpretations of mass spectra, it is essential to have a comprehensive understanding of the fragmentation patterns of analyte ions. Fragmentation patterns of various compounds have been extensively investigated with different reagent ion systems and chemical ionization has been clearly shown to result in less pronounced fragmentation than electron impact ionization (Harrison, 1992). A number of studies using PTR-MS have reported fragmentation patterns of BVOC (e.g. Kim et al., 2009; Tani et al., 2003). In this section, we summarize fragmentation patterns of BVOC groups for the H$_3$O$^+$ reagent ion system based on their functional. We focus here on BVOC relevant to this study, and refer to other references (de Gouw and Warneke, 2007) for other VOC.

In general, reagent ions with higher proton affinities in the Brønsted acid chemical ionization reagent system indicate more pronounced fragmentation for the same analyte ions. Chemical ionization by H$_3$O$^+$, which has a relatively moderate proton affinity (PA=166.5 kcal/mol) among the Brønsted acid reagent ions, leads to moderate fragmentation patterns. Break-up patterns of molecular ions are highly dependent
on functional groups as summarized in Table 1. In addition, for PTR-MS applications, higher ratios of the electric field strength to the number of molecules in the drift tube (E/N) cause a more pronounced degree of fragmentation (Kim et al., 2009; Tani et al., 2003). Usually, E/N is set in the range between 100 and 140 Td (10⁻¹⁷ V cm²) and the experimental results presented here were collected at E/N of ~120 Td (unless otherwise noted). For example, MT (Tani et al., 2003) and SQT (Dhooghe et al., 2008; Kim et al., 2009; Demarcke et al., 2009) decompose by splitting off the same 56 amu (C₄H₈) neutral fragment. This leaves the fragment ions m/z=81⁺ for MT and m/z=149⁺ for SQT as the dominant fragments regardless of the structure of the isomers. Chemical structure, however, can affect the relative abundances of the parent and fragment ions. p-Cymene, a monoterpenoid compound with the molecular mass of 134 amu has a neutral C₃H₆ fragment that leads to an ion detected at m/z=93⁺ (~50% abundance at ~120 Td; Tani et al., 2003). This implies that p-cymene could be a significant interference for toluene measurements in environments with large p-cymene concentrations. Therefore, complementary methods such as GC-MS should be employed to estimate toluene emissions from plants with careful assessments of m/z=93⁺ signals from p-cymene (Heiden et al., 1999; White et al., 2009).

During proton transfer reactions, alcohols split off a H₂O group as the major neutral fragment. This leads for example to a relatively small parent ion abundance for MBO (20–30%) and a negligible parent ion abundance (e.g. less than 1%) for substituted MT alcohols, such as linalool or borneol. Substituted MT alcohols are therefore detected at the same masses as MT, m/z=81⁺ and 137⁺. Bornyl acetate (MW=196) is also detected at m/z=137⁺ and 81⁺ after losing a neutral fragment with the molecular composition of C₂H₄O₂. Many studies have reported noticeable emissions of these oxygenated terpenoid compounds (e.g. Ciccioli et al., 1999); thus, care needs to be taken when estimating MT emissions based on PTR-MS alone. Most aldehyde and epoxide species have common dominant dehydration fragments as summarized in Table 1. Two ketoaldehyde species, pinonaldehyde and caronaldehyde are first generation oxidation products of α-pinene and 3-carene, respectively. The fragmentation pattern
of pinonaldehyde has been reported twice using two different ion reaction chamber configurations—a drift tube (PTR-MS; Wisthaler et al. 2001) and a flow tube (SIFT-MS; Schoon et al., 2004). Results from the two studies are qualitatively similar in that the dehydrated fragment ion (m/z=151+) is the dominant product; other observed fragment ions exhibit the same masses but are more abundant in PTR-MS mass spectra due to the more pronounced fragmentation compared to the SIFT technique. The fragment ion distribution of caronaldehyde observed by the SIFT-MS technique is slightly different from that of pinonaldehyde. Along with the dehydrated ion (m/z=151+), a significant fraction of the parent ion (m/z=169+) and another major fragment (m/z=123+) were observed, both with abundances comparable to that of m/z=151+. These differences could originate from structural differences of the two compounds (e.g. the ring systems). The results suggest that fragmentation patterns of other first generation oxidation products of MTs (e.g. limonenaldehyde) need to be examined for the accurate assessment of product ion yields in photochemical kinetic studies (e.g. Ng et al., 2006). In addition, ketones and ethers listed in the table are detected mostly (>90%) at the parent ion masses.

