Trends in particle phase liquid water during the Southern Oxidant and Aerosol Study

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Abstract

We present in situ measurements of particle phase liquid water. Measurements were conducted 3 June to 15 July 2013 during the Southern Oxidant and Aerosol Study in the southeastern U.S. The region is dominated by biogenic emissions, impacted by anthropogenic pollution, photochemically active, humid, and known to contain high concentrations of organic aerosol mass. Measurements characterized mobility number size distributions of ambient atmospheric aerosols in three states: unperturbed, dry, and dry-humidified. Unperturbed measurements describe the aerosol distribution at ambient temperature and relative humidity. For the dry state, the sample was routed through a cold trap upstream of the inlet then re-heated, while for the dry-humidified state the sample was re-humidified after drying. The total volume of water and semi-volatile compounds lost during drying was quantified by differencing dry and unperturbed volumes from the integrated size spectra, while semi-volatile volumes lost during drying were quantified differencing unperturbed and dry-humidified volumes. Results indicate that particle phase liquid water was always present. Throughout the SOAS campaign, median water mass concentrations at the relative humidity (RH) encountered in the instrument typically ranged from 1 to 5 µg m\(^{-3}\), but were as high as 73 µg m\(^{-3}\). On non-raining days, morning time (6-9am) median mass concentrations exceeded 15 µg m\(^{-3}\). Hygroscopic growth factors followed a diel cycle and exceed two from 7 to 9 a.m. local time. The hygroscopicity parameter kappa ranged from 0.14 to 0.46 and hygroscopicity increased with increasing particle size. An observed diel cycle in kappa is consistent with changes in aerosol inorganic content and a dependency of the hygroscopicity parameter on water content. Unperturbed and dry-humidified aerosol volumes did not result in statistically discernible differences, demonstrating that drying did not lead to large losses in dry particle volume. We anticipate that our results will help improve the representation of aerosol water content and aqueous phase mediated partitioning of atmospheric water-soluble gases in photochemical models.
1 Introduction

Atmospheric aerosols affect human health and welfare, global climate, visibility, and ecosystems. Aerosols are either directly emitted or form in the atmosphere through a myriad of chemical reactions involving a variety of anthropogenic and biogenic precursors (Kanakidou et al., 2005; Hallquist et al., 2009). As a consequence, atmospheric aerosols consist of a mix of chemically diverse lower and higher volatility compounds. While low-volatility species remain predominantly in the particle phase, semi-volatile compounds, including water, undergo equilibrium partitioning between the condensed and gaseous phases (Murphy et al., 1998; Pöschl, 2005; Robinson et al., 2007). Particle phase liquid water and its interactions with aerosol chemical composition is associated with many aerosol health and welfare effects: acid deposition (Calvert et al., 1985), impaired visibility through light scattering (e.g. Malm et al., 1994; Park et al., 2004; Pitchford et al., 2007), climate through effects on aerosol optical depth (Pilinis et al., 1995, Leibensperger et al., 2012), and climate through effects on cloud condensation and ice nuclei (e.g. Cruz and Pandis, 1997; Pöschl, 2005). Quantitative characterization of ambient aerosol size, mass, and chemical composition, including water content, is essential to adequately understand the fate and transport of chemicals in the Earth’s atmosphere, and to develop effective strategies that mitigate aerosol-related problems.

Water is an abundant atmospheric constituent that is present in the condensed phase as a function of relative humidity ($RH$), temperature, aerosol concentration and chemical composition (Zhou et al., 2011). Liquid water is estimated to represent a substantial fraction of total tropospheric aerosol volume at $RH > 85\%$ (Kreidenweis et al., 2008) and is predicted to exceed total aerosol dry mass by 2 to 3 times globally (Liao and Seinfeld, 2005). The aqueous phase also provides a medium for the partitioning of polar, water-soluble gas phase species (Asa-Awuku et al., 2010; Prisle et al., 2010), thus potentially facilitating secondary organic aerosol (SOA) formation (Carlton et al., 2009; Ervens et al., 2011; Carlton and Turpin, 2013). Despite the abundance and importance of aerosol water, it is not routinely measured, actual mass concentrations are not well known, and model predictions are poorly constrained.

The role of water in SOA formation in a particular environment is complex. In addition to the availability and relative abundance of particle phase liquid water, SOA enhancement depends on the amount of semi-volatile and water-soluble gas-phase material (Carlton and Turpin, 2013). For absorptive partitioning, SOA formation is dependent of the activity coefficient of the organic
in the solution (Pankow et al., 2001, Barley et al., 2009). The presence of inorganic material mixed with hydrophobic organic compounds may lead to liquid-liquid phase separation (Bertram et al., 2011) and the presence of liquid-liquid equilibria may influence partitioning (Prisle et al., 2010). Partitioning of water-soluble material is also influenced by the presence of inorganic material through salting effects (Lim et al., 2010; Knote et al., 2014). Semi-volatiles and dissolved gases that are associated with the aqueous phase may be lost by particle drying, which could hamper measurement and predictions of cloud condensation nuclei from dried aerosol streams (Topping and McFiggans, 2012).

Several techniques exist to measure aerosol hygroscopic properties. In general, hygroscopicity measurements characterize the change in aerosol properties in response to perturbations in $RH$. Sorooshian et al. (2008) provide a detailed overview of the different available techniques. Broadly, the approaches can be classified into methods that probe single sizes and methods that probe the entire aerosol. Popular single size methods include the hygroscopicity tandem differential mobility analyzer (HTDMA) technique (Liu et al., 1978; Rader and McMurry, 1986; Suda and Petters, 2013), the laminar flow tube approach (Stratmann et al., 2004; Wex et al., 2005), and the single particle levitation approaches (Tang, 1996; Mitchem and Reid, 2008). The HTDMA technique has been routinely deployed in field experiments, providing rich datasets for particle hygroscopic growth factors in a wide range of environments (e.g. McMurry and Stolzenburg, 1989; Berg et al., 1998; Dick et al., 2000; Swietlicki et al., 2008 (and references therein)). Popular methods that characterize the change in the entire aerosol with humidity include humidified nephelometry (Rood et al., 1985), aerosol hydration spectrometry (Stanier et al., 2004; Hegg et al., 2008; Snider and Petters, 2008; Engelhart et al., 2011), and gravimetric methods (Mikhailov et al., 2013). Single size methods are generally more precise and less ambiguous to interpret relative to bulk techniques. Bulk techniques, however, are useful because they characterize the entire aerosol rather than a subset and thus are needed to directly measure total water volume.

Here we report measurements obtained with an aerosol preconditioning system coupled with a scanning mobility particle sizer (SMPS) to explicitly measure total aerosol liquid water and to estimate the loss of semi-volatile compound volume during particle drying. Our study is motivated by the need to explore the hypothesis that particle water enhances biogenic SOA volume through aqueous-mediated partitioning of biogenically-derived organic species as the
result of anthropogenic perturbations (Carlton and Turpin, 2013). The humid, photochemically active summer of the southeastern U.S is ideal for conducting this study. Biogenic SOA mass concentrations are typically large (Lewis et al., 2004; Kleindienst et al., 2007; Ding et al., 2008) demonstrate positive relationships to RH (Hatch et al., 2011), and have been shown to be enhanced by the presence of sulfates and nitrates that affect particle water uptake (Chan et al., 2010). Further, model predictions suggest liquid water mass concentrations are high (Carlton and Turpin, 2013), and that aqueous phase water attributed to anthropogenic sulfate may influence biogenic SOA mass (Carlton et al., 2010; Hoyle et al., 2011) in the area.

The instrument was deployed as a part of the Southern Oxidant and Aerosol Study (SOAS), a collaborative field campaign during the summer season near Talladega National Forest in Brent, Alabama. The site is situated at (32.903°N, 87.250°W), and at an elevation of 126 m. Data were collected from 3 June to 15 July 2013. Our principle objectives were to measure continuous in situ aerosol volume distributions of the dry, and particle phase liquid water constituents over the six-week time period of SOAS, and to identify chemical and thermodynamic controls on particle phase liquid water content. This work contains a description and analysis of the instrument design, data reduction methods, and field measurement results based on measured RH, explores influences on ambient water content, and provides a method to estimate water content at different RH conditions.

