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Received: 24 September 2015 – Accepted: 3 October 2015 – Published: 27 October 2015

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## Abstract

This paper reports the fluxes and mixing ratios of biogenically emitted volatile organic compounds (BVOCs) 4 m above a mixed oak and hornbeam forest in northern Italy. Fluxes of methanol, acetaldehyde, isoprene, methyl vinyl ketone + methacrolein, methyl ethyl ketone and monoterpenes were obtained using both a proton transfer reaction-mass spectrometer (PTR-MS) and a proton transfer reaction-time of flight-mass spectrometer (PTR-ToF-MS) together with the methods of virtual disjunct eddy covariance (PTR-MS) and eddy covariance (PTR-ToF-MS). Isoprene was the dominant emitted compound with a mean day-time flux of  $1.9 \text{ mg m}^{-2} \text{ h}^{-1}$ . Mixing ratios, recorded 4 m above the canopy, were dominated by methanol with a mean value of 6.2 ppbv over the 28 day measurement period. Comparison of isoprene fluxes calculated using the PTR-MS and PTR-ToF-MS showed very good agreement while comparison of the monoterpene fluxes suggested a slight over estimation of the flux by the PTR-MS. A basal isoprene emission rate for the forest of  $1.7 \text{ mg m}^{-2} \text{ h}^{-1}$  was calculated using the MEGAN isoprene emissions algorithms (Guenther et al., 2006). A detailed tree species distribution map for the site enabled the leaf-level emissions of isoprene and monoterpenes recorded using GC-MS to be scaled up to produce a “bottom-up” canopy-scale flux. This was compared with the “top-down” canopy-scale flux obtained by measurements. For monoterpenes, the two estimates were closely correlated and this correlation improved when the plant species composition in the individual flux footprint was taken into account. However, the bottom-up approach significantly underestimated the isoprene flux, compared with the top-down measurements, suggesting that the leaf-level measurements were not representative of actual emission rates.

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## 1 Introduction

The term volatile organic compound (VOC) describes a broad range of chemical species emitted from natural and anthropogenic sources into the atmosphere. VOCs emitted from the biosphere are commonly termed biogenic VOCs (BVOCs). Of the BVOCs isoprene is almost certainly the dominant species globally with an estimated annual emission of  $535\text{--}578 \times 10^{12}$  gC (Arneeth et al., 2008; Guenther et al., 2012). Isoprene, along with larger terpenoids, are the BVOCs that have received the most attention in the literature to date. Although isoprene is the most commonly measured BVOC global emission estimates continue to differ and there are still large uncertainties associated with the emission estimates of many other compounds. A better understanding of how emissions change with land cover, temperature, soil moisture and solar radiation is required to constrain model descriptions of the effects of BVOCs on atmospheric chemistry in the past, present and future (Monks et al., 2009).

BVOCs are a major source of reactive carbon into the atmosphere and as such exert an influence on both climate and local air quality. BVOCs are oxidised primarily by the hydroxyl radical (OH), itself formed by the photolysis of ozone, to form peroxide radicals ( $\text{RO}_2$ ). In the presence of  $\text{NO}_x$  ( $\text{NO}$  and  $\text{NO}_2$ ) these  $\text{RO}_2$  radicals can oxidise  $\text{NO}$  to  $\text{NO}_2$ , which may undergo photodissociation leading to the net formation of tropospheric ozone (Fehsenfeld et al., 1992). Tropospheric ozone can then impact upon human health, forest productivity and crop yields (Royal Society, 2008; Ashmore, 2005). In addition, BVOC species contribute significantly to the formation of secondary organic aerosol (SOA) in the atmosphere. This affects climate both directly and indirectly by the scattering of solar radiation and by acting as cloud condensation nuclei, increasing cloud cover and therefore altering the Earth's albedo (Hallquist et al., 2009).

The Bosco Fontana campaign was carried out as a part of the ÉCLAIRE (Effects of Climate Change on Air Pollution and Response Strategies for European Ecosystems) EC FP7 project to study the surface/atmosphere exchange within a semi-natural forest situated within one of the most polluted regions in Europe, and its interaction with air

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1 ppmv). The protonated mass of the VOCs ranged from  $m/z$  31 (formaldehyde,  $\text{CH}_3\text{O}^+$ ) to  $m/z$  181 (1,2,4-trichlorobenzene,  $\text{C}_6\text{H}_4\text{Cl}_3^+$ ). Methanol ( $m/z$  33), acetaldehyde ( $m/z$  45), acetone ( $m/z$  59), isoprene ( $m/z$  69), MEK ( $m/z$  73) and the monoterpene  $\alpha$ -pinene ( $m/z$  81 and  $m/z$  137) were present in the calibration gas standard, allowing sensitivities to be calculated directly. Due to reduced quadrupole transmission for high masses, monoterpenes were quantified using the fragment ion at  $m/z$  81. For compounds not contained in the gas standard (acetic acid ( $m/z$  61) and MVK and MACR ( $m/z$  71)) empirical sensitivities were calculated. A relative transmission curve was created using the instrumental sensitivities calculated from the masses present in the standard, and from this curve sensitivities for the unknown masses were calculated (Davison et al., 2009; Taipale et al., 2008). Error in calibration using the gas standard was assumed to be below 5 %, whereas relative errors in calibrations using the relative transmission approach are < 30 % (Taipale et al., 2008).

### 2.2.2 PTR-ToF-MS calibration

Background measurements of the PTR-ToF-MS were made up to three times a day using zero air generated by a custom made catalytic converter. Calibrations were made using a calibration gas (Appel Riemer Environmental Inc., USA) which contained 16 compounds, with masses ranging from 33 to 180 amu. For VOCs not included in the calibration standard, the average instrument sensitivities towards the known  $\text{C}_x\text{H}_y$ ,  $\text{C}_x\text{H}_y\text{O}_z$  or  $\text{C}_x\text{H}_y\text{N}_z$  compound families were used.

### 2.3 Calculation of volume mixing ratios

Volume mixing ratios were calculated from data generated using the PTR-MS using a program written in LabVIEW (National Instruments, Austin, Texas, USA). Volume mixing ratios ( $\chi_{\text{VOC}}$ ) were calculated from the raw PTR-MS data (in counts per second

(cps)) using a method based on those of Taipale et al. (2008) and Tani et al. (2004).

$$X_{\text{VOC}} = \frac{I(\text{RH}^+)_{\text{norm}}}{S_{\text{norm}}} \quad (1)$$

where  $S_{\text{norm}}$  is the normalised sensitivity and  $I(\text{RH}^+)_{\text{norm}}$  represents the background corrected normalised count rate (ncps) for the protonated compound  $R$  which was calculated as shown below.

$$I(\text{RH}^+)_{\text{norm}} = I(\text{RH}^+) \left( \frac{I_{\text{norm}}}{I(\text{H}_3\text{O}^+) + I(\text{H}_3\text{O}^+\text{H}_2\text{O})} \right) \left( \frac{\rho_{\text{norm}}}{\rho_{\text{drift}}} \right) - \frac{1}{n} \sum_{i=1}^n I(\text{RH}^+)_{\text{zero},i} \left( \frac{I_{\text{norm}}}{I(\text{H}_3\text{O}^+)_{\text{zero},i} + I(\text{H}_3\text{O}^+\text{H}_2\text{O})_{\text{zero},i}} \right) \left( \frac{\rho_{\text{norm}}}{\rho_{\text{drift, zero},i}} \right) \quad (2)$$

where  $I(\text{RH}^+)$ ,  $I(\text{H}_3\text{O}^+)$  and  $I(\text{H}_3\text{O}^+\text{H}_2\text{O})$  represent the observed count rate for the protonated compound  $R$ ,  $\text{H}_3\text{O}^+$  and the  $\text{H}_3\text{O}^+\text{H}_2\text{O}$  cluster, respectively. Subscript zero refers to zero air measurements,  $n$  is the number of zero air measurement cycles and  $\rho_{\text{drift}}$  is the drift tube pressure. The drift tube pressure was normalised to 2 mbar ( $\rho_{\text{norm}}$ ) and the sum of the primary ion and first water cluster was normalised to a count rate of  $10^6$  cps ( $I_{\text{norm}}$ ). The compound specific limit of detection (LoD) was calculated using the method described by Karl et al. (2003):

$$\text{LoD} = 2 \times \frac{\sigma_{\text{Background}}}{S_{\text{VOC}}} \quad (3)$$

where  $S_{\text{VOC}}$  is the instrumental sensitivity to the VOC and  $\sigma_{\text{Background}}$  is the mean background normalised count rate.