3.2 BVOC emissions from Ponderosa pine

For the first week of the field campaign, a PTR-MS system was set up to measure mass spectra from a branch enclosure. A three-day averaged mass spectrum (12 to 14 August 2008) of the branch enclosure sample is presented in Fig. 1a for daytime (11 a.m. to 3 p.m.) conditions. Figure 1b presents the transmission-corrected spectrum. The scale on the y-axis represents the relative abundance of the mass spectra normalized to the most abundant peak (m/z=69+, the MBO fragment). Based on the fragmentation patterns, summarized in Table 1, the identified BVOC species and their relative abundances in the transmission-corrected spectrum are summarized in Table 2. The table also summarizes the speciation of MT, oxygenated-MT, and SQT, identified by GC-MS and available proton transfer reaction rates of each species. Emission strengths measured by PTR-MS are proportional to the relative abundances...
normalized by proton transfer reaction constants as summarized in the last column of Table 2. MBO and MT are the dominant BVOCs emitted from Ponderosa pine as observed in earlier studies (Baker et al., 2001). The emission strength of MT that was measured by GC-MS is \( \sim 72\% \) of that measured by PTR-MS, the result is consistent with the findings of Lee et al. (2005), who suggested the existence of reactive MT that can only be measured by PTR-MS. Less than 5% of the discrepancy can be explained by the contributions of oxygenated MT, such as linalool and bornyl acetate, which were detected on \( m/z=81^+ \) and \( 137^+ \). In addition, the GC-MS detected elevated levels of toluene from the branch enclosure (\( \sim 4 \) times higher than ambient concentration). With subtraction of the p-cymene background based on the results of Tani et al. (2004), we can estimate that toluene emissions were less than 1% of MT emissions. This is much lower than the reported value of a previous study (Heiden et al., 1999). Although pronounced acetone, acetaldehyde and acetic acid peaks were observed in the spectrum, the flux of these compounds is strongly dependent on their compensation point and on the concentrations in the surrounding air. Since their concentration in the branch enclosure differs from that of ambient air, ecosystem-scale flux measurements would be preferable to estimate their fluxes (Kesselmeier et al., 1997; Schade and Goldstein, 2001; Karl et al., 2005; Jardine et al., 2008). The total counts listed in Table 2 represent 93% of the total counts in the spectrum of Fig. 1b. This suggests that we have a reasonably good understanding of the composition of BVOC emissions from Ponderosa pine in the mass spectral range investigated here for compounds that can be detected by PTR-MS. However, the lower sensitivities of PTR-MS techniques for high mass compounds of PTR-MS techniques (M.W.>150) could mean that we are missing some heavier compounds.

After a week of conducting mass scan measurements, we changed the PTR-MS measurement mode to multiple ion detection, which enables shorter averaging times due to a better signal to noise ratio. These measurements were conducted for three weeks, focusing on emission rates of MBO, MT, SQT, and oxygenated-MT (\( m/z=153^+ \)). Mean measured daytime emission rates (11 a.m. to 3 p.m.) along with
daily temperature variations are summarized in Table 3. The average emission ratios for MT/MBO and SQT/MT were 0.72 (±0.16) and 0.01 (±0.004), respectively. During the same campaign we conducted ecosystem-scale gradient measurements using a second PTR-MS instrument. From these we calculated an ecosystem-scale flux ratio of 0.50±0.15 and 0.04±0.01 for MT/MBO and SQT/MT respectively. Given significant uncertainties when comparing branch enclosure and ambient flux measurements, MT/MBO and SQT/MT ratios compare well between these different methods. More comprehensive approaches including investigations of photochemical processes inside of the forest canopy and measurements on a large number of branches to characterize branch-to-branch variability are planned for future studies at the MFO site and will address these differences more quantitatively. Emission rates of oxygenated-MT listed in Table 3 are calculated based on the signals on m/z=153⁺, the sensitivity measured for MT and the proton transfer reaction rate of camphor (4.4×10⁻⁹ cm³ s⁻¹; Tani et al., 2003). The estimated emission rates are significantly lower than those of MT but much higher than SQT emission rates, though daily emission rate variations of SQT indicate a wide range (e.g. one order of magnitude). Such large emission variations have been reported previously on longer timescales (Arey et al., 1995; Ciccioli et al., 1999; Vuorinen et al., 2005; Ruuskanen et al., 2007; Bouvier-Brown et al., 2009) but the underlying endogenous (e.g. developmental stage of branch) and exogenous (e.g. temperature and light) controlling factors remain uncertain. Table 3 also contains the results from an exponential fitting function between temperature and emission rates used to parameterize emissions from this. The exponential fitting scheme is analogous to the algorithm of Guenther et al. (1993), E(temp)=E₃₀exp(β(temp−30)), where E₃₀ indicates the emission capacity at 30°C for a given dry needle mass and β indicates the temperature response factor. Analogous measurements of E₃₀ and β from recent Ponderosa pine studies are also shown in Table 3, and β-values between different studies show reasonable agreement but the small difference (∼0.05) in β could make significant differences in BVOC emission estimation. Emission capacity varies significantly between different studies. Leaf cuvette measurements of various Ponderosa pine trees
at the Manitou forest site (Table 3) indicated reasonable agreements with the branch emission measurement results for MBO and MT. In addition, the low emissions of SQT in the branch enclosure measurement were consistent with the results of leaf cuvette measurements for various Ponderosa pine individuals. Bouvier-Brown et al. (2009) observed large variations of emission capacity (factor of 10) for the same Ponderosa pine samplings suggesting that plant-to-plant variations among the same species in an ecosystem can be significant although some levels of uncertainty could be caused by artifacts associated with enclosure measurement techniques. These relatively large emission variations have been reported from experiments for many plant species and ecosystems and could be caused by biotic or abiotic stresses (e.g. herbivory and violent weather events; Duhl et al., 2008).