2 Methods

2.1 Instrument design

A schematic of the instrument and experimental setup is presented in Fig. 1. Ambient air entered the instrument via a preconditioning inlet assembly consisting of a copper tube (9.5 mm ID, ~60 cm length) embedded in an aluminum block that was located ~1 m above ground level under the roof of an open shed. The temperature of the copper tube could be stably controlled between $T_{inlet} - 30\pm0.19$ ºC and $T_{inlet} + 50$ ºC (TE Technology LC-061). An optional Nafion membrane water-to-gas humidifier (PermaPure MH 110-48) was used to condition the aerosol. Subsequently the aerosol passed through a charge neutralizer (Aerosol Dynamics Inc. Model 100; Russell et al., 1996) holding four fresh $^{210}$Po charge strips (NRD StaticMaster 2U500) with a nominal total activity of 2 mCi. The aerosol was routed through an equilibration section (not pictured) before entering a high-flow differential mobility analyzer (DMA; Stolzenburg et al.,
1998). The DMA sheath flow was controlled by a critical orifice (O’Keefe Controls Co., 9 L min⁻¹) and configured in recirculation mode. Monodisperse aerosol exiting the DMA was counted by a condensation particle counter (CPC; TSI 3772) operated at 1 L min⁻¹. Aerosol transit times were 10 s through the inlet loop, 6 s through the charge neutralizer, 5 s in the equilibration section between the Nafion humidifier and the DMA entrance, and 10.4 s through the DMA column. The residence of sample at the measurement RH is comparable to those used in other studies of hygroscopicity (cf. Table 1, Duplissy et al., 2009).

The DMA was operated in scanning mobility particle sizer (SMPS) mode (Wang and Flagan, 1990). Negative voltage applied to the inner rod was held steady at 7 kV for 60 to 190 s, depending on the holding time of the present system state, and was then followed by an exponential decrease to 5 V over 300 s. Mapping between the time-varying electric field and selected particle mobility was achieved using the method of Wang and Flagan (1990). Conversion between particle mobility and mobility diameter followed standard DMA theory (Knutson and Whitby, 1975). The diameter range was determined by the sheath flow rate and DMA dimensions (60 cm tall, 11.6 cm ID outer cylinder, 10 cm OD collection rod) and spanned from 13 nm to ~1.1 µm. The number size distribution was found via a standard inversion that accounts for the transmission of multiply charged particles using the method described in Petters et al. (2009a) with empirically determined adjustments accounting for particle transmission efficiencies that are described in detail in Section 2.2.

Relative humidity and temperature control of the DMA column were unchanged from previous versions of the instrument (Suda and Petters, 2013). Since the instrument was placed inside a well-ventilated shed, and since the objective was to track ambient temperatures, the neoprene insulation used by Suda and Petters (2013) was removed. Despite best efforts the temperature inside the shed was slightly warmer than the outside, resulting in lower relative humidities inside the instrument relative to the values reported by the meteorology station. Both sheath and sample flows were optionally passed through Nafion humidifiers connected to a recirculating water bath. The water temperature determined the dew point temperature of the sheath and sample stream and was actively controlled by LabVIEW to match the ambient dew point temperature measured at the inlet. Temperature of the DMA column was measured using thermistors at the entrance, middle and bottom of the instrument. Two aluminum sleeves with PID controlled thermoelectric heat exchangers that were mounted on the outside were used to
reduce the standard deviation of the three thermistor temperatures to 0.21±0.16°C. The nominal RH inside the instrument was computed from the average of the three thermistor temperatures and the flow rate weighted average of the measured sheath and sample dew point temperatures from the RH sensors embedded in the flows (HC2 Rotronics, Hygroclip, RH = ±0.8% accuracy).

We have previously demonstrated (Suda and Petters, 2013) that this method of humidity control and measurement is sufficient for measuring aerosol hygroscopic growth and activity coefficients in tandem DMA studies at RH < 90%.

The panels in Fig. 1 show three instrument states. In the unperturbed state, no humidity conditioning occurred to the sample prior to particle sizing. The temperature of the preconditioning copper tube and the DMA column temperatures equaled the temperature measured at the inlet. The Nafion humidifier was bypassed. The sheath flow was actively humidified to match the relative humidity of the sample stream. This configuration measured the particle size distribution with minimal perturbation to sample temperature and RH.

In the dry instrument state, the temperature of the copper tube was chilled to 30 °C below the inlet temperature (Fig. 1, panel 2). The temperature drop causes substances with dew point temperatures higher than the tubing temperature to condense onto the wall. Measurements of relative humidity downstream of the copper tube confirmed that the dew point of the sample was less than or equal to the temperature of the aluminum block the copper tube was embedded in. Upon exiting the copper tube the sample flow warmed, resulting in a sharp drop in the water vapor saturation ratio. The sheath flow was conditioned in the same manner as the sample flow. This configuration measured the dry particle size distribution at RH ~ 10%.

The dry-humidified state is similar to the unperturbed state (Fig. 1, panel 3). The difference is that the sample aerosol was dried using the same method as the dry instrument state. A temperature reduction of 30°C is expected to drop the saturation vapor pressure of typical semi-volatile organic compounds by 1-2 orders of magnitude (Booth et al., 2010). Compounds that become supersaturated will condense onto the wall or on the particles. Warming of the sample flow to the original temperature results in lower saturation ratio of gas-phase organic species. Subsequent evaporation of now strongly subsaturated semi-volatile compounds may result in net loss of organic mass from the particle. Note that equilibration time for semi-volatile compounds is longer than the transit time through the equipment (~ 11 s between inlet loop and DMA entrance). For example, the time required to evaporate to a diameter that is within 10% of
the equilibrium size after a 30K warming varies between 30 and 1000 s for $\alpha$-pinene SOA and aged SOA (cf. Figure 2, Riipinen et al., 2010). We note that water soluble organic gases may be affected differently than semi-volatile compounds. Water soluble gases (e.g. glyoxal) have high vapor pressure and low saturation ratio. Due to their low saturation ratio they will not be removed by the cold trap by direct condensation onto the wall. However, some fraction may dissolve into water condensate that is forming due to removal of water vapor. The dry-humidified state was designed to test if the drying procedure resulted in net removal of particle volume.

The full duty cycle of the instrument was unperturbed, dry-humidified, and dry, followed by an automated cleaning cycle (not pictured in Figure 1). During the cleaning cycle the temperature of the copper tube was warmed to 30 °C above outside temperature and back flushed with ambient air to remove water and other condensed substances from the tube. Verifying that the dew-point temperatures measured before and after the copper tube were indistinguishable within experimental uncertainty ensured complete cleaning. Since the temperature of the copper tube had to be adjusted between the instrument states, there was a 160 s delay between the cleaning cycle and unperturbed state, 190 s between the unperturbed and dry-humidified states, 60 s between the dry-humidified and dry states. A total of 600 s was allotted for the cleaning cycle. Approximately two unperturbed, dry-humidified, and dry size distributions were acquired per hour.

2.2 Instrument Performance

Particle sizing accuracy was verified in the laboratory using polystyrene latex spheres (PSL; 102±3 nm; Thermo Scientific, lot #36489). The resulting measured size distribution was in agreement with the PSL size within the accuracy of the PSL spheres. Particle transmission efficiencies were quantified in the lab and the field using mobility selected effloresced ammonium sulfate particles. For these tests, particles were atomized from a stock solution (99.9% pure; Sigma-Aldrich; deionized water, ~ 18.2 MΩ cm) dried in silica gel diffusion dryer (TSI 3062), charge equilibrated, and size selected by a second DMA (same origin and dimensions as the primary DMA, operated at 9:2 sheath-to-monodisperse flow ratio).