## 2.4 Flux calculations from PTR-MS

The 25 min PTR-MS flux files were inspected and incomplete or disrupted files removed. BVOC fluxes were then calculated using a program also written in LabVIEW,

based upon the virtual disjunct eddy covariance technique (vDEC) developed by Karl et al. (2002), also termed continuous flow disjunct eddy covariance (Rinne et al., 2008). This method has previously been successfully applied in a number of studies (e.g. Davison et al., 2009; Langford et al., 2009, 2010a, b; Misztal et al., 2011; Rinne et al., 2007).

This approach allows direct calculation of fluxes of atmospheric constituents, as with standard eddy covariance, yet in this case sampling of scalar concentrations is not continuous. The flux,  $F_x$ , for each compound was calculated using a covariance function between the vertical wind velocity,  $w$ , and the VOC mixing ratios,  $\chi$ :

$$F_x(\Delta t) = \frac{1}{N} \sum_{i=1}^N w'(i - \Delta t / \Delta t_w) \chi'(i) \quad (4)$$

where  $\Delta t$  is the lag time between the PTR-MS concentration measurements and the vertical wind velocity measurements from a sonic anemometer,  $\Delta t_w$  is the sampling interval between wind measurements (0.1 s),  $N$  is the number of PTR-MS measurement cycles in each 25 min averaging period (typically 250 in our study) and primes represent the momentary deviations from the mean concentration or vertical wind speed (e.g.  $w = w' - \bar{w}$ ).

Variations in temperature, pressure and the performance of the sample line pump can cause small deviations in  $\Delta t$ . Therefore these values were calculated using a cross correlation function between  $w'$  and  $\chi'$ . Lag times were calculated individually for each  $m/z$  monitored by the PTR-MS by selecting the absolute maximum value of the covariance function within a 30 s time window (MAX method, Taipale et al., 2010). This analysis resulted in a clear isoprene flux but for most masses a high proportion of the data fell below the limit of detection. These data, especially in the case of acetone, showed a significant amount of flux values with the opposite sign, “mirroring” the true flux. These “mirrored” points occur when the measured flux is of comparable magnitude to the total random error of the system (Langford et al., 2015). As the cross-correlation maximum is likely to be an over-estimate when the noise to signal ratio is greater than one, these points were substituted with fluxes calculated using a fixed lag time.

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The fluxes were filtered using the 70 % stationary criteria as presented by Foken and Wichura (1996), as was applied to the PTR-MS data and corrected for loss of high frequency flux Horst (1997). For a more detailed description of the flux calculation from the PTR-ToF-MS see Schallhart et al. (2015).

## 2.6 Leaf level GC-MS measurements

A portable gas exchange system equipped with a controlled-environment 6 cm<sup>2</sup> broadleaf cuvette (LI6400, Li-COR, Lincoln, USA) was used to measure net photosynthetic rate ( $A$ ) and stomatal conductance ( $g_s$ ) at basal conditions of PAR (1000  $\mu\text{mol m}^{-2} \text{s}^{-1}$ ), leaf temperature (30 °C) and a CO<sub>2</sub> concentration (400 ppm) from fully expanded leaves. When  $A$  reached a steady-state, the outlet tube from the leaf cuvette was replaced with a Teflon tube, and the air stream exiting from the cuvette was used as a sampling port for BVOC (according to the methodology in Loreto et al., 2001) by a silico-steel cartridge packed with 200 mg of tenax (Supelco, PA, USA). Tenax is a very hydrophobic and adsorbent material with high thermal stability generally used for trapping BVOC (Dettmer and Engewald, 2002). The flow rate through the leaf cuvette was maintained at 500  $\mu\text{mol s}^{-1}$ , and a subsample of 200 mL min<sup>-1</sup> (130  $\mu\text{mol s}^{-1}$ ) was pumped through the cartridge with an external pump (AP Buck pump VSS-1) for a total volume of 6 L of air. Blank samples of air without a leaf in the cuvette were collected every day before and after the BVOC samplings. Finally the cartridges were sealed and stored at 4 °C until analysis.

The cartridges were analyzed using a Perkin Elmer Clarus 580 gas-chromatograph coupled with a Clarus 560 Mass-Detector and a thermal-desorber Turbo Matrix (Perkin Elmer Inc., Waltham, MA, USA). The gas-chromatograph was equipped with an Elite-5-MS capillary column (30 m length, 250  $\mu\text{m}$  diameter and 0.25  $\mu\text{m}$  film thicknesses). The carrier gas was helium. The column oven temperature was kept at 40 °C for 5 min, then increased with a 5 °C min<sup>-1</sup> ramp to 250 °C and maintained at 250 °C for 5 min. BVOC were identified using the NIST library provided with the GC/MS Turbomass software. GC peak retention time was substantiated by analysis of parent ions and main

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fragments of the spectra. Commercially available reference standards were used to create the calibration curves and to quantify the emissions. To normalize the BVOC results, the quantities of terpenes collected from the empty cuvette (blanks) were subtracted from the plant emission results. The quantification of total BVOC emission was performed using authentic gaseous standards (Rivoira, Milan, Italy) or liquid standards (Sigma Aldrich, Milan, Italy).

## 2.7 Mapping tree species distribution

Tree species distribution data were obtained from Dalponte et al. (2007) who used a combination of Light Detection and Ranging (LIDAR) and hyperspectral data to develop a high resolution tree species distribution map of the Bosco Fontana natural reserve.

The overall accuracy (kappa coefficient) of this species map is particularly high (0.89), considering the number of classes (23) and the number of training samples (20 % of the data are used in the training set and 80 % in the test set) per class. The LIDAR channels provide relatively sparse information for discriminating between tree species, increasing the overall accuracy of the tree species assignment using the hyperspectral data by only 1 % but the LIDAR data significantly increase the accuracy of understory and underrepresented classes. The kappa coefficient of the main species is also very high (0.88–0.93) showing the effectiveness of this approach for species classification in a very complex forest with 20 different broad-leaves species, some of which, such as *Q. cerris*, *Q. robur* and *Q. rubra*, belong to same genus. For a more detailed discussion of the mapping results and methodology see Dalponte et al. (2007, 2008).

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quency flux measurements so the presented mixing ratios are an average over 25 min. The mixing ratio LoDs, calculated as described above (Karl et al., 2003; Langford et al., 2009; Misztal et al., 2011) were in the same range as those calculated on previous campaigns (Langford et al., 2009; Misztal et al., 2011) and, with the exception of isoprene where the mixing ratio dropped towards zero at night, the recorded mixing ratios generally remained above their respective LoD.