3.3 BVOC and their oxidation product distributions in the forest canopy

Figure 2 presents the diurnal variations of ambient concentrations of MBO, MT, acetone ($m/z=59^+$) and glycolaldehyde+acetic acid ($m/z=61^+$), averaged over a three-week period (18 August to 6 September 2008). The diurnal variations of MBO and MT, two major BVOC emitted by the ecosystem, show a clear anti-correlation. Emissions of MBO are strongly light dependent (Harley et al., 1998) leading to maximum mixing ratios during the day despite a shorter chemical lifetime and dilution into a deeper boundary layer. A similar diurnal trend was also observed at a Ponderosa pine ecosystem in California (Schade and Goldstein, 2001). On the other hand, the lower boundary layer depth during the night seems to be the main controlling factor of the diurnal variation of MT concentrations, with maximal mixing ratios occurring at early morning despite lower emissions resulting from cooler temperatures. The dominant MT species, observed during the field campaign period was β-pinene (26%), followed closely by α-pinene (22%) and 3-carene (21%). The results correspond with emission measurements from the branch enclosure system. The diurnal variation of compounds with sources that include both direct emissions and BVOC oxidation such as acetone, possibly contributed by direct emissions, and acetic acid is more complex due to photochemical production...
during daytime and variations in boundary layer height throughout the day.

In order to compare differences in diurnal oxidation product distributions, Fig. 3 shows an MBO-dominated daytime spectrum (2 to 3 p.m. on 14 August; Fig. 3a) as well as a MT-dominated nighttime spectrum (1 to 2 am on August 18; Figure 3b). Figures 3c and d present transmission-corrected mass spectra for day and night, respectively. Transmission-corrected spectra were calculated based on acetone (m/z=59+) as a reference mass. Major peaks and their transmission-corrected counts from the spectra are summarized in Table 4. The table also shows mixing ratios of NOy and SO2 during the time period, which indicate that the air masses were not influenced by pollution, a conclusion supported by the relatively weak peak strengths of benzene (79+) and toluene (93+), even though ambient toluene concentrations from PTR-MS measurements might reflect an upper limit. Formic acid exhibits the highest contribution during both day and night. Other oxidation products (acetaldehyde, acetone, acetic acid+glycolaldehyde, 2-hydroxy-2-methylpropanal (HMPr), methyl ethyl ketone (MEK)) were also detected in significant amounts. m/z=153+, most likely an oxygenated-MT (e.g. camphor) was also detected in significant amounts during both day and night. A number of studies using GC techniques have reported emissions of oxygenated-MT that could be detected on m/z=153+ by PTR-MS. Noticeable emissions of camphor and other oxygenated-MT have been reported especially from conifers (Ortega et al., 2008). In this ecosystem, we confirmed the emission of oxygenated-MT based on both GC-MS and PTR-MS measurements. We also observed significant concentrations in ambient air. In addition, some unidentified masses in the spectra are summarized in Table 4. m/z=75+, 57+, and 101+ in the table could be oxidation products of isoprene, which is a minor BVOC emission from this ecosystem but is emitted in greater amounts from some upwind ecosystems. Indeed, only small amount of isoprene (∼0.1–0.3 ppbv) were found in canister samples, collected on site and analyzed via GC-MS. The concentrations did not show any strong diurnal variation, suggesting that isoprene at this site is transported from other ecosystems (e.g. aspen groves, oak shrublands, spruce forests, riparian willow zone) with isoprene oxidation products produced during transport. The
significance of transported isoprene oxidation products in this Ponderosa pine dominated ecosystem was also reported by Dreyfus et al. (2002) at the Blodgett Forest research station (CA). The compounds/compound classes listed in Table 4 account for 74% and 73% of the observed ambient mass spectra (without the inclusion of even masses to focus on hydrocarbon ($C_xH_y$) and oxygenated-hydrocarbon ($C_xH_yO_z$) compounds) during day and night respectively. The larger fraction of unidentified counts in ambient mass spectra (~24%), compared to those observed from branch enclosure measurements (~9%), suggests the influence of photochemical VOC production observed in this clean environment. The discussion, again, is limited due to the low sensitivity of PTR-MS for higher mass compounds. Recent technical breakthroughs such as a time-of-flight detector or ion trap mass spectrometers are expected to expand our understanding of the chemistry of semi-volatile organic compounds (Mielke et al., 2008; Jordan et al., 2009).