Monodisperse aerosol was then routed to the SMPS and a second CPC (TSI 3771). Prior to these tests the two CPCs were intercompared sampling lab air side-by-side. Concentrations correlated
well and the systematic offset between the instruments was 7.6%, which is within the manufacturer tolerance (±10%). Using this setup, the kernel function accounting for DMA transfer entering the inversion algorithm (Petters et al., 2009a) for the selected bin resolution (60 bins) was determined for a series of mobility sizes between 20 and 600 nm. Reduced transmission efficiencies were encountered for $D < 50$ nm, presumably due to diffusion losses. To account for reduced transmission a diameter dependent loss correction was included into the inversion matrix. Overall performance of the instrument was confirmed by comparing DMA integrated number concentration with CPC measured number concentration obtained either in simultaneously or sequentially with the size distribution scan. Concentrations from the integrated size distribution and the CPC agreed within 10%.

The aforementioned procedure does not capture losses that may occur in the preconditioning inlet assembly. These losses were different for the unperturbed, dry-humidified, and dry state. To account for differential transmission between the different states, the unperturbed, dry-humidified, and dry states are multiplied by a constant factor 1, 1.2, and 1.12 respectively to match the concentrations of a co-located CPC that was available prior to the start of the campaign (27–29 May) as well as 3–15 July. Between 3 June and 3 July instrument performance was validated by disconnecting the CPC from the DMA during several cleaning cycles. After 3 July the CPC sample flow was reduced to 0.5 L min$^{-1}$ to accommodate 0.5 L min$^{-1}$ flow for a cloud condensation nuclei counter added to the setup. Thus the total flow through the DMA was unchanged. Number distribution readings from the CPC were adjusted accordingly to account for the reduced number of particles counted. The regular system performance checks were also carried out after the change. No degradation of instrument performance was observed. Figure 2 shows example time series of particle number concentration observed with the SMPS and condensation particle counters. The figure shows that there is generally satisfactory agreement between SMPS and CPC derived concentrations, although there are some time periods when the SMPS undercounted relative to the CPC (e.g. Fig. 2 bottom panel 15–20 hr). The reasons for this are not entirely clear but undercounting occurs preferentially when fine mode aerosol are present. This indicates that transmission correction for particles with $D < 50$ nm may not fully account for the combined losses in the inlet assembly and the DMA column. We do not believe that these deviations are of concern since aerosol volume is most sensitive to accumulation mode sizes, and thus the error in volume introduced by these deviations is small.
2.3 Data Reduction

The volume of water associated with the aerosol is described using the hygroscopicity model of Petters and Kreidenweis (2007) and Kreidenweis et al. (2008):

\[ V_w = V_d \kappa \frac{a_w}{1 - a_w}, \]  

(1)

where \(a_w\) is the water activity, \(V_w\) and \(V_d\) are the volume of water and dry aerosol components, and \(\kappa\) is the hygroscopicity parameter. In general \(\kappa\) may vary with water activity and depends on particle chemical composition. For a particle composed of multiple components the particle’s \(\kappa\) can be computed from the volume-weighted mixture of its dry components

\[ \kappa = \sum \varepsilon_i \kappa_i, \]  

(2)

where \(\varepsilon_i\) and \(\kappa_i\) are the volume fraction and hygroscopicity parameter of the \(i^{th}\) component comprising the particle. Water activity and \(RH\) are related via

\[ a_w = RH \exp \left( \frac{4 \sigma_w M_w}{\rho_w RTD} \right)^{-1}, \]  

(3)

where, \(\sigma_w, M_w\) and \(\rho_w\) represent the surface tension, molecular weight, and density of water, respectively, \(T\) is the absolute temperature, \(R\) is the ideal gas constant, \(D\) is the humidified particle diameter, and \(RH\) is the fractional relative humidity. Two hygroscopic growth factors can be defined:

\[ gf_{vol} = \frac{V_w + V_{dry}}{V_{dry}} \text{ and } gf_D = \frac{D}{D_d}, \]  

(4)

where \(gf_{vol}\) and \(gf_D\) are the volume and diameter based growth factors, and \(D_d\) is the dry particle diameter. The dry particle composition can be conceptually divided into a low volatile (lv) and a semi-volatile (sv) fraction. The component parameter sets are \(\{\varepsilon_{lv}, \kappa_{lv}, V_{w,lv} \text{ and } V_{d,lv}\}\) and \(\{\varepsilon_{sv}, \kappa_{sv}, V_{w,sv} \text{ and } V_{d,sv}\}\), respectively. Assuming that volumes are additive (see Section 2.4 for further discussion), the total volume of water and dry components are \(V_w = V_{w,lv} + V_{w,sv}\) and \(V_d = V_{d,lv} + V_{d,sv}\).

Aerosol water and aerosol semi-volatile volumes are quantified using the measured aerosol volume of three consecutively measured instrument states: unperturbed, dry-humidified, and dry. The measured volumes of the three states correspond to
\[ V_{\text{unperturbed}} = V_{w,lv} + V_{w,sv} + V_{d,lv} + V_{d,sv} \]
\[ V_{\text{dry}} = V_{d,lv} \]
\[ V_{\text{dry-humidified}} = V_{w,lv} + V_{d,lv} \]

where \( V_{\text{unperturbed}} \), \( V_{\text{dry}} \), and \( V_{\text{dry-humidified}} \) are the total particle volumes measured for each instrument state, again assuming that volumes are additive. The semi-volatile volumes in Eq. (5) are operationally defined to correspond to the amount that was lost during the drying procedure.

Consequently aerosol water volume associated with the low volatile material is

\[ V_{w,lv} = V_{\text{dry-humidified}} - V_{\text{dry}}. \]

As will be discussed in Section 4, the contribution of \( V_{d,sv} \) to \( V_w \) was not discernible. The semi-volatile volume lost during drying is

\[ V_{sv,\text{instrument}} = V_{\text{unperturbed}} - V_{\text{dry-humidified}}. \]

We note that the instrument defined loss of \( V_{sv,\text{instrument}} \) implies a loss of water that was hygroscopically bound in the particle. The amount of water associated with the semi-volatile fraction can be determined combining Eq. (7) with Eqs. (1) and (2) and solving for the amount of dry semi-volatile volume lost

\[ V_{d,sv} = V_{sv,\text{instrument}} \frac{1 - a_w}{1 - a_w + \kappa_{sv} a_w}. \]

Eq. (8) demonstrates that if \( \kappa_{sv} = 0 \) (corresponding to a hypothetical non-hygroscopic material that is lost during drying) \( V_{sv,\text{instrument}} \) is an accurate measure of \( V_{d,sv} \). For cases where \( \kappa_{sv} > 0 \), \( V_{sv,\text{instrument}} \) will overestimate the amount of semi-volatile material lost during drying. Eq. (8) can be used to estimate the magnitude of this effect.

Volumes entering Eqs. (6) and (7) are derived from the size distribution data. Volume distributions were calculated from the number size distributions assuming particle sphericity following Seinfeld and Pandis (2006). Figure 3 shows example histograms of the inverted 60-bin representation of the number and volume size distribution. The data show that the size distribution is bimodal with mode diameters \( D_{p1} \sim 0.06 \mu m \) and \( D_{p2} \sim 0.2 \mu m \). The relative contributions of number and volume to the ratio of mode #1 to #2 are \( \sim 10:1 \) and \( \sim 1:2 \), respectively. Low number concentrations result in poor counting statistics leading to the apparent noisiness of the volume distribution histograms in the larger size mode. A single false count in the largest diameter bins can significantly bias the total volume derived from the spectra. False
counts sometimes arise due to arcing in the DMA column. Arcing occurs at high relative humidity and high electric potential leading to a breakdown of the electric field in the column. This enables transmission of smaller particles that are falsely sized in the large bins. These particles would appear as a peak at $D \sim 1 \mu m$ (not seen in Fig. 3).