Table 3 summarizes the flux data recorded during the Bosco Fontana measurement campaign. Wind speeds decreased at night, leading to a large proportion of the night time data falling below the  $u_*$  threshold of  $0.15 \text{ ms}^{-1}$ . Consequently, average emission fluxes of all eight compounds are reported for the daytime period 10:00–15:00 LT as well as for the whole campaign. Large fluxes of  $m/z$  69 and  $m/z$  81 (assigned to isoprene and monoterpenes respectively) were observed and are shown in Fig. 4. Fluxes of  $m/z$  33, 45, 59, 61, 71 and 73 (assigned to methanol, acetaldehyde, acetone, acetic acid, MVK + MACR and MEK, respectively) were also observed, but these fluxes were weaker, leading to a high percentage of fluxes failing the LoD check. However, as is described by Langford et al. (2015), when these flux data are averaged to show the average diurnal cycle, it is appropriate to use a combined LoD value appropriate for the same period rather than the LoD attached specifically to each 25 min flux file. It is, though, essential that each individual flux period be processed carefully to avoid the introduction of a bias due to the use of the MAX method of time-lag identification. The LoD for the mean ( $\overline{\text{LoD}}$ ) decreases with the square root of the number of samples averaged ( $N$ ).

$$\overline{\text{LoD}} = \frac{1}{N} \sqrt{\sum_{i=1}^N \text{LoD}^2} \quad (5)$$

Therefore, while the flux time series of methanol, acetaldehyde, acetone, acetic acid, MVK + MACR and MEK are not presented here, the campaign average diurnal fluxes are shown (Fig. 5). As discussed above, 25 min averaged flux files flagged as below the

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LoD were included in these diurnal averages. Flux files falling below the  $0.15 \text{ m s}^{-1}$  wind speed threshold were also included to prevent the night time flux being biased high for depositing compounds. For compounds showing emission, night-time fluxes are close to zero anyway and the application has little influence on the results. Data flagged for non-stationarity were excluded. For a more detailed discussion of the fluxes and mixing ratios of each BVOC and comparison made with other temperate and Mediterranean ecosystems, see the Supplement.

The fluxes of isoprene and monoterpenes calculated using both the PTR-MS and the PTR-ToF-MS instruments are displayed in Fig. 4 and summarised in Table 3. The isoprene fluxes calculated using both instruments show very good agreement ( $R^2 = 0.91$ , slope 1.3 and intercept 0.17). The monoterpene fluxes, calculated using  $m/z$  81 with the PTR-MS and  $m/z$  81.070 with the PTR-ToF-MS show an  $R^2 = 0.50$ . Three additional mass spectral peaks are observed at  $m/z$  81 in the PTR-ToF-MS:  $m/z$  80.92, 80.99 and 81.03, however statistically significant fluxes from these peaks could not be calculated using the PTR-ToF-MS. Owing to the lower sensitivity of the PTR-MS at  $m/z$  81 and the lower sampling frequency of the disjunct sampling protocol (Rinne and Ammann, 2012), the monoterpene flux calculated using this instrument is significantly noisier than the flux calculated using the PTR-ToF-MS.

PTR-MS and PTR-ToF-MS mass scans were averaged over a ten day period (14–24 June). A comparison of these mass scans over the range  $m/z$  33 to 100 at unit mass resolution is displayed in Fig. 6, with masses reported relative to  $m/z$  59 (acetone). A good agreement between the PTR-MS and PTR-ToF-MS is seen for all masses, except for  $m/z$  33 where the PTR-MS gives a significantly higher signal. As both instruments have comparable sensitivities at this mass (11.6 and ca. 10–12 ncps ppbv<sup>-1</sup> for the PTR-MS and PTR-ToF-MS respectively) this discrepancy must be the result of interference from another ion at this mass.  $\text{O}^{17}\text{O}^+$  could interfere with the methanol signal at  $m/z$  33 but as a significant peak is not observed at  $m/z$  34 ( $\text{O}^{18}\text{O}^+$ ) a large contribution from  $\text{O}^{17}\text{O}^+$  to  $m/z$  33 is unlikely. This suggests that there is a greater formation of  $\text{O}_2\text{H}^+$  in the PTR-MS than in the PTR-ToF-MS under these particular op-

eration parameters. No major mass spectral peaks are observed in one instrument alone, indicating that there is no artefact formation or unexpected loss of chemical species with either instrument. The mass scans show a much cleaner spectrum than was reported by Misztal et al. (2011) above an oil palm plantation in South-East Asia, suggesting an atmosphere dominated by fewer chemical species at higher concentrations.

### 3.2.1 BVOC correlations

Scatter plots were used to investigate the relationship between the measured species. Methanol, acetone and MEK (Fig. 7) all showed a bimodal relationship with two linear groupings, one at lower temperature (ca.  $< 20^{\circ}\text{C}$ ) and another at higher temperatures (ca.  $> 20^{\circ}\text{C}$ ). This suggests that either there are two different sources contributing to the mixing ratios (most likely an atmospheric background and a photochemical source at higher temperatures) or that a second compound contributes to the nominal mass at higher temperatures. As few compounds have been reported to contribute to  $m/z$  33 or 59, an additional source at higher temperatures seems more likely.

### 3.2.2 Short-chain oxygenated BVOCs

A mean methanol mixing ratio of 6.2 ppbv at 4 m above the canopy was recorded over the duration of the campaign, making it the dominant BVOC observed at Bosco Fontana. Mean acetaldehyde, acetone and acetic acid mixing ratios were 3.4, 3.2 and 1.9 ppbv at 4 m above the canopy, respectively. Methanol, acetaldehyde and acetic acid mixing ratios all followed similar diurnal cycles (Fig. 3), with mixing ratios remaining stable through the night before a drop in the morning, probably caused by expansion of the planetary boundary layer after sunrise. Then mixing ratios increased again in the late afternoon as emissions accumulated in a shrinking boundary layer. Acetone mixing ratios remained on average stable throughout the day (Fig. 3).

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that emissions of MVK and MACR increase with temperature stress (Jardine et al., 2012). The mid-day (10:00–15:00 LT) mixing ratios of MVK + MACR at 4 m above the canopy showed a positive correlation with those of isoprene ( $R^2 = 0.49$ ), suggesting that the oxidation of isoprene was responsible for the formation of MVK and MACR.

The production of MVK and MACR from isoprene at the Bosco Fontana site has been modelled by Schallhart et al. (2015), who estimated that 4–27 % of the MVK + MACR flux was formed from isoprene oxidation products. MVK and MACR mixing ratios recorded at 4 m above the canopy (Fig. 3) increase in the morning as isoprene concentrations rise, before boundary layer expansion causes them to drop in the middle of the day. The mixing ratios then increase again in the evening as the boundary layer contracts. The flux of MVK + MACR (Fig. 5) peaked in the early afternoon with a mean day-time flux of  $0.05 \text{ mg m}^{-2} \text{ h}^{-1}$  comparable to the 0.03 and  $0.08 \text{ mg m}^{-2} \text{ h}^{-1}$  observed, respectively, by Kalogridis et al. (2014) and Spirig et al. (2005) over European oak and mixed forests.