To quantitatively assess contributions of oxidation products of BVOC in this ecosystem, we conducted laboratory kinetic experiments for two major BVOC, MBO and β-pinene. OH- and ozone-initiated oxidation products and their yields from β-pinene were quantified with PTR-MS by Wisthaler et al. (2001) and Lee et al. (2006), respectively but the products of OH or ozone initiated oxidation of MBO have not been reported with PTR-MS. For each oxidation experiment, the PTR-MS was operated in a mass-scanning mode ($m/z=40^+$ to $210^+$). A background mass spectrum for OH oxidation experiments was taken for all chemicals (air matrix 800 Torr), introduced into the environmental chamber. For ozone experiments, we also obtained background spectra before ozone was injected into the environmental chamber. As oxidation reactions progressed, spectra were taken and the blank spectrum subtracted to get the net product distribution spectra due to photo-oxidation.

An example of a net spectrum is presented in Fig. 4. The mass discrimination-corrected spectrum in the figure is product distributions from the oxidation of MBO by OH. Peaks of the spectrum are normalized by acetone counts ($m/z=59^+$), the dominant peak in the spectrum. Other pronounced peaks can be matched with known product
molecular ions such as 2-hydroxy-2-methylpropanal (HMPr, m/z=89\textsuperscript{+}) and its dehydrated ion (m/z7=1\textsuperscript{+}), and glycolaldehyde (m/z=61\textsuperscript{+}). The protonated glycolaldehyde ion was shown to have a dehydrated fragment of m/z=43\textsuperscript{+} with an intensity equal to the parent ion abundance in the E/N configuration of laboratory experiments (∼130 Td). However, due to interferences from OH precursors on m/z=43\textsuperscript{+}, the pronounced peak at this mass could not be quantified in the product distribution spectrum. The relative abundances of products and their product yields (brackets) calculated from spectra are summarized in Table 5. The table also shows product yields, calculated from previous FT-IR experiments (Carrasco et al., 2007; Atkinson and Arey et al., 2003, and references therein). Overall, the FT-IR observed product yields of this study show reasonable agreement with previous studies. The estimated yield of acetone from the PTR-MS experiments in Table 5 shows reasonable agreement with previous FT-IR experiment results. The glycolaldehyde yield is estimated from the calculated proton transfer reaction constant (kₐₒₒₒ =2.97×10⁻⁹ cm³ s⁻¹ from the average-dipole-orientation theory) and the estimated product ion counts, twice that of the parent ion (m/z=61\textsuperscript{+}) counts in the spectrum, to account for fragmentation. PTR-MS detected acetone, HMPr and formic acid and FT-IR detected formaldehyde, acetone, and formic acid as major ozone-initiated oxidation products of MBO in the dry condition. Carrasco et al. (2007) reported same results in the dry condition. However, the same study also found that only HMPr and formaldehyde are detected as oxidation products in humid condition (RH>20%).

Product distributions of β-pinene oxidation with OH and ozone indicate reasonable agreement with previously reported results. Both PTR-MS and FT-IR measured nopinone as a major oxidation product. FT-IR detected formaldehyde and acetone and PTR-MS detected acetone, formic acid, hydroxy norpinone (only by ozone oxidation), and 3-oxonorpinone as oxidation products. PTR-MS detected relatively small amount of m/z=169\textsuperscript{+} in the product spectrum by ozone-initiated oxidation. The peak is possibly from dehydrated fragments of protonated pinic acid (m/z=187\textsuperscript{+}). The product mass spectrum of OH oxidation experiments indicates pronounced m/z=101\textsuperscript{+} (with a relative
abundance of 25.7) similar to what we observed in the ambient spectra at the Mani-
tou forest site. The oxidation products abundances described above explain 75.9% of
total counts in the OH oxidation spectrum and 75.4% of total counts from the ozone
oxidation spectrum.