To filter out possible contributions from arcing, reduce the noisiness from low counting statistics, and identify the mode diameters in each scan, log-normal distribution functions (Hatch and Choate, 1929) are fitted to the distribution data. A non-linear least square fitting routine is used to minimize the residual between the data and the distribution function. Artifacts from rogue counts at high diameters are filtered since a 3rd mode is not allowed. Distribution functions are fit to both the number and volume distributions separately. Example fits to the histograms are superimposed in Fig. 3. To test the efficacy of the fits to represent the data, the fit and data integrated moments were compared. For each distribution the relative difference between integrated number concentration from the data and the fit was 1.4±5%, where 1.4% denotes the average error and ±5% the fit-to-fit variability of the error. Similarly, the statistics for the integrated volume are 1.6%±5.3%. The variability in the data greatly exceeds any artificial error introduced by the two-mode parameterization, indicating that the two-mode parameterization is appropriate to model the size distribution. Measured volume concentrations were correlated with mass concentrations from a co-located tapered element oscillating microbalance (Patashnick and Rupprecht, 1991, see supplementary information). The offset between these measurements is broadly consistent with an aerosol density of 1.5 $g \ cm^{-3}$, a value that is reasonable for the measured aerosol chemical composition. Note that the average number and volume fraction of particles with $D < 50$ nm to smaller mode is 30.6% and 19%, respectively. For those sizes instrument performance is degraded due to diffusional losses. However, since the contribution of $D < 50$ nm particles is less than 50% and since the potential error is likely serious only for $D < 30$ nm we do not expect that diffusional losses affect the fit-determined mode diameter.

The fitted volumes $V_{\text{unperturbed}}$ and $V_{\text{dry}}$ enter Eq. (6) to find $V_w$. Furthermore, $V_{\text{dry}}$ defines $V_d$ in Eq. (1). Calculations were also performed using the data distributions directly and summary statistics are provided in the online supplement. For these calculations, we assume $a_w$ is equivalent to $RH$ measured by the instrument, and the implications of this assumption are discussed later on in Section 2.4.
Alternatively, growth factors are defined by calculating the ratio of the shift in the mode diameters from the log-normal fit between $V_{\text{unperturbed}}$ and $V_{\text{dry}}$:

$$g_{fi} = \frac{D_{pgi,\text{unperturbed}}}{D_{pgi,\text{dry}}}$$  \hspace{1cm} (9)

where $g_{fi}$ is the diameter growth factor of the $i^{th}$ log-normal mode and $D_{pgi,\text{unperturbed}}$ and $D_{pgi,\text{dry}}$ are the fit-returned mode diameters of the $i^{th}$ log-normal mode for the sequential unperturbed and dry measurement cycles. Conversion from RH to water activity is performed using Eq. (3) and with $D_{pgi,\text{unperturbed}}$ as the particle diameter. The resulting diameter growth factors can be used to define $\kappa$ values that characterize the hygroscopicity of the individual modes:

$$\kappa_{Di} = (g_{fi}^3 - 1)(1 - a_w)a_w^{-1}$$  \hspace{1cm} (10)

where $\kappa_{Di}$ is the diameter based hygroscopicity parameter for the $i^{th}$ log-normal mode.

The fitted volumes $V_{\text{unperturbed}}$ and $V_{\text{dry-humidified}}$ enter Eq. (7) to find $V_s$, instrument. Since the scans were sequential, relative humidity was not necessarily the same between the two scans. To account for RH variations from the target value the $V_{\text{dry-humidified}}$ volume was empirically corrected to the RH corresponding the unperturbed cycle as recommended by Gysel et al. (2009). In our case, the correction is performed as follows. First, $\kappa_{\text{vol, dry-humidified}}$ is determined from $V_{\text{dry-humidified}}$ and $V_{\text{dry}}$ using Eq. (1) and $RH_{\text{dry-humidified}}$. Second, we obtain the corrected state using:

$$V_{\text{dry-humidified, corrected}} = V_{\text{dry}}\kappa_{\text{vol}} \frac{RH_{\text{unperturbed}}}{1 - RH_{\text{unperturbed}}} + V_{\text{dry}}$$  \hspace{1cm} (11)

where $V_{\text{dry-humidified, corrected}}$ is the corrected volume for the dry-humidified state, $\kappa_{\text{vol}}$ is the size distribution integrated volume based hygroscopicity parameter computed from Eq. (1), and $RH_{\text{unperturbed}}$ is the average RH measured during the unperturbed state.

### 2.4 Sources of uncertainty

Since $RH_{\text{unperturbed}}$ was slightly lower than that measured at the meteorological station due to slight heating of the shed, the reported water contents represent a lower estimate. The relative difference between the two $RH$ sensors, computed as $(RH_{\text{met}} - RH_{\text{shed}})/RH_{\text{met}}$, was 15%. Eq. (11) can be used to estimate the corrected water content at conditions deviating from instrument conditions.

The DMA measures mobility diameter, which for spherical particles is equal to the volume equivalent diameter. During the unperturbed and dry-humidified state, particles that
contain water will likely have a nearly spherical shape because water preferentially adsorbs at the corners and edges of the particle (Mikhailov et al., 2004). However, some particles may have a crystalline structure when dried. Particle shape irregularities increase the drag inside the DMA and result in a larger electric mobility diameter measured. Compounds with a cubic structure, for example, have a shape factor of 1.08 (e.g. Kelly and McMurry, 1992). Gysel et al. (2002) calculated that the relation between the volume equivalent diameter and the mobility diameter of a cubic particle is 0.96, which indicates a 4% overestimate of particle diameter by the DMA for cubic particles. As a result, the volume distribution calculated from the mobility diameters of irregularly shaped particles during the dry state may be overestimated, which would result in an underestimate of water content when we difference the dry-humidified and dry states.

The calculations also assume volume additivity, i.e. that aerosol species volumes are independent of solution concentration and of species mixing fractions. Detailed studies of water activity for inorganic compounds show that excess volume of mixing can be large for some systems, particularly at low water content (Clegg et al., 1998; Wexler and Clegg, 2002). However, other studies find that volume additivity holds, although this may be due to the cancellation of excess volumes (Stokes and Robinson, 1966). Mikhailov et al. (2004) found volume additivity to be a safe assumption for aerosols composed of proteins and salts for which the individual densities and behavior in solution were known. In the absence of detailed knowledge of composition and the component behavior in solution, as is the case with ambient aerosol, volume additivity has typically been assumed (Dick et al., 2000; Speer et al., 2003). Dick et al. (2000) constrained their results with the calculation that for simple aqueous inorganic solutions volume additivity mis-estimates particle phase liquid water by -7% for H₂SO₄ at RH = 90% and by < 5% for deliquesced NH₄HSO₄, (NH₄)₃H(SO₄)₂, and (NH₄)₂SO₄, thus providing an estimate of error for different ammonium-to-sulfate ratios for the inorganic fraction of ambient aerosols. In sum we conclude that the assumption of volume additivity may introduce a small error in our calculations when molecular interactions in the solution deviate from the pure components.

Since multiple sizes contribute to the fitted volumes, the conversion from instrument RH (Section 2.1) to \( a_w \) via Eq. (3) is not straightforward. We therefore assume \( a_w \) is equivalent to RH averaged over the scan to compute \( \kappa_{vol} \). The expected difference between RH and water activity
are between 0.01 and 0.02 in absolute units (e.g. Tang, 1996) and will result an RH dependent overestimate in $\kappa_{vol}$ ranging from 4 to 11%.

Changes in aerosol solution from stable to metastable conditions during the dry-humidified state can also introduce errors into the calculations of water volume. Inside the Nafion tube during the dry-humidified state, the RH approaches ~100% at the temperature that approximates the dew point temperature of the outside air. Thus most particles that can deliquesce at subsaturated relative humidity will contain water exiting the Nafion. Particles may or may not effloresce prior to entering the DMA. If $V_{dry-humidified}$ exceeds $V_{unperturbed}$, it may be possible that a sufficiently large volume of water was added by the deliquescence of water on particles that were effloresced under unperturbed conditions.

Some compounds do not effloresce and thus may retain water at low RH. In that case the measured $V_{dry}$ will be overestimated and the instrumented-inferred $g_{fvol}$ and $\kappa_{vol}$ will be biased low. The relative error in the inferred $\kappa_{vol}$ at $a_w=0.12$, for example, is $e_\kappa = (\kappa_{vol,dry} - \kappa_{vol,0.12})/\kappa_{vol,dry}$, where $\kappa_{vol,dry}$ is the true hygroscopicity, and $\kappa_{vol,0.12}$ is the hygroscopicity one would calculate if particle-bound water was present at $a_w=0.12$. Eq. (1) can be used to estimate the magnitude of the particle-bound water at $a_w = 0.12$, assuming that $\kappa$ is invariant with $a_w$. From that the total particle volume ($V_w + V_d$) can be computed. If the total volume is mistakenly assumed to equal the dry volume, a $\kappa_{vol,0.12}$ can be computed. The estimated error is independent of $a_w$ and scales with $\kappa_{vol,dry}$. For $\kappa_{vol,dry} = 0.01$ and $\kappa_{vol,dry} = 0.6$, the error is $e_\kappa = -0.1\%$ and $e_\kappa = -7.6\%$, respectively. We also note that the dry instrument state corresponds to $RH = 12.5\% \pm 2.3\%$, where 2.3% denotes the variability of the RH in absolute units at the dry state throughout the campaign.