MEK may be directly emitted by plants (Fall, 2003) or formed photochemically (Luecken et al., 2012). MEK mixing ratios 4 m above the forest canopy remained stable through the night at ca. 0.6 ppbv before a dropping in the morning, probably caused by expansion of the planetary boundary layer, to ca. 0.3 ppbv and rising again in the evening (Fig. 3). A plot of the mixing ratios of MEK against those of acetone reveals a bimodal distribution suggesting two distinct sinks or sources (Fig. 7), the first occurring at lower temperatures (ca. 12–20 °C) with a MEK to acetone ratio of ca. 0.17 and the second at higher temperatures (ca. 20–34 °C) with a MEK to acetone ratio of ca. 0.06. A relationship between acetone and MEK has been reported by Riemer et al. (1998) who observed an MEK to acetone ratio of 0.07 at temperatures between 20 and 37 °C. This compares well with the observations at Bosco Fontana. This trend was not observed when data were coloured by PAR indicating that the bimodal distribution is not driven by the faster rate of reaction of MEK than of acetone with OH. A low MEK emission flux was observed in the afternoon with a mean day-time flux of  $0.02 \text{ mg m}^{-2} \text{ h}^{-1}$ .

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### 3.2.4 Isoprene and monoterpenes

Isoprene mixing ratios 4 m above the canopy began to rise in the mid-morning from a night-time zero, peaking in the late afternoon at ca. 2 ppbv before falling again to zero in the late evening (Fig. 3). Isoprene fluxes were not observed at night, but increased in the morning to a peak in the mid afternoon before dropping to zero again in the evening (Fig. 5) with a mean day-time flux of  $1.9 \text{ mg m}^{-2} \text{ h}^{-1}$ .

Isoprene fluxes correlated with leaf temperature (estimated using a method based on that described by Nemitz et al. (2009) and explained in more detail in the Supplement) giving an  $R^2 = 0.75$  for an exponential fit, PAR ( $R^2 = 0.75$  for an exponential fit) and with sensible heat flux ( $H$ ) ( $R^2 = 0.67$ ). The relationship between isoprene fluxes and mixing ratios, temperature and PAR is displayed in Fig. 8. An exponential relationship between temperature and both fluxes and mixing ratios was observed for the periods when PAR was greater than zero. Table 4 compares isoprene flux measurements with the fluxes recorded during other field campaigns in the Mediterranean region and the isoprene emission factor under basal conditions. As would be expected the flux of isoprene is shown to be highly dependent on ecosystem type. When fluxes are normalised to standard conditions the fluxes observed on this campaign are lower than those observed over woodland dominated by isoprene emitting oak species due to the lower proportion of isoprene emitting species in the canopy but closer in magnitude to that observed over a mixed pine and oak forest.

The campaign mean monoterpene mixing ratio 4 m above the canopy was 0.2 ppbv. The diurnal profile (Fig. 3) shows a night-time mixing ratio of ca. 0.18 ppbv which increases to ca. 0.21 ppbv in the morning remaining stable through the day and dropping again to ca. 0.18 ppbv at night. The monoterpene flux (Fig. 5) peaked in the early afternoon with a campaign mean mid-day flux of  $0.12 \text{ mg m}^{-2} \text{ h}^{-1}$ . Monoterpene mixing ratios were not significantly correlated with leaf surface temperature or with PAR ( $R^2 = 0.11$  and  $0.12$  respectively). However, the flux displayed a correlation with both leaf surface temperature and PAR ( $R^2 = 0.44$  and  $0.39$  respectively).

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### 3.3 Calculation of isoprene and monoterpene canopy level emission factors

Although other approaches do exist, isoprene fluxes are widely modelled using the Model of Emissions of Gases and Aerosols from Nature (MEGAN, Guenther et al., 2006). MEGAN calculates isoprene fluxes based on the product of an emission activity factor ( $\gamma$ ), a canopy loss and production factor ( $\rho$ ) and a canopy emission factor ( $\varepsilon$ ). Therefore, plotting isoprene flux against  $\gamma \times \rho$  enables the calculation of a canopy-specific isoprene emission factor (Fig. 9), giving a canopy emission factor of  $1.68 \text{ mg m}^{-2} \text{ h}^{-1}$  at standard conditions ( $1000 \mu\text{mol m}^{-2} \text{ s}^{-1}$  PAR and 303 K) for the campaign period. For the purpose of this work,  $\gamma$  was calculated using the algorithms described by Guenther et al. (2006). Radiative transfer through the canopy was modelled using the canopy model applied by Müller et al. (2008) based on that of Goudriaan and van Laar (1994) and ambient temperature was recorded 4 m above the canopy. The standard light and temperature conditions for MEGAN canopy scale emissions factors are  $\sim 1500 \mu\text{mol m}^{-2} \text{ s}^{-1}$  and 303 K (Guenther et al., 2006). In order to allow for direct comparison with the GC-MS data and literature emissions factors the factor which sets the emission activity to unity at standard conditions ( $C_{\text{CE}}$ ) was increased to 1.42 to give standard light and temperature conditions of  $1000 \mu\text{mol m}^{-2} \text{ s}^{-1}$  and 303 K, respectively.

The emission factor is lower than those calculated by Kalogridis et al. (2014) and Baghi et al. (2012) from oak (*Quercus pubescens*) dominated forests in southern France ( $7.4$  and  $5.4 \text{ mg m}^{-2} \text{ h}^{-1}$ , respectively). However, this is to be expected, owing to the high proportion of low or non-isoprene emitting species such as *Carpinus betulus*, *Corylus avellana*, *Sambucus nigra* and *Acer campestre* present in the forest at Bosco Fontana.

Monoterpene emission from plants may take the form of pool or de novo emission. Emission from stored pools is temperature controlled whereas de novo is driven by photosynthesis and is therefore controlled by light as well as temperature (Ghirardo et al., 2010). Emission from stored pools was modelled using the monoterpene–temperature

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creased in the understory vegetation but it is unlikely that, despite their high rates of growth, the *Populus* coverage changed significantly in the 4 years between mapping and this campaign. Whilst the hyperspectral/LIDAR tree species data for this site provides a unique opportunity for comparing the canopy-scale measurements with a detailed bottom-up estimate, the hyperspectral/LIDAR data provides information on projected tree species area as seen from above, whilst the flux is regulated by leaf mass and its exposure to radiation. Thus there are uncertainties in the ability of the hyperspectral/LIDAR in detecting understorey vegetation and a single conversion factor was used between projected tree area and leaf mass. However, understorey vegetation is less exposed to sunlight reducing its emission. Indeed, the main reason for the underestimate of isoprene flux is probably that the leaf level isoprene emission rate recorded from the leaves sampled at ground level (albeit taken at the edge of sun exposed clearings) are not representative of those at the canopy top. Substituting the measured *Q. robur* and *Q. rubra* emission factors with those reported by Karl et al. (2009) caused the bottom-up estimate to give 130 % of the measured flux and improved the correlation between bottom-up estimates and canopy-scale measurements further.

The speciated monoterpene flux also showed good agreement with the above canopy flux ( $R^2 = 0.72$ ) and captured 57 % of the flux. The discrepancy between the magnitude of the speciated monoterpene flux and the above canopy flux was within the range expected to be caused by the loss of monoterpenes within the canopy through oxidation and deposition.

The contribution of different species to the isoprene and monoterpene fluxes over the course of an example day is shown in Fig. 10. As is shown, the isoprene flux was dominated by *Q. robur* but was sensitive to the species composition within the flux footprint. The change in wind direction around 14:00 LT reduced the contribution of *Q. rubra* to the total flux, with the contribution of *Populus × canescens* increasing significantly. The monoterpene flux was predicted to have been dominated by *C. betulus*, the dominant tree species in the canopy at Bosco Fontana. A greater number of tree

species contributed to the monoterpene flux, and emissions were therefore much more uniform across the canopy and less affected by changes in wind direction.