To compare the product distributions of the oxidation experiments with the daytime
ambient spectrum in the forest canopy, we conducted a simple time-dependent cal-
culation to get ratios of [glycolaldehyde]/[MBO] and [nopinone]/[β-pinene]. The cal-
culations were constrained by the measured daily variations of MT, MBO (Fig. 2) and
ozone. Daily variations of OH concentrations were set to follow daily variations of
\( J_{NO_2} \), calculated by the TUV (Total Ultraviolet and Visible http://cprm.acd.ucar.edu/
Models/TUV/) model with a maximum value of \( 2.5 \times 10^6 \) molecules cm\(^{-3} \) (OH). The
yields of glycolaldehyde and nopinone from MBO and β-pinene oxidation, respec-
tively, were assigned the values found in this study. The OH loss rates of glycolalde-
hyde (\( k_{glycolaldehyde-OH} = 1.1 \times 10^{-11} \) at 298 K) and nopinone (\( k_{nopinone-OH} = 1.7 \times 10^{-11} \) at
298 K) were taken from Bacher et al. (2001) and Calogirou et al. (1999), respectively.
The daily variations of glycolaldehyde and nopinone in the model become consistent
in 48 h of the model time. From the stabilized diurnal variations, the average ratios of
[glycolaldehyde]/[MBO] (=1.62) and [nopinone]/[β-pinene] (=0.824) between 2 p.m. to
3 p.m., the time period of the average daytime ambient mass spectrum in Fig. 3c were
obtained. Based on the ratio, a composite spectrum was calculated from four mass
spectra of the laboratory oxidation experiments. The oxidation product distribution
mass spectra were scaled based on ambient oxidant concentrations of OH (assumed
to be \( 2.5 \times 10^6 \) molecules cm\(^{-3} \)) and ozone (45 ppbv from ambient measurements) and
the amount of MBO and β-pinene in the ambient spectrum of Fig. 3c. The calculated
composite spectrum along with the ambient spectrum is presented in Fig. 5.

The composite spectrum is dominated by the known oxidation products of MBO and
β-pinene such as acetone, glycolaldehyde, HMPr, and nopinone. Those compounds
comprised more than half of the total counts in the ambient spectrum as described in
the above section. However, a significant fraction of peaks in the ambient spectrum

20834
are not present in the composite spectrum. The additional peaks may come from oxidation products of other MT species such as α-pinene and 3-carene, which represented 22% and 20% of the ambient MT concentrations. However, the relatively weak signal on m/z=151+, a major fragment for both pinonaldehyde (an α-pinene oxidation product) and caronaldehyde (a 3-carene oxidation product) suggests that photo-oxidation products of these two MTs can only account for a small fraction of unidentified peaks. Therefore, further investigations such as possible oxidation product contributions from oxygenated-MT (m/z=153+) are necessary. As ambient mass spectra indicated influences of transported oxygenated products of VOC not emitted in the ecosystem (e.g. isoprene), a significant portion of unidentified peaks in the spectra may be attributable to this process. In addition, the possibility of significant emissions of semi-volatile organic compounds, detected by PTR-MS with low sensitivity (i.e. with amu>150), leads us to speculate that their oxygenated products could contribute to the unidentified peaks, although many studies have demonstrated that a significant portion of oxidation products of SVOC is partitioned into the aerosol phase due to their low vapor pressure (Kroll and Seinfeld, 2008).

4 Summary

We examined emissions and photochemical processes within a Ponderosa pine ecosystem with comprehensive PTR-MS and GC-MS measurements. Most of the peaks in the mass spectra of the branch enclosure emission system could be allocated to BVOC, which were identified by GC-MS. The identified species comprise most of the signals (∼93%) in the mass spectrum. Both PTR-MS and GC-MS observed MBO and MT as major emissions. Emission measurements from a leaf cuvette system with GC-FID analysis showed relatively consistent emission rates of MBO and total MT from different Ponderosa pine trees in the ecosystem. Significantly lower amounts of SQT emissions were quantified by PTR-MS and GC-MS than emission rates previously reported for Ponderosa pines. As the importance of SQT has been highlighted for
their significant SOA (secondary organic aerosol) formation potential (Sakulyanontvit-taya et al., 2008), comprehensive research on the biotic and abiotic factors controlling SQT emission is necessary. Oxygenated-MT such as camphor were also quantified by both PTR-MS and GC-MS. The estimated emission rates were in the range of 0.1 to 0.4 µg g⁻¹ h⁻¹, lower than MT emission rates but significantly higher than SQT emission rates.