Although losses of semi-volatile compounds during drying are expected, the total amount lost unlikely corresponds to what would be expected from thermodynamic equilibrium. Several reasons are listed here. First, fast kinetics for aerosol dissolution are required. However, this may not be the case if the initial RH is low, as the aerosol becomes viscous and formation of a homogenous liquid phase may be slow (Vaden et al., 2011; Renbaum-Wolff et al., 2013). Second, evaporation rates must be fast enough to remove semi-volatiles in the inlet even though evaporation rates may be slow (Bilde et al., 2003). Succinic acid, for example, has been measured to have an evaporation rate of 2.76 nm s$^{-1}$ at 299.8 K and 64.1% RH (Koponen et al.,
The residence time of the preconditioning inlet may not be adequate to capture full evaporation of succinic acid. Third, the phase state of the particles after drying is uncertain. Sub-cooled liquid vapor pressure may be up to three orders of magnitude higher than solid vapor pressure (Booth et al., 2010). Since partitioning between gas and particle phase is dependent on the saturation vapor pressure of the organic compound in the liquid state (Pankow, 1994), semi-volatile content may be underestimated if particles are in the solid state after drying. However, water in the organic phase could reduce organic equilibrium partial pressure according to Raoult’s law (Pankow and Chang, 2008), though this effect did not play a strong role on α-pinene SOA formation (Prisle et al., 2010). Detailed studies with proxy systems are needed to quantify the amount removed as a function of compound volatility in the context of particle phase/viscosity/diffusion and non-ideal solution effects. Therefore, the tests with the preconditioning inlet only test whether the selected drying procedure lead to loss of material during SOAS and is not intended to be used as a measure of the semi-volatile fraction.

### 2.5 Chemical Composition

PM$_{2.5}$ or PM$_1$ water soluble ions were measured by a Particle-Into-Liquid-Sampler coupled with an Ion Chromatograph (PILS-IC; Metrohm 761 Compact IC. Similar setups are described in previous field studies (Orsini et al., 2003, Hecobian et al., 2010, Liu et al., 2012). Metrosep A Supp 5 anion column and C 4 150 cation column (Metrohm USA, Riverside FL) were used to separate PILS liquid sample anions sulfate, nitrate, chloride, oxalate, acetate, formate and cations ammonium, sodium, potassium, calcium, magnesium with 20 min duty cycle. Water soluble organic carbon (WSOC) was measured by coupling the PILS with a total carbon analyzer or a total carbon analyzer (TOC, Model 900, GE Analytical Instruments; Boulder, CO). The TOC was operated in normal mode with a 6-min duty cycle. The PILS sample air flow rate was 16.8 ± 0.35 L/min and URG cyclones inlets provided PM cut sizes of PM$_{2.5}$ for the 1st half of field study (June 1 to June 22) and PM$_1$ for the latter half (June 23 to July 15). Honeycomb acid (phosphoric acid) and base (sodium carbonate) coated denuders remove interfering gases. Water soluble organic matter (WSOM) was then estimated from WSOC measurements using a conversion factor of 2.1 (Turpin and Lim, 2001). The sample inlet was ~7m above ground level and ~4m long. The inlet line was insulated inside a trailer (typical indoor T was 25°C) and less than 1m in length to minimize possible changes in aerosol
composition prior to measurement. The trailer was located adjacent to the shed housing the SMPS system. Periodic 1 hour blank measurements were made every day by placing a HEPA filter (Pall corp.) on the cyclone inlet. All data were blank corrected.

3 Results

Figure 4 provides a time-series of the measured data for the six weeks of data collection. A campaign-based statistical summary of selected parameters encountered in the instrument is summarized in Tables 1 and 2. Temperatures measured by the instrument typically fluctuated between ~24 °C during nighttime and ~30 °C during daytime. Minor fluctuations in the dew point temperature ($T_{dew}$) indicate that the diel cycle of RH was driven mostly by temperature variations – highest at low temperatures and lowest at high temperatures. Daily fluctuations for RH within the instrument were typically within 60% to 77%. Periods of heavy rainfall included 5–7 June, 18 June, and 3–6 July. The highest amount of precipitation for a single day was 45.8 mm on 6 June. Periods with little to no rainfall included 8 June, 11–12 June, 14–16 June, 19–22 June, 25–26 June, and 9–12 July. Periods with noticeably high amounts of rainfall, such as 5 and 6 June, resulted in low levels of dry ($V_d$) and water aerosol ($V_w$) volume. During periods with no rainfall, such as the days between 19 and 22 June, dry particle volume steadily increased, presumably due to the lack of an efficient aerosol sink.

Statistics of the campaign average bimodal distributions of number concentration, surface area, and volume for the three-instrument inlet states are summarized in Table 2. A graphical interpretation of Table 2 is provided in Fig. 5. Average number concentrations were ~2300 cm$^{-3}$ and 500 cm$^{-3}$ for the smaller and larger diameter mode, respectively. Surface area distributions are higher for the unperturbed and dry-humidified states than for the dry state due to the additional water uptake. For the smaller diameter mode, surface areas average at 54.0 µm$^2$ cm$^{-3}$ for both the unperturbed and dry-humidified states, and 54.4 for the dry state, while for the larger mode, the averages are 130, 127, and 95 µm$^2$ cm$^{-3}$ for the unperturbed, dry-humidified, and dry states respectively. This indicates that water uptake increases the aerosol surface area by 33.7% for the large diameter mode, while the effect for the small diameter mode is not observed, suggesting that the smaller mode is less hygroscopic. (Note that the surface area of the small mode nominally shrinks by 0.07% upon humidification. Possible reasons are noise in the measurement and that some fraction of the particles is picked up in mode #1 for the “dry” fit and
mode #2 for the humidified fit.). Similarly, the volume distributions for the unperturbed and dry-humidified state are both larger than the distribution for the dry state. Volume averages are 2.20, 2.18, and 1.49 µm³ cm⁻³ for the smaller diameter mode and 6.50, 6.93, and 3.87 µm³ cm⁻³ for the larger diameter mode, for the unperturbed, dry-humidified, and dry states, respectively. The observed size distribution statistics are consistent with previous measurements of accumulation mode remote continental aerosol size distributions (Jaenicke, 1993; Martin et al., 2010; Levin et al., 2012). Notably, small particle events indicative of nucleation (Levin et al., 2012) were not observed during the campaign. Number and volume concentrations were significantly higher during SOAS than during the Amazonian Aerosol Characterization Experiment (Martin et al., 2010), suggesting anthropogenic influence, likely from sulfates, on the SOAS site (Carlton and Turpin, 2013). One would expect that the presence of sulfates would result in more hygroscopic aerosol in Alabama compared to the pristine Amazon rainforest.

Figure 4 summarizes trends of particle phase liquid water (\(\mathcal{V}_w\)) throughout the campaign at the RH encountered in the instrument. Measured water concentrations always exceeded 0 µg m⁻³, typically varied between 1 and 5 µg m⁻³, and peaked at 73 µg m⁻³ on 17 June. The campaign time-series data indicate a strong diel cycle for \(\mathcal{V}_w\). A campaign-average representation of the diel cycle is presented in Fig. 6. We note that this average diel cycle was suppressed during periods with heavy washout. Analysis contrasting selected time periods is provided in Appendix A. Figure 6 shows that water volume peaked during the early morning hours ~7 a.m. to 9 a.m. LT. On average, the median value for water concentrations was 2.9 µg m⁻³, and the interquartile range was between 1.3 and 4.9 µg m⁻³ (Table 1). For 7 a.m. to 9 a.m. LT, however, the average median value for water mass concentrations was 5.8 µg m⁻³. The start of the decreasing transition for RH occurred in the morning at approximately 6 a.m. LT. The peak in water content occurred shortly after RH decreased and RH continued its decline until it increased again at 4 p.m. LT. Water mass concentrations leveled off after the morning peak, and despite the RH increase after 4 p.m. LT, water mass concentrations did not experience a noticeable increase in response during that time period. The partial decorrelation of RH and water volume suggests that either aerosol dry volume or aerosol hygroscopicity evolved during the day.