The fit between the above canopy measured isoprene and monoterpene fluxes and the “bottom-up” flux estimate was improved by optimising the leaf-level emission factors, within the constraints displayed in Table 6, using  $\text{Chi}^2$  minimisation as implemented by the solver function in Microsoft Excel. Use of the optimised isoprene and monoterpene emission factors gave a good correlation with the measured fluxes with  $R^2$  values of 0.75 and 0.76, respectively for isoprene and monoterpenes. The optimised isoprene and monoterpene emission factors are presented in Table 6 and show a reasonable agreement with literature values (Karl et al., 2009).

## 4 Conclusions

Direct above-canopy fluxes of methanol, acetaldehyde, acetic acid, isoprene, MVK + MACR, MEK and monoterpenes were calculated using the method of virtual disjunct eddy covariance from mixing ratio data obtained with a PTR-MS above a semi-natural mixed oak and hornbeam forest in northern Italy from 13 June to 11 July 2012. Isoprene was the dominant BVOC emitted with a mean day-time flux of  $1.91 \text{ mg m}^{-2} \text{ h}^{-1}$ . When normalised to standard conditions (temperature of  $30^\circ\text{C}$ , PAR of  $1000 \mu\text{mol m}^{-2} \text{ s}^{-1}$ ) using the MEGAN model (Guenther et al., 2006), a canopy scale emission factor of  $1.68 \text{ mg m}^{-2} \text{ h}^{-1}$  was derived. Mixing ratios of VOCs measured at 4 m above the forest canopy were dominated by those of methanol, with a campaign mean mixing ratio of 6.2 ppbv.

The isoprene fluxes obtained using the PTR-MS/vDEC system showed good agreement with those obtained using a direct eddy covariance (with volume mixing ratios measured with a fast response PTR-ToF-MS instrument). Monoterpene fluxes recorded using the PTR-MS were noisier and marginally higher than those recorded using the PTR-ToF-MS due to a lower sensitivity and, probably, the inclusion of isobaric compounds. Comparison of mass scan data generated using the PTR-MS and

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PTR-ToF-MS showed very good agreement and no significant masses observed in one instrument but not in the other within the mass range  $m/z$  33–100.

Up-scaling leaf-level isoprene and monoterpene emissions to the canopy scale, using a high spatial resolution tree species database and a 2-D footprint model, showed significantly better correlation with the measured above canopy fluxes than was obtained using a canopy scale emission factor. Leaf-level isoprene emissions resulted in an underestimate of the above-canopy isoprene flux and this was assumed to be the result of differences in isoprene emission rates from leaves sampled at ground-level and those at the canopy top.

Overall, the data obtained give confidence in the measurement of biogenic VOC fluxes by the method of virtual disjunct eddy covariance and highlight the importance of using leaf-level emissions data from sun-lit canopy-top leaves when up-scaling leaf-level emissions to produce a “bottom-up” canopy-scale emissions estimate.

**The Supplement related to this article is available online at  
doi:10.5194/acpd-15-29213-2015-supplement.**

*Acknowledgements.* W. J. F. Acton would like to thank Alex Guenther for his advice on the use of MEGAN. This work was funded by the EU FP7 grants ÉCLAIRE (grant 282910) and PEGASOS (grant 265148), as well as by a BBSRC/Ionicon Analytik GmbH Industrial CASE studentship awarded to W. J. F. Acton. We acknowledge access to the measurement site provided by the Italian Corpo forestale dello Stato and provision of the site infrastructure by the Catholic University of Italy at Brescia and in particular by Giacomo Gerosa, Angelo Finco and Riccardo Marzuoli.

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**Table 5.** Leaf level isoprene and monoterpene emission ( $\text{mg m}^{-2} \text{h}^{-1}$ ) from single leaves under basal conditions (temperature:  $30^\circ\text{C}$  and PAR:  $1000 \mu\text{mol m}^{-2} \text{s}^{-1}$ ). ND signifies not detected.

Tree species	isoprene flux (standard error)	$\alpha$ -pinene flux (standard error)	sabinene flux (standard error)	$\beta$ -pinene flux (standard error)	limonene flux (standard error)	sum monoterpene flux
<i>Carpinus betulus</i>	$2.25 \times 10^{-3}$ ( $1.50 \times 10^{-3}$ )	$1.07 \times 10^{-2}$ ( $6.00 \times 10^{-3}$ )	$1.81 \times 10^{-2}$ ( $1.36 \times 10^{-2}$ )	$5.14 \times 10^{-2}$ ( $1.23 \times 10^{-2}$ )	$5.83 \times 10^{-1}$ ( $2.36 \times 10^{-1}$ )	$6.63 \times 10^{-1}$
<i>Quercus robur</i>	$2.39 \times 10^0$ ( $6.12 \times 10^{-1}$ )	$2.81 \times 10^{-2}$ ( $1.45 \times 10^{-2}$ )	ND	$4.70 \times 10^{-3}$ ( $3.08 \times 10^{-3}$ )	$2.16 \times 10^{-1}$ ( $6.49 \times 10^{-2}$ )	$2.49 \times 10^{-1}$
<i>Quercus rubra</i>	$9.14 \times 10^{-1}$ ( $2.02 \times 10^{-1}$ )	ND	ND	$7.95 \times 10^{-3}$ ( $2.22 \times 10^{-3}$ )	$2.34 \times 10^{-2}$ ( $7.11 \times 10^{-3}$ )	$3.13 \times 10^{-2}$
<i>Corylus avellana</i>	$4.97 \times 10^{-4}$ ( $3.93 \times 10^{-4}$ )	$1.30 \times 10^{-2}$ ( $8.00 \times 10^{-3}$ )	ND	$2.08 \times 10^{-2}$ ( $4.80 \times 10^{-3}$ )	$7.57 \times 10^{-1}$ ( $4.15 \times 10^{-1}$ )	$7.90 \times 10^{-1}$
<i>Acer campestre</i>	$4.40 \times 10^{-4}$ ( $3.11 \times 10^{-4}$ )	$5.14 \times 10^{-2}$ ( $2.95 \times 10^{-2}$ )	ND	$2.27 \times 10^{-1}$ ( $3.54 \times 10^{-2}$ )	$1.07 \times 10^{-1}$ ( $1.41 \times 10^{-2}$ )	$3.85 \times 10^{-1}$
<i>Sambucus nigra</i>	$4.09 \times 10^{-3}$ ( $3.66 \times 10^{-3}$ )	ND	ND	$9.67 \times 10^{-3}$ ( $2.69 \times 10^{-3}$ )	$2.49 \times 10^{-1}$ ( $1.41 \times 10^{-1}$ )	$2.59 \times 10^{-1}$
<i>Cornus sanguinea</i>	$4.00 \times 10^{-1}$ ( $4.00 \times 10^{-1}$ )	$1.11 \times 10^{-3}$ ( $1.11 \times 10^{-3}$ )	ND	$1.95 \times 10^{-2}$ ( $4.91 \times 10^{-3}$ )	$2.28 \times 10^{-1}$ ( $1.73 \times 10^{-1}$ )	$2.49 \times 10^{-1}$