The observed ambient temporal variation patterns of MBO and MT, the major BVOC emitted from the ecosystem, and their major oxidation products, acetone and glycolaldehyde+acetic acid (m/z=61⁺) were controlled by diurnal emission and boundary layer characteristics, and were similar to observations reported previously. In the transmission corrected ambient mass spectrum obtained during daytime, ~73% of the total signal could be explained by known compounds. To explore sources of unidentified peaks, OH- and ozone-initiated oxidation spectra of the primary emission species at the site (MBO and β-pinene) were generated using a laboratory chamber facility. In general, the oxidation product distributions in the PTR-MS spectra corresponded with those found in previous studies. With four different laboratory-derived product distribution spectra, we calculated the oxidation product distribution spectrum of the ecosystem. This spectrum could not explain most of the unidentified peaks, which make up ~27% of the total transmission-corrected signals in the ambient spectrum. The potential sources for these unidentified species include 1) BVOC oxidation products, transported from the other ecosystems upwind, and 2) oxidation products of unidentified BVOC emitted from the ecosystem that are not detected due to analytical limitations of PTR-MS and GC techniques (e.g. semi-volatile organic compounds). Recent technical breakthroughs such as the time-of-flight mass spectrometry technique are expected to address these issues.
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Emissions of BVOC in a Ponderosa pine ecosystem

S. Kim et al.


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Lee, A., Schade, G. W., Holzinger, R., and Goldstein, A. H.: A comparison of new measure-


Table 1. A summary of fragmentation patterns of BVOC relevant to this study. Major product ions (relative abundance > 10%) are listed and minor product ions are listed in the brackets at a drift tube energy of ~120 Td.

<table>
<thead>
<tr>
<th>Classes</th>
<th>Species</th>
<th>Fragments</th>
</tr>
</thead>
<tbody>
<tr>
<td>Terpenoid</td>
<td>Monoterpenes∗</td>
<td>( m/z = 81^+ ) and ( 137^+ ) (67^+, 93^+, and 95^+)</td>
</tr>
<tr>
<td></td>
<td>Sesquiterpenes+</td>
<td>( m/z = 149^+ ) and ( 205^+ ) (81^+, 95^+, 109^+, 123^+, 135^+, and 137^+)</td>
</tr>
<tr>
<td></td>
<td>p-cymene∗</td>
<td>( m/z = 93^+ ) and ( 135^+ )</td>
</tr>
<tr>
<td>Alcohol</td>
<td>MBO</td>
<td>( m/z = 69^+ ), ( 87^+ ) (41^+)</td>
</tr>
<tr>
<td></td>
<td>Linalool</td>
<td>( m/z = 81^+ ) and ( 137^+ )</td>
</tr>
<tr>
<td></td>
<td>Borneol</td>
<td>( m/z = 81^+ ) and ( 137^+ )</td>
</tr>
<tr>
<td>Ketone</td>
<td>norpinone∗∗</td>
<td>( m/z = 139^+ ) (121^+)</td>
</tr>
<tr>
<td></td>
<td>camphor∗</td>
<td>( m/z = 153^+ )</td>
</tr>
<tr>
<td>Aldehyde</td>
<td>glycolaldehyde</td>
<td>( m/z = 61^+ ) and ( 43^+ )</td>
</tr>
<tr>
<td></td>
<td>pinonaldehyde**</td>
<td>( m/z = 151^+, 107^+, ) and ( 71^+ ) (43^+, 81^+, 99^+, 109^+, 123^+, ) and ( 169^+ ))</td>
</tr>
<tr>
<td></td>
<td>Caronaldehyde++</td>
<td>( m/z = 169^+, 151^+, ) and ( 123^+ ) (107^+, 109^+, 125^+, 139^+, ) and ( 151^+ ))</td>
</tr>
<tr>
<td></td>
<td>HMPri</td>
<td>( m/z = 71^+ ) and ( 89^+ )</td>
</tr>
<tr>
<td>Epoxide</td>
<td>( \alpha )-pinene oxide</td>
<td>( m/z = 153^+, 135^+, 109^+ ) (43^+, 93^+, ) and ( 95^+ ))</td>
</tr>
<tr>
<td>Ether</td>
<td>methylchavicol</td>
<td>( m/z = 149^+ )</td>
</tr>
<tr>
<td>Ester</td>
<td>Methyl salicylate+++</td>
<td>( m/z = 153^+ )</td>
</tr>
<tr>
<td></td>
<td>bornyl acetate</td>
<td>( m/z = 81^+ ) and ( 137^+ )</td>
</tr>
</tbody>
</table>

* Tani et al. (2003, 2004)
** Whisthaler et al. (2001) and Schoon et al. (2004)
† Kim et al. (2009) and Demarcke et al. (2009)
++ Schoon et al. (2004)
+++ Karl et al. (2008)
Table 2. Observed compounds/compound classes, product ions and relative abundances in the transmission-corrected mass spectrum (3rd column). The proton transfer reaction rate corrected abundances (PTR Corr. Abd.) along with proton transfer rate constants (PTR constants) are also summarized in 5th and 4th column.