The evolution of composition is demonstrated further by the volumetric growth factor data. Figure 7 shows the diurnal profile of \(g_{\text{vol}}\) overlaid with expected \(g_{\text{vol}}\) assuming an invariant \(\kappa\) throughout the day. Analysis in the Appendix is performed for a volumetric growth factor
profile for a single day as an illustrative example of a day with no rainfall. The data shows that $gf_{vol}$ is always higher than 1, and exceeds 2 between 7 and 9 a.m. LT. Overall, the $gf_{vol}$ median is 1.61 (Table 1). The data also show that the observed interquartile range for $gf_{vol}$ is between 1.37 and 2.01. These values are similar to those found during the Pittsburgh Air Quality Study (Khlystov et al., 2005). The overlaid expected growth factor for different values of $\kappa$ in the plot suggest that the aerosol hygroscopicity during the campaign changed systematically throughout the day, which is further explored in the diel cycle of $\kappa_{vol}$ shown in Fig. 8. Overall, the median observed value for $\kappa_{vol}$ is 0.27 and the interquartile range is from 0.14 to 0.46. At night, $\kappa_{vol}$ is steady at median values of ~0.18 and quartiles of ~ 0.08 to 0.3. After 7 a.m. LT, $\kappa_{vol}$ increases to about a median value of ~0.48 and quartiles of ~ 0.2 to 0.7 followed by a gradual decrease throughout the rest of the day. The hygroscopicity statistics for the two size modes are summarized in Table 1. Medians of $\kappa$ for the smaller and larger diameter mode average to $\kappa_{D1} = 0.13$ and $\kappa_{D2} = 0.29$. In comparison, medians for $\kappa_{vol}$ average to 0.27, which more closely matches $\kappa_{D2}$ than $\kappa_{D1}$. $\kappa_{D1}$ also exhibits greater variability than either $\kappa_{vol}$ or $\kappa_{D2}$, which is most likely attributable to noisiness in the fittings on a scan by scan basis that increases variability in the diameter growth factors for the mode.

The measured $\kappa_{vol}$ are consistent with a mix of hygroscopic organic and inorganic compounds (Petters and Kreidenweis, 2007; Kreidenweis et al., 2008; Suda et al., 2012). The observed $\kappa_{vol}$ are significantly larger than those in organic dominated environments encountered in remote continental forests (Gunthe et al., 2009; Sihto et al., 2011; Levin et al., 2012; Mikhailov et al., 2013), which vary between 0.1 < $\kappa$ < 0.2. In these studies the aerosol were dried prior to measurement. We therefore also tested whether drying removed semi-volatile compounds and thus bias the observed $\kappa_{vol}$.

Results for the diel cycle of the semi-volatile fraction computed from Eqs. (7) and (11) are summarized Fig. 9. No clear diel trend is noticeable. At times the average semi-volatile volume concentration is negative. However, the 95% confidence interval of the mean consistently envelops zero. This is consistent with the null hypothesis that no semi-volatiles were removed in our preconditioning system and that the measurement has random error. We note that the confidence limits are ~ ±0.5 µg m$^{-3}$, which implies that point-to-point variability in the semi-volatile retrieval led to significant uncertainty. Losses of semi-volatile material less than 0.5 µg m$^{-3}$ may have occurred but were undetectable with our method.
The diel evolution of the hygroscopic species sulfate, nitrate, ammonia, and WSOM is shown in Fig. 10. Ammonia and WSOM are approximately constant while sulfate and nitrate show a diel cycle with a daytime maximum. We note that these speciated compounds only represent a subsection of the total aerosol since dust and elemental carbon are not included. Dust and elemental carbon are non-hygroscopic and therefore will not affect total water mass. Their presence will, however, lower the derived net $\kappa$ value of the aerosol. Overall, the sum of the median mass concentrations for sulfate (1.7 $\mu$g m$^{-3}$), nitrate (0.07 $\mu$g m$^{-3}$), ammonia (0.47 $\mu$g m$^{-3}$), and WSOM (3.15 $\mu$g m$^{-3}$) is 5.39 $\mu$g m$^{-3}$ and these species likely played an important role in governing aerosol water uptake during SOAS.

4 Discussion

The dynamics governing the total water volume that is available for water-mediated partitioning and condensed aqueous phase reactions are complex. First, our results show that at minimum 7% of the aerosol volume is composed of water at the measured RH. Day-to-day variations in aerosol hygroscopicity are relatively minor and total water volumes scale with dry aerosol volume and RH. The fractional water content is highest during the early morning hours when temperatures are lowest and RH is at a maximum. During those times volume growth factors exceed two, implying that the aerosol composition is dominated by water. Second, on timescales longer than the diel cycle, dry aerosol volume is the dominant determinant of absolute water content. For example, between 3 and 9 June (Fig. 4), dry aerosol volume increased from 1 to 10 $\mu$m$^3$ cm$^{-3}$, with concomitant increases in water volumes. Dry aerosol volume concentrations result from a balance in production (emissions and secondary production via chemical reactions) and removal (dry deposition, washout, and venting to the free troposphere) processes. The buildup of dry aerosol volume during stagnation events seems to be tied to the meteorological conditions. Warm temperatures, high actinic flux, and absence of deep convection and precipitation lead to accumulation of aerosol volume. The availability of a significant aqueous phase (by total volume) is thus correlated with, and contingent on, suitable weather conditions. Finally, the diel cycle of aerosol hygroscopicity suggests that significant changes in aerosol composition may occur on the time scale of a few hours. This is supported by changes in the inorganic aerosol fraction derived from the PILS measurements shown in Fig. 10e. Variation ranges from 20% to 80% for the inorganic fraction and the diel cycle in organic fraction mimics
that of $\kappa$ (Fig. 10f). However, the actual relationship between chemical composition and $\kappa$ is complex since $\kappa$ may strongly vary with water content (or $RH$) for concentrated solutions (Kreidenweis et al., 2008). Assuming that the PILS measured chemical composition captures most of the species responsible for water uptake and assuming a representative composition for the WSOM fraction we estimated the $RH$ dependence using the E-AIM model (Clegg et al., 1998). These calculations show that the reduction of $RH$ from 90% to 40% may cause a 60% increase in $\kappa$ (see supplemental information). The diel fluctuation in $\kappa_{vol}$ is from 0.14 to 0.46. Notably, the aerosol is most hygroscopic when the fractional water content is larger, which is consistent with the modeled $RH$ dependency of $\kappa$. The magnitude of the $RH$ dependence, however, is insufficient to explain the full diel $\kappa$-cycle which is driven by a combination of the two effects. We note that deconvolution of the two effects will require closure calculations between time-resolved chemical composition and $\kappa$ that are contingent upon complete speciation of the aerosol (including black carbon and dust), detailed information about the composition of WSOM, and estimation of aerosol solubility and density (see supplemental information).

Figure 11 shows that there was not strong diel profile in dry aerosol volume. Thus diel changes in aerosol hygroscopicity are indicative of dynamic aerosol chemistry that does not involve major volume changes. Another possible explanation for the observed $\kappa_{vol}$ changes are chemical transformations of the organic fraction itself that lead to corresponding changes in $\kappa_{org}$. Specifically, $\kappa_{org}$ has shown to increase with higher oxygen-to-carbon ratio (Jimenez et al., 2009), smaller molecular size (Petters et al., 2009b), and higher solubility in water (Petters and Kreidenweis, 2008). The presence of an aqueous phase combined with the morning actinic flux could drive aqueous phase oxidation reactions or lead to hydrolysis reactions resulting in smaller, more oxidized, and water-soluble organic molecules or ions. Perhaps consistent with aqueous phase processes, the hygroscopicity parameter $\kappa$ increases with particle size, similar to trends of $\kappa$ with particle size were observed in previous field studies (Gunthe et al., 2009; Wiedensohler et al., 2009; Levin et al., 2012). The higher $\kappa$ values for the larger mode suggest the presence of more hygroscopic solutes such as more oxidized or hydrolyzed organic species, sulfates or nitrates. A direct consequence of the size dependence is that the aqueous phase volume is predominately located with the larger particle sizes both due to the larger dry aerosol volume and the larger hygroscopicity. This initiates the
question: of the three parameters influencing particle phase liquid water (dry aerosol volume, hygroscopicity, and \( RH \)), which are the most important under ambient conditions?