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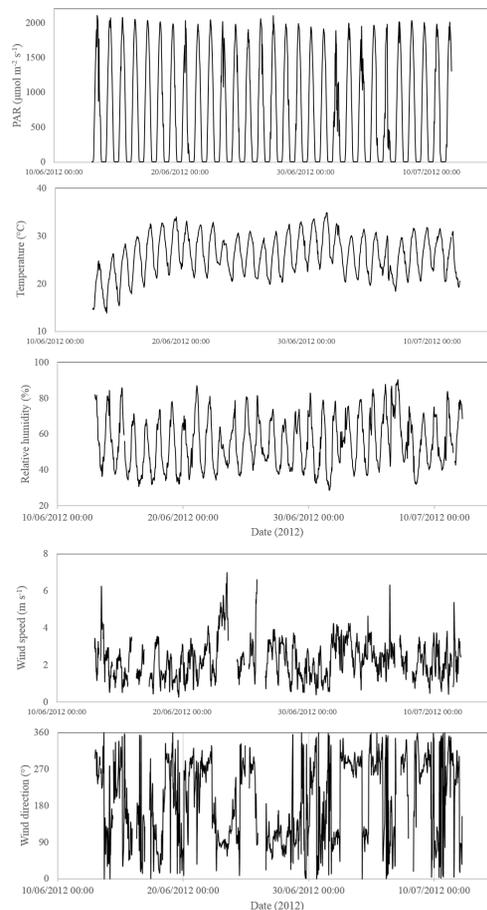


**Table 6.** Species specific isoprene and monoterpene emission factors (for a standard temperature of 30 °C and a PAR value of 1000  $\mu\text{mol m}^{-2} \text{s}^{-1}$ ) derived from optimising the leaf level emission factors to give the best fit with the measured above canopy isoprene and monoterpene fluxes within the constraints displayed.

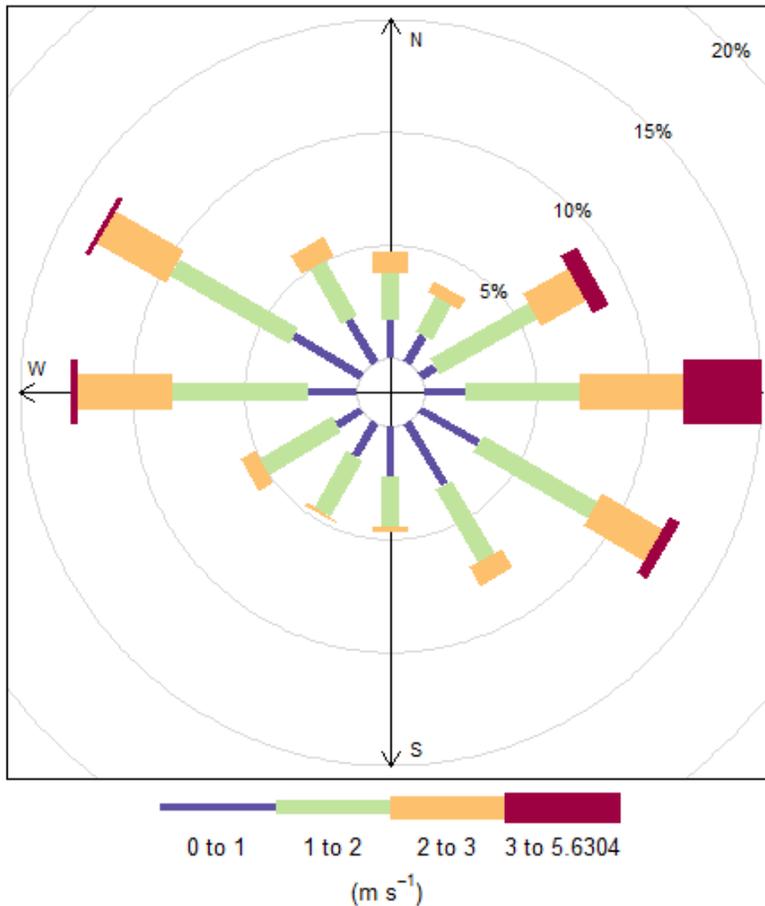
Species	Isoprene emission factor ( $\text{mg m}^{-2} \text{h}^{-1}$ )	Isoprene constraint ( $\text{mg m}^{-2} \text{h}^{-1}$ )	Monoterpene emission factor ( $\text{mg m}^{-2} \text{h}^{-1}$ )	Monoterpene constraint ( $\text{mg m}^{-2} \text{h}^{-1}$ )
<i>Acer campestre</i>	0.00	< 1.0	0.15	< 0.50
<i>Acer negundo</i>	0.00	< 1.0	0.33	< 0.64
<i>Alnus glutinosa</i>	0.01	< 1.0	0.22	< 0.50
<i>Carpinus betulus</i>	0.00	< 1.0	0.57	< 0.63
<i>Corylus avellana</i>	0.00	< 1.0	0.23	< 0.50
<i>Fraxinus angustifolia</i>	0.00	< 1.0	0.00	< 0.50
<i>Juglans nigra</i>	0.00	< 1.0	0.12	< 0.50
<i>Juglans regia</i>	0.36	< 1.0	0.15	< 0.50
<i>Morus</i> sp.	0.00	< 1.0	0.19	< 0.50
<i>Platanus hispanica</i>	2.97	< 4.4	0.50	< 0.50
<i>Populus</i> × <i>canescens</i>	10.66	< 16.1	0.29	< 0.50
<i>Populus</i> × <i>hybrida</i>	8.06	< 16.1	0.00	< 0.50
<i>Prunus avium</i>	0.00	< 1.0	0.01	< 0.50
<i>Quercus cerris</i>	0.02	< 1.0	0.07	< 0.50
<i>Quercus robur</i>	7.46	< 16.1	0.19	< 0.50
<i>Quercus rubra</i>	1.38	< 8.1	0.02	< 0.50
<i>Robinia pseudoacacia</i>	1.38	< 2.8	0.01	< 0.50
<i>Rubus</i> sp.	0.00	< 1.0	0.01	< 0.50
<i>Tilia</i> sp.	0.00	< 1.0	0.00	< 0.50
<i>Ulmus minor</i>	0.01	< 1.0	0.01	< 0.50
Grass	0.06	< 1.0	0.06	< 0.15
Not woodland	0.06	< 1.0	0.08	< 0.15
Outside forest	0.06	< 1.0	0.06	< 0.50

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**Figure 1.** Time series of meteorological conditions recorded over the campaign period. From top to bottom: PAR ( $\mu\text{mol m}^{-2} \text{s}^{-1}$ ), air temperature ( $^{\circ}\text{C}$ ), relative humidity (%), wind speed ( $\text{m s}^{-1}$ ) and wind direction ( $^{\circ}$ ).