<table>
<thead>
<tr>
<th>Species</th>
<th>Product Ions</th>
<th>Abundances</th>
<th>Detected Species</th>
<th>PTR Constants</th>
<th>PTR Corr. Abd.</th>
</tr>
</thead>
<tbody>
<tr>
<td>MBO</td>
<td>69+ and 87+</td>
<td>50.74</td>
<td></td>
<td>2.6</td>
<td>50.15</td>
</tr>
<tr>
<td>MT</td>
<td>81+ and 137+</td>
<td>29.97</td>
<td>β−pinene (25.1%), 3-carene (17.1%), limonene (15%), mycene(17.3%) and ten other species</td>
<td>2.3</td>
<td>33.48</td>
</tr>
<tr>
<td>other MT</td>
<td>93+, 133+, 135+</td>
<td>2.81</td>
<td>p-cymeme, o-cymene, and menthatriene</td>
<td>2.4</td>
<td>3.05</td>
</tr>
<tr>
<td>O-MT</td>
<td>153+, 151+</td>
<td>2.49</td>
<td>verbenone, car-3-en-2-one, and carvone (m/z=151⁺), and camphor, t-carveol, and 2-hydroxy methyl ester (m/z=153⁺)</td>
<td>4.4</td>
<td>1.45</td>
</tr>
<tr>
<td>m/z=169+</td>
<td>169+</td>
<td>0.36</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>SQT</td>
<td>205+</td>
<td>0.15</td>
<td>Mostly Isolongifolene</td>
<td>3.0</td>
<td>0.13</td>
</tr>
<tr>
<td>Acetaldehyde</td>
<td>45+</td>
<td>3.36</td>
<td></td>
<td>3.4</td>
<td>2.57</td>
</tr>
<tr>
<td>Acetone</td>
<td>59+</td>
<td>8.35</td>
<td></td>
<td>3.0</td>
<td>7.15</td>
</tr>
<tr>
<td>Acetic Acid</td>
<td>61+</td>
<td>1.79</td>
<td></td>
<td>2.3</td>
<td>2.02</td>
</tr>
</tbody>
</table>
Table 3. Daily variations of emission rates of four BVOC observed from branch enclosures, measured by PTR-MS. Leaf cuvette measurement results are also summarized for measurements made at the same ecosystem with different Ponderosa pine trees. Emission capacity ($E_{30}$) and $\beta$ derived from this study and previous studies of Ponderosa pine of different ecosystems are also presented.

<table>
<thead>
<tr>
<th>Date</th>
<th>MBO (µg g$^{-1}$ h$^{-1}$)</th>
<th>MT (µg g$^{-1}$ h$^{-1}$)</th>
<th>OH-MT (µg g$^{-1}$ h$^{-1}$)</th>
<th>SQT (µg g$^{-1}$ h$^{-1}$)</th>
<th>Temp</th>
</tr>
</thead>
<tbody>
<tr>
<td>21 Aug 2008</td>
<td>4.85</td>
<td>3.6</td>
<td>0.400</td>
<td>0.0343</td>
<td>30.9</td>
</tr>
<tr>
<td>25 Aug 2008</td>
<td>4.67</td>
<td>4.66</td>
<td>0.438</td>
<td>0.0825</td>
<td>30.5</td>
</tr>
<tr>
<td>26 Aug 2008</td>
<td>5.07</td>
<td>4.52</td>
<td>0.460</td>
<td>0.0453</td>
<td>30.3</td>
</tr>
<tr>
<td>27 Aug 2008</td>
<td>4.56</td>
<td>3.67</td>
<td>0.427</td>
<td>0.0425</td>
<td>30.7</td>
</tr>
<tr>
<td>28 Aug 2008</td>
<td>3.57</td>
<td>2.44</td>
<td>0.197</td>
<td>0.0243</td>
<td>27.0</td>
</tr>
<tr>
<td>2 Sep 2008</td>
<td>2.43</td>
<td>1.81</td>
<td>0.156</td>
<td>0.0179</td>
<td>24.3</td>
</tr>
<tr>
<td>3 Sep 2008</td>
<td>3.08</td>
<td>1.51</td>
<td>0.130</td>
<td>0.00641</td>
<td>28.9</td>
</tr>
<tr>
<td>4 Sep 2008</td>
<td>2.47</td>
<td>1.02</td>
<td>0.109</td>
<td>0.00623</td>
<td>26.4</td>
</tr>
<tr>
<td>30 Jul to 25 Sep 2008**</td>
<td>4.70 (2.43)</td>
<td>1.97 (0.73)</td>
<td>NA***</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Basal Emission