We address this question using sensitivity analysis following the approach of (Feingold, 2003; McFiggans et al., 2006). The sensitivity is defined as \( S(X_i) = \partial \ln V_w / \partial \ln X_i \), where \( X_i \) is one of \( V_d, RH \), or \( \kappa_{vol} \). Comparing values of \( S(X_i) \) describes their relative importance given equal perturbations \( \delta V_d, \delta RH \), or \( \delta \kappa_{vol} \). Based on Eq. (1), sensitivities \( S(\kappa_{vol}) \) and \( S(V_d) = 1 \), while \( S(RH) = 1/(1 - RH) \). These sensitivities, combined with values for \( \delta X_i \) are summarized in Table 3. The interpretation of \( S(X_i) = 1 \) is that e.g. a 10% relative change in parameter \( X_i \) will lead to a 10% relative change in \( V_w \). Changes in hygroscopicity, dry aerosol volume, and \( RH \) are potentially cross-correlated. For example, the condensation of sulfuric acid onto organic aerosol will lead to both increases in \( \kappa \) and \( V_d \), thereby influencing \( V_w \) via both pathways. In contrast, the condensation of nonhygroscopic organic material (i.e., \( \kappa = 0 \)) will lead to no increase in \( V_w \) as the effects of increased volume and decreased hygroscopicity cancel after application of the ZSR mixing rule for \( \kappa \) (Petters and Kreidenweis, 2007). From observations during SOAS, the perturbations \( \delta \kappa \) and \( \delta V_d \) are comparable, resulting in 0–200% variability of \( V_w \) depending on the hygroscopicity of the compound that controls the variability of the dry aerosol volume.

Although the relative variability in \( RH \) is typically lower than \( \kappa \) or \( V_d \), the relative sensitivity of water volume to changes in \( RH \) is much larger. At \( RH = 40\%, 70\%, \) and \( 90\% \), \( S(RH) = 1.66, 3.33, \) and \( 10 \), respectively. Thus, at \( RH = 90\% \), a 1% relative fluctuation in \( RH \) will be equivalent to controls on water content by a 10% change in dry aerosol volume. As a result, the diel trends in water are primarily controlled by changes in relative humidity. The strong dependence of \( V_w \) on relative humidity is similar to that reported in previous studies (i.e., Khlystov et al., 2005). Since dew points are relatively constant during the day at the site location (see Fig. 4), we believe that the diel \( V_w \) cycle is indirectly controlled by temperature. Correlations between ambient temperature, relative humidity, and particle phase liquid water can potentially obfuscate the attribution of semi-volatile partitioning into the condensed phase to either the presence of water due to increased \( RH \) or reduced vapor pressure due to reduced temperature. Although this conceptual distinction is important, it is irrelevant for this study because loss of semi-volatile volumes \( V_{sv} \) during our drying procedure was not statistically discernible (Fig. 9). The absence of this effect is likely due to the mismatch of residence time (~ 11s) and slow equilibration time of the organic phase with after a perturbation in of the gas-phase saturation ratio, which is system
dependent and may exceed 60 min (Riipinen et al., 2010, Cappa and Jimenez et al., 2010). Improved methodology will need to lengthen residence time, reduce the uncertainty to resolve removal of less than 0.5 µg m\(^{-3}\) amounts, and establish volatility limits that can be detected using proxy system, lengthen residence time between the exit of the cold trap and SMPS to ensure full equilibration, and test whether the conclusion is valid when single particle sizes are considered.

5 Conclusions

We present measurements of dry aerosol volume, particle phase liquid water, and dry aerosol volume changes that may occur during particle drying. The measurements were obtained during the SOAS campaign at Talladega National Forest in Brent, AL from 3 June to 15 July 2013 to collect six weeks of continuous in situ measurements of ambient aerosols. Aqueous phase liquid water was always present, comprising at minimum 7% of aerosol volume at the measured RH. Water mass concentrations at the RH encountered in the instrument typically ranged from 1 to 5 µg m\(^{-3}\). Liquid water was the dominant aerosol constituent from ~7 a.m. to 9 a.m. LT. The diel water-content cycle was dominated by changes in RH, which mostly fluctuated due to changes in temperature. A strong diel cycle in aerosol hygroscopicity is consistent with observed systematic changes in aerosol composition and a water-content dependency of \(\kappa\). Aerosol hygroscopicity increased with particle size. The method of particle drying used in this study did not lead to removal of significant material that was associated with the aqueous phase.

Acknowledgements

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Appendix A

Conclusions based on campaign-based average diel cycles may be influenced by different regimes. To explore the possible influence of averaging we repeat the analysis performed in Fig. 7 for a single day in Fig A1. Comparing Figs. 7 and A1 suggests that the reported diel trend accurately captures the behavior for a single day. An unusual cold and rainy period occurred between 4 and 8 July 2013. During that time, aerosol volume was low and diel temperature and RH fluctuations were minimal. Fig. A2 contrasts the diel cycle for particle phase liquid water for a stagnation event with a pronounced diel cycle and the cold period. The data show that no diel cycle was observed during this atypical period, suggesting that conclusions reached about typical regional southeastern U.S. aerosol must be interpreted in the context of the meteorological setting.

References


Table 1. Mean and quartiles of number concentration $N_{\text{conc}}$, relative humidity $RH$, temperature ($T$), particle phase water volume $V_w$, semi-volatile volume $V_{sv}$ lost during drying, volume of solutes $V_d$, $\kappa_{\text{vol}}$, $\kappa_{D1}$, $\kappa_{D2}$, and $g_{f\text{vol}}$.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Units</th>
<th>Mean</th>
<th>25% Quartile</th>
<th>50% Quartile</th>
<th>75% Quartile</th>
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<tbody>
<tr>
<td>$N_{\text{conc}}$</td>
<td>cm$^{-3}$</td>
<td>2629</td>
<td>1531</td>
<td>2155</td>
<td>3136</td>
</tr>
<tr>
<td>$RH$</td>
<td>%</td>
<td>68.94</td>
<td>59.95</td>
<td>70.85</td>
<td>77.10</td>
</tr>
<tr>
<td>$T$</td>
<td>$^\circ$C</td>
<td>27.48</td>
<td>24.85</td>
<td>26.73</td>
<td>30.06</td>
</tr>
<tr>
<td>$V_w$</td>
<td>$\mu$m$^3$ cm$^{-3}$</td>
<td>4.00</td>
<td>1.34</td>
<td>2.88</td>
<td>4.86</td>
</tr>
<tr>
<td>$V_{sv}$</td>
<td>$\mu$m$^3$ cm$^{-3}$</td>
<td>0.18</td>
<td>-0.65</td>
<td>0.22</td>
<td>1.05</td>
</tr>
<tr>
<td>$V_d$</td>
<td>$\mu$m$^3$ cm$^{-3}$</td>
<td>5.29</td>
<td>2.95</td>
<td>4.65</td>
<td>7.17</td>
</tr>
<tr>
<td>$\kappa_{\text{vol}}$</td>
<td>dimensionless</td>
<td>0.33</td>
<td>0.14</td>
<td>0.27</td>
<td>0.46</td>
</tr>
<tr>
<td>$\kappa_{D1}$</td>
<td>dimensionless</td>
<td>0.47</td>
<td>-0.11</td>
<td>0.13</td>
<td>0.78</td>
</tr>
<tr>
<td>$\kappa_{D2}$</td>
<td>dimensionless</td>
<td>0.38</td>
<td>0.12</td>
<td>0.29</td>
<td>0.56</td>
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<tr>
<td>$g_{f\text{vol}}$</td>
<td>dimensionless</td>
<td>1.78</td>
<td>1.37</td>
<td>1.61</td>
<td>2.01</td>
</tr>
</tbody>
</table>
Table 2. Parameters describing two-mode distributions of number concentration, surface area, and volume for the unperturbed ($C_{\text{unperturbed},i}$), dry-humidified ($C_{\text{dry-humidified},i}$), and dry states ($C_{\text{dry},i}$).