**Figure 2.** Wind direction and wind speed observed during the Bosco Fontana field campaign.

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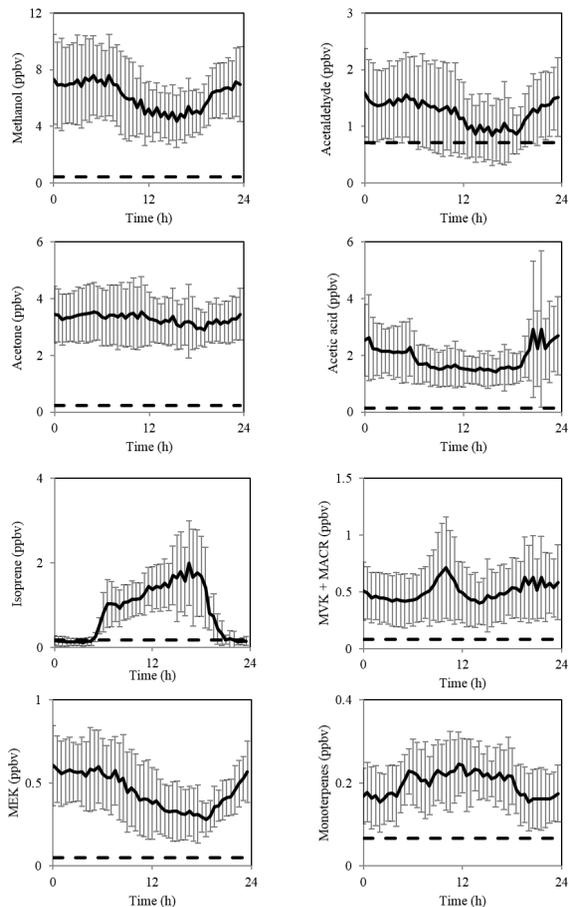
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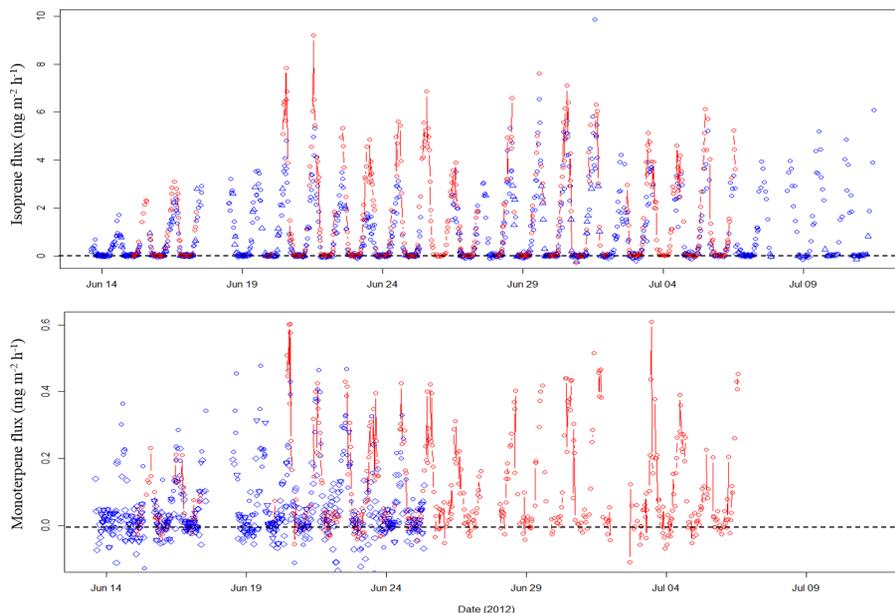




**Figure 3.** Mean 4 m above-canopy diurnal volume mixing ratios of volatile organic compounds measured during the Bosco Fontana field campaign. Error bars represent one standard deviation from the mean and the dashed line denotes limit of detection.

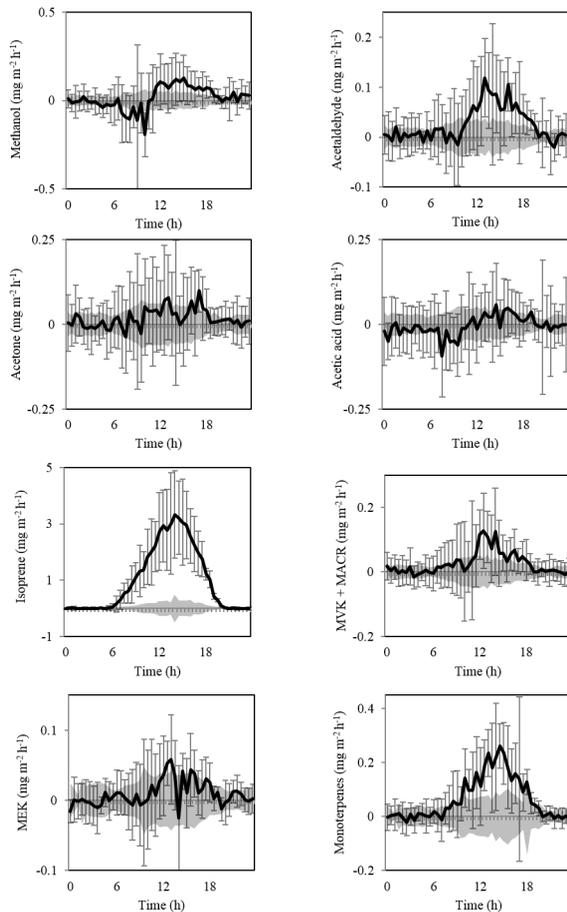
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**Figure 4.** Time series of isoprene (top) and monoterpene (bottom) fluxes ( $\text{mg m}^{-2} \text{h}^{-1}$ ) measured using the method of vDEC. Blue circles, triangles and diamonds represent 25 min averaged flux data from the PTR-MS which respectively passed all tests, fell below the  $u_*$  threshold or the LoD. Red circles and lines represent PTR-ToF-MS isoprene and monoterpene fluxes with 30 min averaged flux files failing the stationarity test removed.

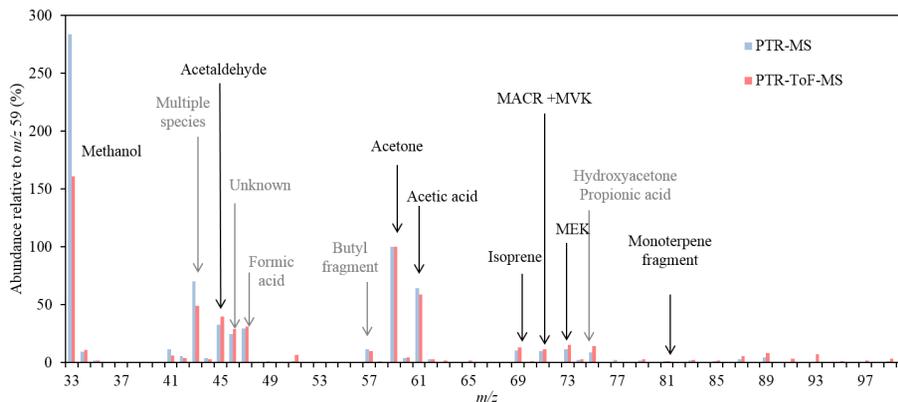
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**Figure 5.** Mean diurnal fluxes of volatile organic compounds measured using vDEC. Shaded area represents the limit of detection of the averaged data, and error bars represent one standard deviation between days from the mean.

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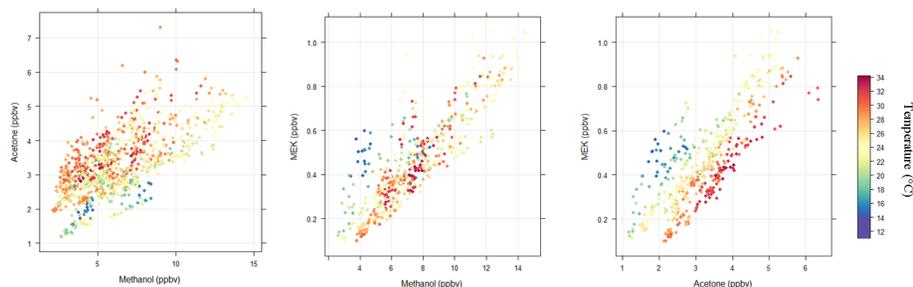


**Figure 6.** Comparison of PTR-MS (blue) and PTR-ToF-MS (red) mass scans relative to  $m/z$  59 at unit mass resolution averaged between 14 and 24 June. Compounds recorded in flux mode using the PTR-MS are presented in black with compounds tentatively identified in grey.