<table>
<thead>
<tr>
<th>Previous Studies</th>
</tr>
</thead>
<tbody>
<tr>
<td>$E_{30}&gt;7$++</td>
</tr>
<tr>
<td>$\beta=0.159$--0.781*</td>
</tr>
</tbody>
</table>

$E_{30}=0.5,+$  & $E_{30}=0.070,+$  \\
$\beta=0.11$  & $\beta=0.15$  \\

* Bouvier-Brown et al. (2009)  
** Leaf cuvette-GC FID measured averages (for a total of 12 days in the periods).  
*** Mostly under the detection limit.  
+ Helmig et al. (2007)  
++ Harley et al. (1998)
**Table 4.** Major peaks in the ambient transmission corrected spectra and their normalized counts (Fig. 3).

<table>
<thead>
<tr>
<th>Species</th>
<th>m/z</th>
<th>Transmission Corrected Counts</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Day</td>
</tr>
<tr>
<td>Acetaldehyde</td>
<td>45</td>
<td>84.73</td>
</tr>
<tr>
<td>Formic Acid</td>
<td>47</td>
<td>251.75</td>
</tr>
<tr>
<td>Acetone</td>
<td>59</td>
<td>145.06</td>
</tr>
<tr>
<td>Acetic Acid</td>
<td>61</td>
<td>79.19</td>
</tr>
<tr>
<td>MBO</td>
<td>69 and 87</td>
<td>48.07</td>
</tr>
<tr>
<td>HMPR</td>
<td>71 and 89</td>
<td>34.20</td>
</tr>
<tr>
<td>Methyl Ethyl Ketone</td>
<td>73</td>
<td>24.64</td>
</tr>
<tr>
<td>MT</td>
<td>81 and 137</td>
<td>13.52</td>
</tr>
<tr>
<td>Oxygenated-MT</td>
<td>153</td>
<td>23.66</td>
</tr>
<tr>
<td>MS75</td>
<td>75</td>
<td>14.64</td>
</tr>
<tr>
<td>MS57**</td>
<td>57</td>
<td>31.46</td>
</tr>
<tr>
<td>MS101+</td>
<td>101</td>
<td>23.59</td>
</tr>
<tr>
<td>MS109</td>
<td>109</td>
<td>23.70</td>
</tr>
<tr>
<td>NO\textsubscript{y}</td>
<td></td>
<td>5.2 ppbv</td>
</tr>
<tr>
<td>SO\textsubscript{2}</td>
<td></td>
<td>0.36 ppbv</td>
</tr>
</tbody>
</table>

* Possibly hydroxy acetone.
** Possibly butene or fragment of hydroxy acetone.
+ Possibly isoprene hydroperoxides.
Table 5. Relative abundances, normalized by the acetone peak of the oxidation products of MBO by OH. The estimated product yields are presented in the brackets. The glycolaldehyde abundance and yield are calculated with considerations of the fragmented portion (~50%, see the text for details).

<table>
<thead>
<tr>
<th>Products</th>
<th>PTR-MS</th>
<th>FT-IR*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Formaldehyde</td>
<td>NA</td>
<td>0.29–0.35</td>
</tr>
<tr>
<td>Acetone</td>
<td>100 (0.51)</td>
<td>0.52–0.67</td>
</tr>
<tr>
<td>HMPR</td>
<td>23.7</td>
<td>0.19–0.31</td>
</tr>
<tr>
<td>glycolaldehyde</td>
<td>83.8 (0.43)</td>
<td>0.50–0.78</td>
</tr>
</tbody>
</table>

* Carrasco et al. (2007) and Atkinson and Arey (2003)
Fig. 1. (a) A averaged mass spectrum and (b) transmission-corrected mass spectrum of branch enclosure measurements (11 a.m. to 3 p.m. of 12 August to 14 August 2008).
Fig. 2. Averaged diurnal variations of BVOC and their oxidation products for three weeks in the summer of 2008 (18 August to 6 September).
Fig. 3. Representative ambient mass spectra of (a) the daytime (2 to 3 p.m. on 14 August 2008) and (b) the nighttime (1 to 2 a.m. on 18 August 2008) periods. Their transmission corrected mass spectra are presented (c) and (d), respectively.
Fig. 4. The product distribution mass spectrum of the OH initiated MBO oxidation experiment.
Fig. 5. The transmission corrected daytime ambient mass spectrum (red from Fig. 3c) and the steady-state product distribution spectrum (black) of MBO and β-pinene, calculated from the product distribution mass spectra of oxidation experiments.