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Units</th>
<th>Number N</th>
<th>Surface S</th>
<th>Volume V</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>1</td>
<td>2</td>
<td>1</td>
</tr>
<tr>
<td>$C_{\text{unperturbed},i}$</td>
<td>cm$^{-3}$</td>
<td>2254</td>
<td>507</td>
<td>54.0</td>
</tr>
<tr>
<td>$g_{\text{md,unperturbed},i}$</td>
<td>µm</td>
<td>0.099</td>
<td>0.24</td>
<td>0.11</td>
</tr>
<tr>
<td>$g_{\text{sd,unperturbed},i}$</td>
<td></td>
<td>1.54</td>
<td>1.43</td>
<td>1.54</td>
</tr>
<tr>
<td>$C_{\text{dry-humidified},i}$</td>
<td>cm$^{-3}$</td>
<td>2238</td>
<td>511</td>
<td>54.0</td>
</tr>
<tr>
<td>$g_{\text{md,dry-humidified},i}$</td>
<td>µm</td>
<td>0.11</td>
<td>0.23</td>
<td>0.12</td>
</tr>
<tr>
<td>$g_{\text{sd,dry-humidified},i}$</td>
<td></td>
<td>1.49</td>
<td>1.49</td>
<td>1.5</td>
</tr>
<tr>
<td>$C_{\text{dry},i}$</td>
<td>cm$^{-3}$</td>
<td>2355</td>
<td>403</td>
<td>54.4</td>
</tr>
<tr>
<td>$g_{\text{md,dry},i}$</td>
<td>µm</td>
<td>0.096</td>
<td>0.2</td>
<td>0.10</td>
</tr>
<tr>
<td>$g_{\text{sd,dry},i}$</td>
<td></td>
<td>1.47</td>
<td>1.43</td>
<td>1.47</td>
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</table>
Table 3. Table of sensitivity ratios $S(X_i)$ based on Eq. (1) for $X_i$ equal to $\kappa_{vol}$, $V_d$, and $RH$. The table also reports the typical range of these parameters during the SOAS campaign.

<table>
<thead>
<tr>
<th>$X_i$</th>
<th>$S(X_i)$</th>
<th>Typical range during SOAS</th>
<th>Average of range</th>
</tr>
</thead>
<tbody>
<tr>
<td>$\kappa_{vol}$</td>
<td>1</td>
<td>$0.1 &lt; \kappa_{vol} &lt; 0.6$</td>
<td>0.3</td>
</tr>
<tr>
<td>$V_d$</td>
<td>1</td>
<td>$1 &lt; V_d &lt; 10 \mu g m^{-3}$</td>
<td>5 $\mu g m^{-3}$</td>
</tr>
<tr>
<td>$RH$</td>
<td>$\frac{1}{1 - RH}$</td>
<td>$0.4 &lt; RH &lt; 1$</td>
<td>0.7</td>
</tr>
</tbody>
</table>
Fig. 1. Instrument setup schematic. Red lines indicate the sample flow, black lines indicate the sheath flow, orange lines indicate the cleaning cycle flow, and grey lines indicate no flow. The column central to the figure is the differential mobility analyzer (DMA). Sheath and sample flows in the diagram are downward in the center and upward at the sides. The cleaning cycle is similar to the unperturbed cycle but with solenoid valve A toggled such that flushing air flows toward the actuated ball valve G and is split between the sample path to the CPC and the inlet, reversing the flow through the temperature controlled preconditioner.
Fig. 2. Top panel: Example time series from 8 June. Symbols correspond to article number concentration obtained from integration over the size distribution (red = unperturbed, blue = dry-humidified, black = dry). Grey line corresponds to 1 Hz particle concentration from the CPC upstream of the SMPS with the inlet opened to ambient air during cleaning cycles. The plot shown here is a sample plot of the comparisons for 8 June. Bottom panel: Example time series from 14 July. Symbols are the same as in the top panel. Grey line corresponds to 1 Hz particle concentration from a second CPC sampling side-by-side with the SMPS.
Fig. 3. Example data from 5 July 2013. Histograms represent measured 60-bin representation of the number size distribution (top row) and derived volume size distributions (bottom row) for the unperturbed (left panels) dry-humidified (middle panels) and dry (right panels) instrument states. Red lines correspond to a bimodal log-normal distribution fitted to the data. Data-derived and fit-derived number and volume concentrations are provided in the legend. Blue numbers indicate the mode diameters obtained from the fit to the distributions.
**Fig. 4.** From top to bottom in forefront of figure: Time series of 1) temperature ($T$, °C) in brown, 2) dew point temperature ($T_{dew}$, °C) in dark grey, 3) relative humidity ($RH$, %) in green, 4) aerosol dry volume ($V_d$, µm$^3$ cm$^{-3}$) in pink, 5) volumetric growth factor ($gf_{vol}$, dimensionless) in light blue, 6) volumetric $\kappa$ ($\kappa_{vol}$, dimensionless) in orange, and 7) particle phase liquid water ($V_w$, µm$^3$ cm$^{-3}$) in dark blue. Precipitation (Precip, mm day$^{-1}$) is represented in grey bars at the bottom of the plot. All of the data except for precipitation were recorded by the SMPS. Gaps in the data indicate periods when the instrument was not operating. Precipitation data is from the ARA SEARCH network.
**Fig. 5.** Summary number concentration (top row), surface area (middle row), and volume (bottom row) distributions for 3 June to 15 July 2013. The left panels indicate the unperturbed state, the middle panels the dry-humidified state, and the right panels the dry state. The central black lines show the median distribution, and the shaded gray regions represent the interquartile range.
Fig. 6. Campaign-averaged diel profiles of total particle phase liquid water mass concentrations and RH binned into hourly intervals. The central lines indicate the median and the shaded regions indicate the interquartile range.
Fig. 7. Diel profile of volumetric growth factor, $gf_{\text{vol}}$, binned into hourly intervals. The central lines indicate the median and the shaded area represents interquartile range. Grey dotted lines indicating median growth factors that were calculated using the same RH and dry volume, and assumed $\kappa$ values ranging from 0 to 0.6.
Fig. 8. The top, middle, and bottom panels depict diurnal trends of smaller diameter mode, larger diameter mode, and volumetric $\kappa$, respectively. The trends are campaign-averaged and binned into hourly intervals. The central lines indicate the median values and the shaded area represents interquartile range.
Fig. 9. Diurnal trend of semi-volatile volumes lost during drying binned into hourly intervals. The central white line shows the mean of the measurements. The shaded blue area indicates a 95% confidence interval for the mean.
Fig. 10. Diurnal trend of select aerosol chemical species measured by PILS-IC and WSOM calculated from PILS-WSOC. The central black line shows the median of the measurements. The shaded area indicates the interquartile range. Panel (e) shows the ratio of $\sum(b) + (c) + (d)$ over $\sum(a) + (b) + (c) + (d)$. Panel (f) is identical to Fig. 8.
Fig. 11. Diurnal trend of dry aerosol volumes binned into hourly intervals. The central black line shows the median of the measurements. The shaded red area indicates the interquartile range.
Fig. A1. Same as Fig. 7 for 21 June 2013, a day with no rainfall. Grey dotted lines indicating median growth factors that were calculated using the same $RH$ and dry volume, and assumed $\kappa$ values ranging from 0 to 0.6. The black dotted line indicates the $RH$. 
Fig. A2. Comparison of particle-phase liquid water for a 5 day period with heavy rainfall (4–8 July, blue) versus a 5 day period with no rainfall (18–22 June, brown). The central lines indicate the median and the shaded regions indicate the interquartile range.