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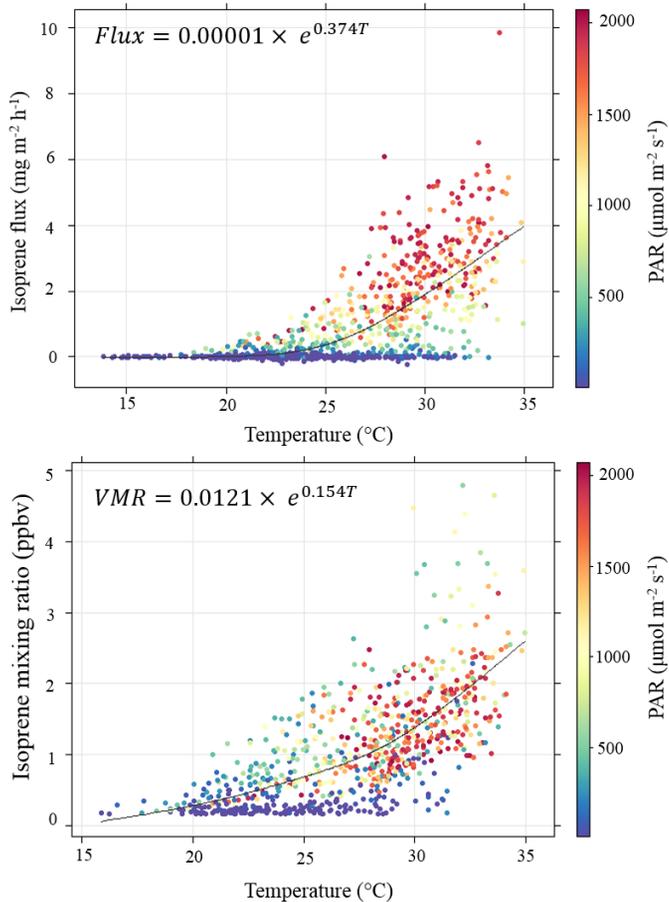
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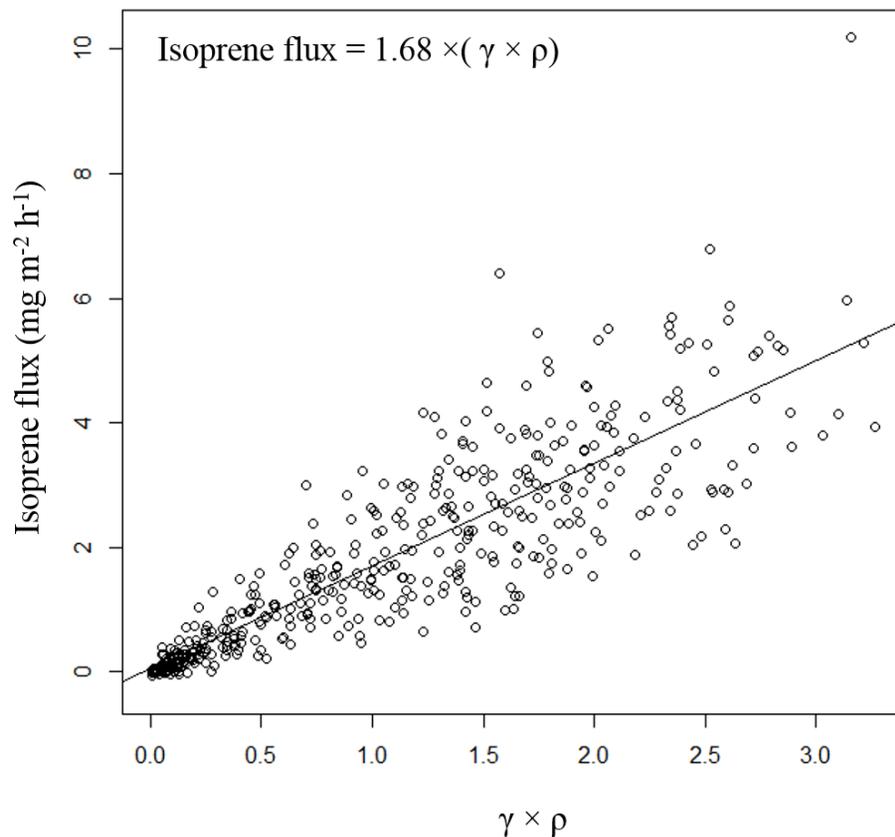


**Figure 7.** Scatter plots displaying the relationship between the volume mixing ratios of methanol, acetone and MEK measured 4 m above the canopy, coloured by temperature.

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**Figure 8.** The exponential relationship between temperature (°C) and isoprene fluxes ( $\text{mg m}^{-2} \text{h}^{-1}$ ) and volume mixing ratios (ppbv), coloured according to the magnitude of photosynthetically active radiation ( $\mu\text{mol m}^{-1} \text{s}^{-1}$ ).



**Figure 9.** Measured isoprene fluxes against the product of  $\gamma$  (emission activity factor, itself the product of the temperature, light and leaf area index activity factors) and  $\rho$  (the canopy loss and production factor).

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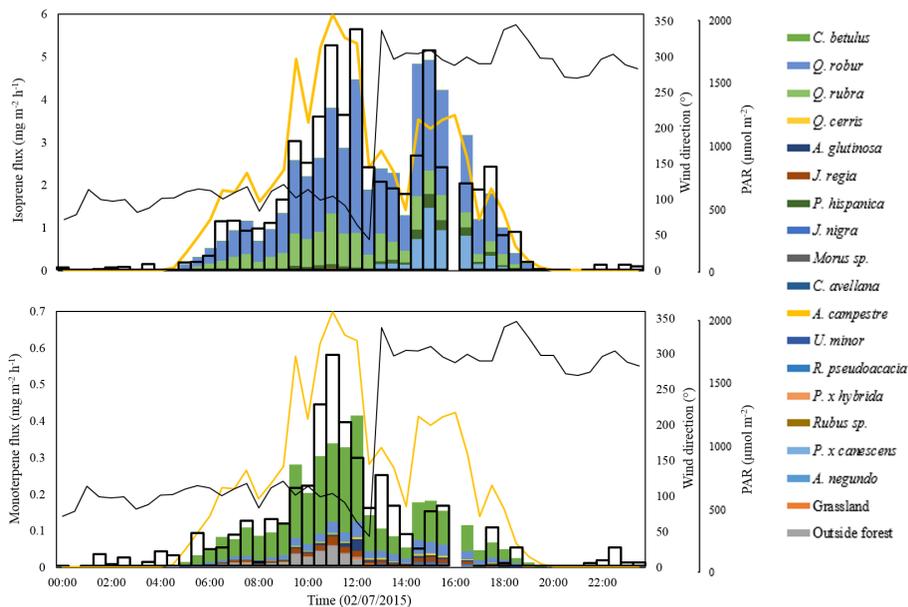
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**Figure 10.** The contribution of individual tree species to the speciated isoprene and monoterpene flux on the 2 July 2012 with PAR displayed as a yellow line, wind direction as a black line and the flux recorded using the PTR-MS as bold black bars.

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